

# Gold nanoparticle-mediated photothermal therapy: applications and opportunities for multimodal cancer treatment

Rachel S. Riley<sup>1</sup> and Emily S. Day<sup>1,2\*</sup>

Photothermal therapy (PTT), in which nanoparticles embedded within tumors generate heat in response to exogenously applied laser light, has been well documented as an independent strategy for highly selective cancer treatment. Gold-based nanoparticles are the main mediators of PTT because they offer: (1) biocompatibility, (2) small diameters that enable tumor penetration upon systemic delivery, (3) simple gold-thiol bioconjugation chemistry for the attachment of desired molecules, (4) efficient light-to-heat conversion, and (5) the ability to be tuned to absorb near-infrared light, which penetrates tissue more deeply than other wavelengths of light. In addition to acting as a standalone therapy, gold nanoparticlemediated PTT has recently been evaluated in combination with other therapies, such as chemotherapy, gene regulation, and immunotherapy, for enhanced antitumor effects. When delivered independently, the therapeutic success of molecular agents is hindered by premature degradation, insufficient tumor delivery, and off-target toxicity. PTT can overcome these limitations by enhancing tumor- or cell-specific delivery of these agents or by sensitizing cancer cells to these additional therapies. All together, these benefits can enhance the therapeutic success of both PTT and the secondary treatment while lowering the required doses of the individual agents, leading to fewer off-target effects. Given the benefits of combining gold nanoparticle-mediated PTT with other treatment strategies, many exciting opportunities for multimodal cancer treatment are emerging that will ultimately lead to improved patient outcomes. © 2017 Wiley Periodicals, Inc.

How to cite this article: WIREs Nanomed Nanobiotechnol 2017, e1449. doi: 10.1002/wnan.1449

#### **INTRODUCTION**

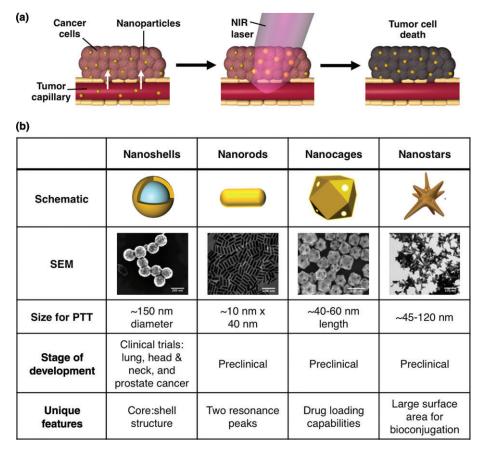
Nanoparticle-mediated PTT has been rapidly developing as a standalone therapy for cancer because it enables selective hyperthermia of tumor tissue while avoiding damage to healthy tissue. In PTT, plasmonic nanoparticles (NPs) are delivered into tumors and are irradiated with laser light, which causes the NPs' conduction band electrons to undergo synchronized oscillations that result in either the absorption or scattering of the applied light.<sup>1</sup> The absorbed light is converted into heat, which irreversibly damages the surrounding diseased tissue (Figure 1(a)). While NPs made of various materials can be employed for PTT,<sup>2–4</sup> gold-based nanoparticles (AuNPs), which we define here as those consisting either entirely or partially of gold (such as silica core/gold shell 'nanoshells'), have emerged as the lead therapeutic platform because they offer several major benefits. First, AuNPs enable simple gold-thiol bioconjugation chemistry for surface functionalization with therapeutic molecules, targeting ligands, or

<sup>\*</sup>Correspondence to: emilyday@udel.edu

<sup>&</sup>lt;sup>1</sup>Department of Biomedical Engineering, University of Delaware, Newark, DE, USA

<sup>&</sup>lt;sup>2</sup>Helen F. Graham Cancer Center & Research Institute, Newark, DE, USA

Conflict of interest: The authors have declared no conflicts of interest for this article.



**FIGURE 1** (a) In photothermal therapy, systemically-delivered nanoparticles accumulate within solid tumors *via* the enhanced permeability and retention effect, which exploits the leaky tumor vasculature. Once within the tumor, near-infrared light is applied to cause the nanoparticles to generate heat to kill the surrounding tumor tissue. (b) Four of the most commonly employed AuNPs for photothermal therapy include silica core/ gold shell nanoshells, gold nanorods, hollow gold nanocages, and nanostars. (Nanorod, nanocage, and nanostar images reprinted with permission from Ref 10. Copyright 2011 Wiley, Ref 11. Copyright 2007 Wiley-VCH Verlag GmbH & Co. KGaA, and Ref 12. Copyright 2015 Elsevier)

passivating agents that enhance biocompatibility as described in detail below. Additionally, the optical properties of AuNPs" can be tuned by controlling their structural dimensions so that they maximally absorb near-infrared (NIR) light ( $\lambda \approx 650-1350$  nm), which is ideal for PTT because it can safely penetrate deeply through healthy tissue to reach AuNPs embedded within tumors. A third benefit of AuNPs is that they can passively accumulate within tumors via the enhanced permeability and retention (EPR) effect, which exploits the inherently leaky and unorganized tumor vasculature.<sup>5</sup> Because of these benefits, AuNPmediated PTT has been thoroughly investigated for tumor ablation in animal models and has even entered clinical trials.<sup>1,6–9</sup> However, PTT as a standalone treatment is limited because it does not affect metastatic lesions or tumor cells outside of the area of irradiation, both of which may lead to disease recurrence. To overcome these limitations and enhance treatment outcome, researchers have recently begun to explore the use of AuNP-mediated PTT in combination with secondary therapeutic approaches that take advantage of the physiological and cellular changes within tumors afforded by PTT. Additionally, they have begun to utilize optical imaging techniques to guide and assess treatment in realtime. In this review, we first discuss the design features of AuNPs that are ideal for photothermal applications and then describe how PTT aids in the success of supplemental therapies. We also highlight the use of imaging to guide PTT. Finally, we discuss opportunities and challenges for the use of AuNPmediated PTT in multimodal strategies for cancer management.

## Design of Gold Nanoparticles for Photothermal Therapy

AuNPs used in photothermal applications must meet several design criteria such as having plasmon

resonance tunability, high photothermal conversion efficiency,<sup>13,14</sup> and simple surface functionalization or encapsulation chemistry. Based on these design criteria, nanoshells (NSs), nanorods (NRs), nanocages (NCs), and nanostars have emerged as the most common photothermal transducers. Here, we use these formulations as examples to discuss the desirable properties of AuNPs for PTT as depicted in Figure 1(b). This section is not meant to be a comprehensive overview, as others have discussed this topic elsewhere,<sup>1,15</sup> but we highlight key features that researchers should consider as they plan to use PTT in individual or multimodal therapeutic strategies.

First, AuNPs used in PTT must be designed to absorb light within the first (650-850 nm) or second (950-1350 nm) NIR window because these wavelengths of light can safely and deeply penetrate healthy tissue to reach AuNPs embedded within tumors. An important feature of AuNPs is that their structural dimensions can be tuned to yield maximal absorption within one of these two regions of light. The majority of AuNPs have been designed to maximally absorb within the first NIR window, which can safely penetrate 2-3 cm of tissue. For example, the peak plasmon resonance of NSs, which consist of spherical silica cores with thin gold shells, can be tuned by adjusting their core diameter-to-shell thickness ratio. NSs with 120 nm diameter cores and 15 nm thick shells are typically used for PTT because they maximally absorb 800 nm light.<sup>16,17</sup> Likewise, NRs have a short and long axis, thereby offering two absorption peaks corresponding to the transverse  $(\lambda \approx 500-550 \text{ nm})$  and longitudinal  $(\lambda \approx 650-850)$ nm) surface plasmon resonance (SPR).<sup>18,19</sup> The rod length can be shortened or elongated to achieve peak longitudinal SPR in the NIR region.<sup>10</sup> NRs on the order of 10 nm x x40 nm maximally absorb ~800 nm light and are therefore the most common for PTT. While most AuNPs have been designed to maximally absorb within the first NIR window, more recently novel AuNP designs have emerged for PTT in the second NIR window. This is because these longer wavelengths can safely penetrate up to 10 cm of tissue to reach deeply embedded tumors.<sup>20-22</sup> For example, Tchounwou et al. demonstrated the use of AuNP-coated single-walled carbon nanotubes as a multifunctional platform for imaging and PTT within the second NIR window.<sup>22</sup> Although we expect that PTT utilizing second window NIR light will gain attention in the coming years, here we focus on PTT within the first NIR window because it has been more commonly studied in combination PTT applications.

In addition to absorbing NIR light, AuNPs intended for use in PTT should also display high photothermal conversion efficiency, which is dictated by the AuNPs' structural dimensions (i.e., size and shape). The photothermal conversion efficiency of AuNPs of different sizes and shapes, and how this measurement can be determined, has been extensively studied in literature, and we point the reader towards these published works for more in-depth discussion.<sup>13,15,23,24</sup> In general, for spherical AuNPs, those with smaller diameters have higher conversion efficiency than those with larger diameters. Additionally, non-spherical AuNP designs, including NRs, nanostars, and NCs, are more efficient photothermal transducers than their spherical counterparts like NSs due to their larger absorption cross sections.<sup>15,19</sup>

Notably, the size and shape of AuNPs also influence their ability to extravasate from vasculature and penetrate solid tumors, which will ultimately impact the success of PTT because it will influence how evenly heat is distributed throughout the tumor.<sup>25,26</sup> For example, Perrault et al. compared the tumor penetrating abilities of 20 nm, 60 nm, and 100 nm solid gold spheres and found that larger size limits the distance NPs can travel away from blood vessels and into tumors.<sup>26</sup> However, tumor retention of smaller AuNPs is challenging because they are rapidly cleared from the extracellular milieu. Therefore, it is critical to determine the optimal size for penetration and retention based on the tumor characteristics (i.e., the 'leakiness' of its vasculature, the organization of its lymphatic vessels, and the density of the tumor extracellular matrix). This is particularly important given that the EPR effect is now known to be heterogeneous in distinct tumor types.<sup>27</sup> Intriguingly, PTT can be used to overcome these tumor penetration and retention limitations by amplifying tumor vessel leakiness and extracellular matrix permeability to enhance the delivery of more AuNPs or secondary therapeutic molecules, which we discuss later in this review.<sup>28,29</sup>

Another important consideration for AuNP design is their ability to encapsulate or be functionalized with biomolecules or drugs to enable combination therapy. The gold exterior of AuNPs is beneficial for bioconjugation because it enables simple gold-thiol bonding. One advantage of anisotropic materials like nanostars, which are spiked AuNPs, is that they offer a high surface-to-volume ratio for bioconjugation.<sup>30</sup> This can be advantageous because loading density can influence how AuNPs interact with cells and tissues. One disadvantage of loading molecules on the exterior of solid AuNPs, however, is that it leaves them susceptible to immune recognition or degradation. Hollow, porous AuNPs, such as NCs, offer the ability to load therapeutic agents in the nanoparticle core.<sup>11</sup> Since most hollow AuNPs destabilize upon NIR irradiation, the release of encapsulated molecules can be triggered 'on demand.'<sup>31</sup> Although this photothermal reshaping is desirable for drug delivery, it causes the AuNPs to lose their peak SPR to preclude their repeated use for PTT. Using NRs, researchers have demonstrated that one way to counteract this reshaping upon irradiation is by coating the NPs with a silica shell.<sup>32</sup> A similar strategy could be employed for other AuNPs that are known to reshape upon irradiation.

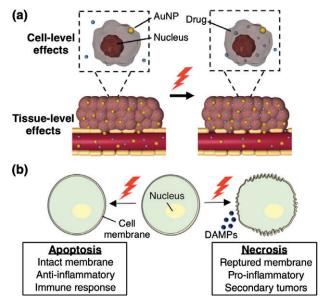
Here, we focused on how AuNPs' size, shape, and surface modifications can influence their heat generation and tumor penetration. In addition, these features also impact the AuNPs' biodistribution and biocompatibility, as demonstrated in various cell and animal models.<sup>12,33–35</sup> Similar to bulk gold, AuNPs are generally considered to be chemically inert and biocompatible.35 However, upon intravenous administration, AuNPs are rapidly coated with serum proteins that alter their biological identity and change how they are presented to cells.<sup>36–38</sup> Further, AuNPs are quickly recognized by the mononuclear phagocytic system, also known as the reticuloendothelial system (RES), leading to their rapid clearance from the blood. AuNPs' surface coating largely dictates the formation of the protein corona and clearance from the body. To reduce non-specific protein adsorption, extend circulation time, and enhance biocompatibility, AuNPs are often coated with passivating agents, such as poly(ethylene) glycol (PEG), in addition to targeting agents or other therapeutic molecules. The length and density of PEG on AuNPs is important, as it influences how well the AuNPs are shielded from protein corona formation and clearance from the body.<sup>37</sup> Among the various types of AuNPs used for PTT, NSs coated with PEG (also known as AuroShells, which are being commercialized by Nanospectra Biosciences, Inc.) are the furthest along in development and are currently the only AuNP for PTT being evaluated in clinical trials. These trials are examining the safety and efficacy of nanoshell-mediated PTT against lung, head and neck, and prostate cancers.<sup>7–9</sup> The first publication regarding these human trials indicates that PEG-coated NSs have an excellent clinical safety profile,39 supporting their continued use and providing evidence that PEG is a viable surface coating for AuNPs intended for human PTT. Although there are several clinical trials ongoing, efficacy data from these trials has not yet been published. In the Opportunities and Challenges section of this review, we further discuss ongoing clinical applications of AuNPs for PTT.

Overall, there are many features that contribute to whether a specific AuNP design will work well for PTT, and we have highlighted a few here. As previously mentioned, investigators have begun studying PTT in combination with other therapeutic strategies for overall enhanced therapeutic efficacy by exploiting the physiological and cellular level effects of PTT in tumors as described in the next section.

## Physiological Effects of PTT and the Mechanism of Cell Death Induced by PTT

AuNP-mediated PTT causes several physiological and biological changes to the surrounding tumor environment that can maximize the cytotoxic effects of PTT itself, enhance the efficacy of secondary therapies, or decrease required treatment dosages for secondary agents.<sup>40–42</sup> For example, the heat generated by AuNPs can increase the permeability of tumor vessels and cancer cell membranes to increase tumor and cellular uptake, respectively, of additional AuNPs or drugs (Figure 2(a)). We highlight some of the benefits of combination strategies that exploit these effects of PTT later in this review.

In addition to increasing tumor vessel and cell membrane permeability, PTT also causes intracellular



**FIGURE 2** Schematics demonstrating the physiological and biological effects of photothermal therapy. (a) The heat generated by AuNPs in response to NIR light enhances the permeability of tumor vasculature and cell membranes to increase the accumulation of secondary therapies, such as chemotherapeutic drugs. (b) PTT can lead to either cellular necrosis or apoptosis, which cause different cellular and immune responses. The mechanism of cell death depends on the applied irradiation parameters.

effects such as DNA damage and protein denaturation. Recently, researchers have begun to study the mechanisms of cell death triggered by PTT more closely because the mechanism of cell death (i.e., necrosis versus apoptosis) activates different cell signaling pathways that may influence treatment success and be beneficial for potential combination therapeutic strategies (Figure 2(b)). Conventional PTT uses high energy irradiation to cause rapid nanoparticle heating leading to cellular necrosis; while this strategy is effective for the ablation of established tumors,<sup>16,17,43</sup> it can also induce undesirable effects. For example, high energy PTT resulting in cellular necrosis can trigger the release of cellular waste and damage-associated molecular patterns (DAMPs) that induce inflammation, which may lead to increased secondary tumor growths.44,45 Conversely, low energy PTT initiates cellular apoptosis, which may lead to beneficial immunogenic responses.44,45 For example, cellular apoptosis can potentially discourage inflammation by causing phagocytes to produce anti-inflammatory molecules such as TGF-B.<sup>46</sup> Further, macrophages and dendritic cells (DCs) enter an anti-inflammatory state when in contact with apoptotic cells. The ability to induce apoptosis versus necrosis with PTT depends on many factors, including the AuNP photothermal properties, AuNP concentration in diseased tissue, and irradiation conditions. Given that initiating apoptosis versus necrosis with PTT can result in very distinct cellular effects, researchers should carefully control PTT parameters for their particular application. In addition, investigators should consider utilizing PTT in combination with secondary treatment strategies to capitalize on the tissue- or cellular- level effects induced by PTT. Below, we review how the physiological changes and mechanisms of cell death induced by PTT can be combined with chemotherapy, gene regulation, and immunotherapy for synergistic approaches to cancer treatment. We then describe how imaging can be used to guide and assess PTT in real-time.

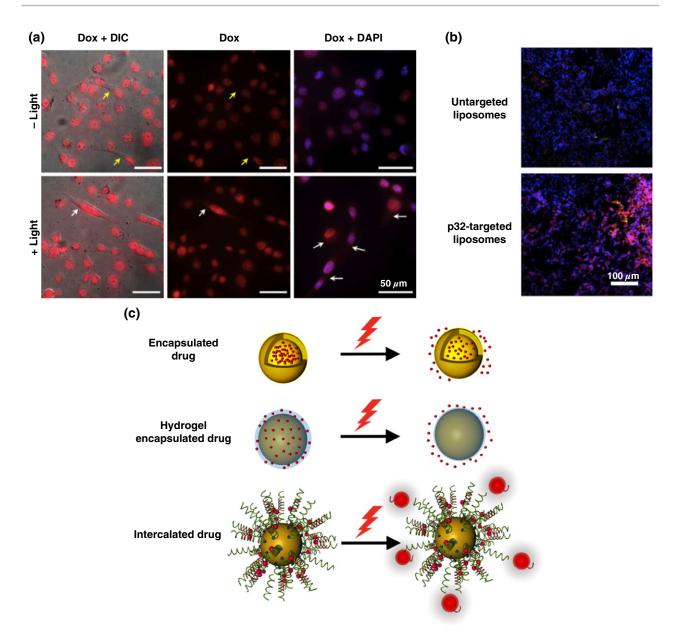
# ENHANCING CHEMOTHERAPY WITH GOLD NANOPARTICLE-MEDIATED PHOTOTHERMAL THERAPY

Although mortalities due to many cancers have not significantly improved in the past few decades, chemotherapy remains the current standard-of-care treatment. While cytotoxic at high doses, chemotherapy success is often hindered by acquired drug resistance and adverse side effects that limit the maximum

administered dose. As previously mentioned, hyperthermia increases both vascular and cell membrane permeability, as well as extracellular matrix permeability.<sup>28</sup> These physiological changes can amplify the effect of subsequently applied chemotherapy by increasing intratumoral and intracellular drug content.<sup>42,47</sup> Further, drug cytotoxicity may be enhanced by heat application.<sup>48</sup> One of the early indications of the synergy between hyperthermia and chemotherapy was demonstrated by Hahn et al., who showed optimal cytotoxicity by co-treating cells with doxorubicin and heat at 43°C.49 The clinical feasibility of widespread hyperthermia, however, is limited because it may increase drug cytotoxicity to healthy cells in addition to tumor cells. Comparatively, PTTmediated by AuNPs can provide localized, tumorspecific heating, thereby avoiding the detrimental effects of widespread hyperthermia.

In one investigation of combined chemotherapy and PTT, Hauck et al. demonstrated that NRmediated low intensity PTT initiates apoptosis in cancer cells, which in turn sensitizes the cells to cisplatin therapy and results in synergistic cytotoxicity.<sup>50</sup> We recently demonstrated that NS-mediated PTT of breast cancer cells leads to enhanced cell permeability and therefore increased intracellular drug content leading to decreased viability compared to cells that do not receive PTT (Figure 3(a)).<sup>42</sup> Notably, in addition to increasing the cellular uptake of drugs and the cellular response to drugs, hyperthermia can also be used to increase the amount of drugs that infiltrate tumors. This is because PTT can both increase tumors' vascular permeability and alter the expression of proteins found on cancer cell membranes, thereby providing a handle for targeting of additional therapeutic agents. In an elegant example of this technique, Park et al. fabricated a cooperative system that targeted drug-loaded liposomes to the p32 protein, which is upregulated following NRmediated PTT, to improve tumor delivery of the drugs and enhance treatment outcome versus PTT or chemotherapy individually (Figure 3(b)).<sup>29</sup> Strategies such as this that exploit the physical effects of PTT for enhanced treatment outcome have outstanding promise for future applications.

While the aforementioned strategies delivered AuNPs and chemotherapy separately, researchers have begun to create more complex AuNP-drug conjugates for combination PTT and chemotherapy. This should reduce off-target toxicity by limiting the amount of chemotherapy that reaches normal tissues and enabling drug release only upon irradiation. In these systems, drugs are physically encapsulated within AuNPs or bound to their surfaces using



**FIGURE 3** (a) Breast cancer cells treated with silica core/gold shell nanoshells and doxorubicin (red) and exposed to NIR light display increased drug uptake compared to cells that were not irradiated. Blue indicates nuclei, yellow arrows indicate cytoplasmic regions of cells not irradiated, and white arrows indicate enhanced doxorubicin content in the cytoplasm following irradiation. (Reprinted with permission from Ref 42. Copyright 2015 Dovepress) (b) Tumor tissue extracted from mice treated with nanorod-mediated hyperthermia and doxorubicin-loaded liposomes targeted to p32, a protein, that is upregulated in response to stress, shows enhanced liposome (green) and doxorubicin (red) distribution compared to mice treated with hyperthermia and untargeted liposomes. (Reprinted with permission from Ref 29. Copyright 2010 PNAS) (c) Schematics showing the mechanisms of light induced release of drugs from AuNPs.

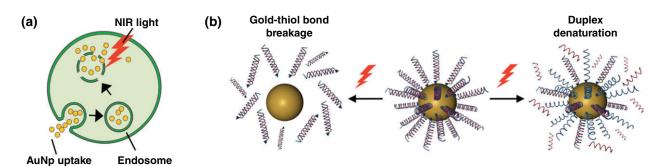
linkers that hold them in place until the heat produced during light irradiation triggers their release (Figure 3(c)). Hollow AuNPs, such as NCs, are ideal for drug encapsulation and light-induced release strategies.<sup>31,51,52</sup> For example, Wang et al. developed multifunctional NCs by encapsulating doxorubicin and coating the particles with hyaluronic acid (HA).<sup>52</sup> In this design, the HA acted as both a targeting agent to bind CD44 receptors and as a biodegradable capping agent to avoid premature drug release. Upon irradiation, the encapsulated doxorubicin was released only following degradation of the HA, which resulted in a drastic decrease in cell viability relative to controls.<sup>52</sup> Additional mechanisms of light-induced drug release include coating AuNPs with thermally responsive drug-containing hydrogels or oligonucleotides that contain intercalated drugs (Figure 3(c)). For instance, Strong et al. created NSs coated with a thin poly(N-isopropylacrylamide-co-acrylamide) hydrogel layer that remained swollen under physiological conditions, but expelled water and encapsulated drugs at higher temperatures achieved during NIR irradiation.<sup>53</sup> Similarly, thiolated ligands attached to AuNPs can be used to carry intercalating agents that are released upon photothermal irradiation (Figure 3(c)).<sup>54,55</sup> Overall, strategies for combination PTT and chemotherapy such as those reviewed here have great promise to increase treatment efficacy while minimizing off-target effects.

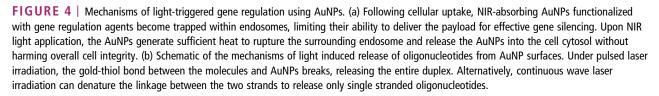
## THERMALLY RESPONSIVE GOLD NANOPARTICLES FOR GENE REGULATION

Gene regulation is a technique that aims to treat cancer by using nucleic acids to inhibit the expression of genes that drive tumor progression, and it is particularly promising for targets that are considered 'undruggable' by small molecules due to lack of an effective binding site. The main type of nucleic acid used for gene regulation is small interfering RNA (siRNA), which suppresses gene expression by triggering degradation of targeted messenger RNA molecules inside the cell cytoplasm, thereby halting production of the encoded disease-associated protein.<sup>56–58</sup> Unfortunately, several challenges have hindered the translation of gene regulation from the laboratory to the clinic. First, siRNAs and other nucleic acids are rapidly cleared from the bloodstream and quickly degraded in the presence of nucleases, making in vivo delivery difficult. Second, intracellular delivery of these negatively charged

molecules requires the use of cationic transfections agents that display undesirable toxicity. Further, many siRNA delivery strategies are not cell-specific, and therefore may lead to uptake and gene silencing in all cell types. AuNPs have been exploited as tools for gene regulation because they offer simple bioconjugation chemistry, enabling them to carry siRNAs or other nucleic acids for enhanced stability, intracellular delivery, and gene regulation both in vitro and in vivo.<sup>59-61</sup> However, a common challenge to achieving sufficient gene regulation with siRNA-AuNP conjugates is that they become trapped in intracellular compartments upon endocytosis.<sup>62</sup> To improve upon this issue and enable gene regulation only in desired tissues, PTT can be used to enable endosomal escape and trigger gene silencing on demand (Figure 4(a) and (b)).<sup>62-65</sup> A mechanistic study completed by Braun et al. demonstrated that hollow NSs conjugated with siRNA and Tat peptides for cellular uptake become trapped within endosomes following endocytosis. However, irradiating the samples with a pulsed NIR laser to induce NS heating caused the endosomes to rupture, enabling the release of the siRNA into the cytosol.<sup>62</sup> Similarly, Huschka et al. demonstrated that silica core/gold shell NSs coated with GFP-silencing DNA or siRNA drastically decreased expression GFP only following irradiation.66

There are two proposed mechanisms for lightinduced release of oligonucleotides from AuNP surfaces depending on the laser power and temperature reached during irradiation (Figure 4(b)).<sup>67</sup> First, a pulsed laser is used to break the gold-thiol bond between the oligonucleotides and the AuNPs, releasing the entire molecule from the AuNP surface.<sup>62</sup> The pitfall of this method is that the dissociation of the molecules from the NP surface results in NP





instability, and the free exposed thiols can be detrimental to live cells.<sup>67</sup> The second mechanism of release typically uses a continuous wave laser to denature the linkage between the sense and antisense strands of oligonucleotides with the goal of releasing only the unthiolated strand from the AuNP, although this may result in the gold-thiol bond breaking as well. This method requires longer irradiation times and is less efficient for release compared to the pulsed laser approach due to the lower laser powers used. However, researchers have optimized laser parameters to use the pulsed laser for duplex denaturation. For example, Poon et al. demonstrated that decreasing laser powers inhibited the gold-thiol bond breakage while still enabling duplex denaturation.<sup>67</sup> In addition to understanding the role that light irradiation/heating conditions play in facilitating the release of oligonucleotides from AuNPs, it is also important to consider how particle shape may influence the delivery of gene regulatory agents. Gold NRs, e.g., have been vastly studied for photorelease

following irradiation to trigger payload release.<sup>68–70</sup> Here, we have briefly reviewed the use of PTT to trigger the release of oligonucleotides from AuNP surfaces, as well as the use of PTT to release AuNPs and oligonucleotides from endosomes, as both of these strategies result in enhanced gene regulation. A more detailed discussion of this topic can be found in the recent review article published by Kim et al.<sup>71</sup> Looking forward, we encourage researchers to carefully study the signaling pathways that are activated in cells in response to PTT, and to investigate whether inhibiting these pathways via gene regulation can produce synergistic effects on tumor growth.

because they melt and reshape into spherical particles

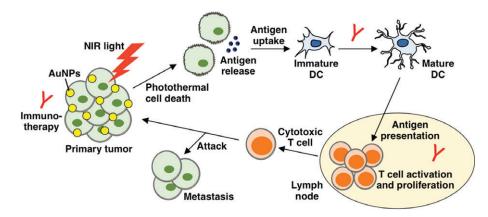
## COMBINING PHOTOTHERMAL THERAPY WITH IMMUNOTHERAPY FOR PROLONGED ANTI-CANCER EFFECTS AND METASTASIS INHIBITION

As previously discussed, PTT as a standalone therapy faces risk of recurrence and is not suitable for treating disseminated metastatic disease. Combining PTT with immunotherapy may prevent tumor recurrence and inhibit metastases.<sup>2,55,72–77</sup> Immunotherapy is a rapidly emerging, promising technique in which the host immune system is stimulated to recognize and kill cancer cells that have adapted various means of avoiding immune recognition.<sup>78</sup> One way cancer cells avoid recognition is by secreting cytokines such as IL-10 that suppress DC maturation (mature DCs

capture tumor antigens and present them to T cells for activation). Cancer cells also produce factors such as TRAIL and FasL that induce apoptosis of cytotoxic T cells (which find and destroy cancer cells), and down-regulate surface antigens and costimulatory molecules to decrease T-cell recognition and stimulation. Finally, cancer cells attract immune suppressive cells to the tumor microenvironment, including tumor-associated macrophages, myeloid derived suppressor cells, and T regulatory cells (Tregs, which suppress DCs and cytotoxic T cells by presenting inhibitory molecules such as cytotoxic lymphocyte antigen 4 (CTLA-4) and programmed cell death ligand 1 (PD-L1)). Immunotherapy counteracts these mechanisms by delivering tumor-specific T cells, immune checkpoint inhibitors (antibodies against CTLA-4 or PD-L1), or CpG oligodeoxynucleotides (which are recognized by toll-like receptor 9 (TLR9) on B cells and plasmacytoid DCs to stimulate an immune response). Each of these strategies has been evaluated in combination with PTT (mediated by both AuNPs and non-AuNPs), and the results indicate that dual PTT/immunotherapy is

advantageous to either therapy alone.<sup>2,55,72–77</sup> The rationale to combine PTT with immunotherapy is based on the fact that heat causes dying cancer cells to release antigens and heat shock proteins (HSPs) that are captured by antigen presenting cells such as DCs to mediate an immune response (Figure 5).<sup>72,73</sup> Further, the immunostimulatory environment created by PTT can enhance immunotherapies to prolong anti-cancer effects and eliminate both primary tumors and metastatic disease. For example, Bear et al. demonstrated in a murine melanoma model that PTT mediated by hollow gold NSs promotes the expression of pro-inflammatory cytokines and chemokines that induce DC maturation in tumor-draining lymph nodes and that prime antitumor CD8<sup>+</sup> effector T cells.<sup>74</sup> Combining PTT with adoptive T-cell transfer (ATCT) decreased tumor recurrence by 21.8% compared to PTT alone and also suppressed distant, untreated contralateral tumors more effectively than either therapy alone.<sup>74</sup> Thus, ATCT can help prevent primary tumor recurrence following PTT, and PTT can enhance the ability of ATCT to eradicate distant disease.

To our knowledge, PTT mediated by AuNPs has not yet been evaluated in combination with immune checkpoint inhibitors, but a recent study by Wang et al. demonstrated that PTT mediated by single-walled carbon nanotubes (SWNTs) is synergistic with anti-CTLA-4 therapy.<sup>75</sup> They observed that SWNT-mediated PTT increased DC maturation in tumor-draining lymph nodes, and the combination of



**FIGURE 5** Schematic depicting the impact of combined photothermal therapy and immunotherapy on primary tumors and metastases. (Reprinted with minor modifications with permission from Ref 72. Copyright 2016 World Scientific Publishing Company)

PTT and anti-CTLA-4 therapy suppressed distant metastasis (either contralateral flank tumors or lung metastasis) in a murine model of breast cancer better than either therapy alone.<sup>75</sup> Given the success of this study, it is likely that AuNP-mediated PTT would also result in impressive anti-tumor effects when combined with immune checkpoint inhibitors. Finally, CpG-mediated immunotherapy has also been tested in combination with PTT and appears promising.<sup>2,55</sup> Tao et al., for instance, showed that NRs coated with CpGs that were intercalated with doxorubicin could enable in vivo combination chemotherapy, light-triggered drug release, hyperthermia, and immunotherapy.55 This is an excellent example of how AuNPs can be exploited for multimodal anticancer therapy.

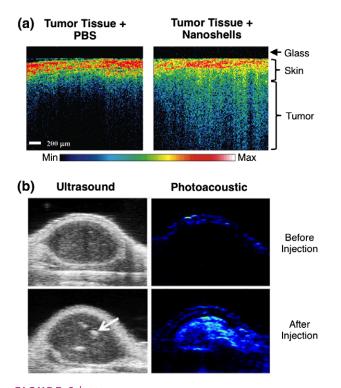
While most research combining PTT with immunotherapy has used PTT to enhance the effect of immunotherapy, researchers have also begun to exploit immune cells to enhance PTT. Specifically, since immune cells such as antigen-specific cytotoxic T cells, cytokine-induced killer cells, and macrophages have a unique ability to 'home' to the tumor microenvironment in vivo, researchers have attached AuNPs such as NRs, nanoprisms, and NSs,<sup>79-82</sup> as well as non-AuNPs such as Prussian blue NPs,<sup>77</sup> to the surface of these cells or loaded them within the cells' interior to enhance tumor delivery and improve the efficacy of PTT. This technique, which has been referred to as 'cellular backpacking' or 'Trojan horse' delivery, capitalizes on the natural homing ability of immune cells for enhanced treatment outcomes and shows great promise for further development.

Overall, there is great potential for combining AuNP-mediated PTT with immunotherapy, as it can enable treatment of both primary tumors and distant metastases and may also prevent long-term tumor recurrence. Given that PTT and immunotherapy are mutually beneficial, we anticipate that the field of photothermal immunotherapy will rapidly expand in the next decade to result in improved patient outcomes.

## IMAGE-GUIDANCE AND ASSESSMENT OF PTT USING GOLD NANOPARTICLES

A key advantage of AuNPs is that their unique photonic properties enable them to be used as optical contrast agents, so imaging can be used to guide and assess PTT in real-time.<sup>83–85</sup> Two imaging modalities that have been extensively studied for this purpose in pre-clinical settings are optical coherence tomography (OCT) and photoacoustic (PA) imaging, which exploit the AuNPs' light scattering or absorption properties, respectively. Here, we describe each of these in detail. We also describe the use of magnetic resonance thermal imaging for image-guided PTT.

OCT uses a low coherence light source applied through the depth of a sample and detects the resultant reflection at micrometer-level resolution.<sup>86,87</sup> Since AuNPs provide reflective signal beyond the native tissue itself, tumors or other tissues containing AuNPs display enhanced OCT signal compared to tissue not harvesting AuNPs (Figure 6(a)).<sup>87</sup> Using NSs as contrast agents and photothermal transducers, Gobin et al. showed that OCT could be used to ensure sufficient NS accumulation within tumors prior to PTT. More specifically, they showed that systemically-delivered NSs enhanced the OCT signal in tumors but not in healthy tissue in a murine cancer model, and subsequent application of NIR light led to a reduction in tumor size.87 In this manner, researchers can utilize OCT for image-guided therapy



**FIGURE 6** (a) Silica core/gold shell nanoshells accumulated within tumor tissue can be visualized with optical coherence tomography, as depicted by increased signal throughout the tissue depth. (b) *In vivo* ultrasonic and photoacoustic images demonstrate enhanced signal from biodegradable gold-based NPs accumulated within the tumor. NPs were composed of clusters of small gold nanoparticles that produced an ultrastrong plasmonic coupling effect for imaging and PTT. The arrow indicates NPs. ((a) Reprinted with permission from Ref 87. Copyright 2007 American Chemical Society and (b) Ref 88. Copyright 2013 Wiley-VCH Verlag GmbH & Co. KGaA)

to ensure ablation of the entire tumor. More recently, researchers have demonstrated that the absorptive properties of photothermally active NPs can be exploited to enhance contrast of phase-sensitive OCT (PSOCT) ex vivo. While conventional OCT generates contrast based on light scattering, PSOCT generates contrast based on small alterations in the sample's optical path length, which is influenced by tissue temperature.<sup>87</sup> Since AuNPs generate heat upon light irradiation, they can enhance PSOCT contrast in a technique known as photothermal OCT.<sup>89</sup> Photothermal OCT has recently been applied in vivo, and has proven useful for analysis of highly-scattering tissues because it can separate the AuNPs from the scattering background.<sup>89</sup> Overall, OCT has shown much promise as a tool to guide PTT in ex vivo and preclinical in vivo studies, but its low penetration depth may limit its clinical utility.

An additional technique used for image guidance and assessment of PTT is photoacoustic (PA) imaging (Figure 6(b)).<sup>32,88</sup> In PA imaging, tissue is irradiated with a pulsed laser and light absorption produces a temperature rise in the tissue leading to its subsequent thermo-elastic expansion. The pressure induced by this expansion leads to propagation of acoustic waves, which are detected by an ultrasound transducer at the surface of the body. AuNPs embedded in tissue provide increased PA signal because they convert light to heat more efficiently than native tissue itself. Therefore, PA imaging can be utilized to guide and assess PTT efficacy.<sup>90</sup> Notably, since PA imaging incorporates an ultrasound transducer, this technology readily allows for acquisition of traditional ultrasound images. Overlaying PA images with ultrasound images can provide morphological information and reveal the distribution of AuNPs in tumor tissue. Further, as shown in Figure 6 (b), AuNPs can inherently increase contrast of both ultrasound and PA imaging.<sup>88</sup> Another benefit of PA imaging is that it can also be used to remotely monitor the thermal dose applied to the tissue during PTT. This was demonstrated by Kim et al., who showed that PA imaging could be used to determine the distribution of NRs within subcutaneous human epithelial tumors in mice and to map the temperature distribution during irradiation.<sup>32</sup> As previously mentioned, the temperature achieved during PTT will influence whether the cells undergo apoptosis or necrosis, which ultimately influences therapeutic success. Using techniques such as PA imaging to monitor the temperatures reached during PTT may allow researchers to optimize irradiation parameters during treatment to control the mechanism of cell death. Finally, a very exciting application of PA imaging is the analysis of micrometastases in lymph nodes.<sup>91,92</sup> Since cancer typically spreads first to the lymph nodes prior to distant organs, the ability to detect this earliest stage of metastasis and provide PTT in a single setting holds much promise as an intervention strategy.

In addition to OCT and PA imaging, magnetic resonance thermal imaging (MRTI) has also been studied as a tool to non-invasively guide or assess AuNP-mediated PTT using pre-clinical *in vivo* models of human cancer.<sup>93–95</sup> In MRTI, a temperature map is generated in real-time during irradiation that can be interpreted by physicians to control the thermal doses applied.<sup>93,94</sup> In one example of this technique, Rylander et al. demonstrated the use of MRTI in prostate tumors and correlated the temperatures reached during irradiation with expression of heat shock proteins.<sup>95</sup> Since MRTI temperature mapping can be used to control and optimize PTT parameters in real time, it may be possible to use this technology

during PTT to control the mechanism of cell death and also to minimize any adverse impacts to adjacent healthy tissue. It should be noted that this technology is best applied to static tissues, as motion can easily corrupt the accuracy of MRTI.<sup>96</sup>

In this section, we have introduced three imaging techniques that are the most commonly studied for guidance and assessment of PTT. Confirming that AuNPs have adequately accumulated in tumors prior to irradiation, and that the desired temperature has been achieved upon irradiation, is critical to ensure success of PTT as a standalone therapy or in combination with chemotherapy, gene regulation, or immunotherapy. While the ability to guide PTT in humans with OCT, PA imaging, or MRTI has not yet been evaluated, we anticipate that this will occur in the near future as PTT progresses through clinical development. Indeed, MRTI is already used for temperature mapping during cryotherapy, radiofrequency ablation, and focused ultrasound ablation, so its implementation for clinical guidance of PTT should be straightforward.<sup>97</sup> For a discussion of the clinical status and potential of each of these imaging modalities, we point the reader towards several recent papers that provide an excellent summary of the benefits and limitations of each of these imaging modalities in the clinical setting.98-101

## ADDITIONAL APPLICATIONS OF PHOTOTHERMALLY ACTIVE GOLD NANOPARTICLES

While this review has emphasized the use of PTT in cancer therapy, PTT also poses opportunities for other biological applications outside cancer management. For example, since AuNPs can passively accumulate in atherosclerotic plaques, they have been used for imageguided PTT of atherosclerosis directed by intravascular photoacoustic imaging (IVPA).<sup>102,103</sup> Additionally, PTT has been studied for the targeted ablation of bacterial cells to treat antibiotic-resistant infections such as methicillin-resistant Staphylococcus aureus (MRSA).<sup>104,105</sup> Moreover, the heat-generating photothermal effect has also been investigated for global health and environmental applications. Neumann et al. demonstrated that specialized AuNPs designed to absorb light across the entire solar spectrum could produce sufficient heat in response to sunlight to generate steam for sterilization and sanitization purposes.<sup>106,107</sup> These applications of photothermally active AuNPs are particularly interesting and important for developing countries that

do not currently have access to clean energy and water supplies.

# **OPPORTUNITIES AND CHALLENGES**

Excitingly, PTT has rapidly progressed from concept to clinical application, with PTT-mediated by NSs already in three distinct Phase 1 clinical trials (Figure 1(b))that focus on the treatment of lung, head and neck, and prostate cancers.7-9 While efficacy data has not yet been published, clinical trials evaluating safety of NSs are promising.<sup>39</sup> In addition, PTT mediated by both NSs and NRs has begun to be applied to veterinarian medicine, with a special emphasis on treating canine and feline tumors.<sup>108,109</sup> Although we do not focus on translatability here, a comprehensive overview of clinical trials that utilize AuNPs for photothermal applications has recently been published by Pedrosa et al.<sup>110</sup> We anticipate that the next decade will witness the commencement of additional clinical trials of other more recently developed AuNPs, the expansion into applications beyond cancer, and the evaluation of multimodal strategies.

There are several critical opportunities and challenges that should be addressed as PTT continues to develop as a standalone therapy and as a component of multifunctional strategies for cancer management. Detailed studies should be performed to understand how adjusting irradiation conditions, AuNP types, and the temperatures achieved during PTT influence the underlying impact to cells, including the mechanism of cell death, as this is crucial for successful therapeutic outcome. Conventional PTT, while effective for tumor ablation with high laser energies and temperatures, is challenging as a standalone therapy because it may result in cellular necrosis and therefore increased inflammation and secondary tumor growth. Further, it is difficult to ensure complete ablation of all tumor cells, and PTT is not feasible as a treatment for metastatic disease. However, there are increasing opportunities to use PTT in combination with other therapies such as chemotherapy, gene regulation, and immunotherapy, to provide synergy for more thorough cancer treatment. The combination of PTT and these secondary techniques is mutually beneficial. That is, PTT can enhance secondary therapies by increasing molecular stability and tumor delivery, and these other therapies can enhance PTT by ensuring complete elimination of cells. Finally, PTT can be combined with imaging strategies to guide therapeutic application and monitor treatment success. Overall, using PTT in combination with imaging and/or second therapeutic strategies offers great potential for increasing tumor regression while decreasing offtarget effects to healthy tissue.

#### CONCLUSION

AuNP-mediated PTT has been thoroughly investigated as a standalone therapy for cancer, but its recent use in combination with other imaging and treatment modalities presents improved therapeutic results. AuNPs are ideal photothermal transducers because their optical properties can be tuned by adjusting their structural dimensions so that they strongly absorb tissue-penetrating NIR light. Further, AuNPs provide several essential features for use in multimodal therapies including simple goldthiol bioconjugation chemistry and efficient tumor accumulation and cellular uptake. PTT can exploit these qualities of AuNPs to provide controlled release of conjugated or encapsulated molecules

specifically at diseased sites. Additionally, AuNPmediated PTT can sensitize cancer cells to chemotherapy, gene regulation, and immunotherapy by increasing cell permeability and intracellular delivery. Controlled release and hyperthermia-induced sensitization can lower the required therapy dosages and increase localized efficacy. Lastly, PTT can modulate cellular apoptosis or necrosis depending on the laser power and resultant temperature reached during irradiation. The mechanism of cell death is important because necrotic cells can promote secondary tumor growth, while apoptotic cells may elicit an immune response to decrease the formation of secondary tumors. Therefore, it is critical to understand the mechanism of cell death to optimally design multimodal cancer therapies. Owing to the extensive synergy between hyperthermia and other treatment modalities, it is likely that AuNPmediated PTT will become a hallmark of multifunctional cancer therapies.

#### ACKNOWLEDGMENTS

This work was supported by the University of Delaware Research Foundation, by grant number IRG14-251-07-IRG from the American Cancer Society, by a Science and Engineering Research Grant from the W.M. Keck Foundation, and by the National Institutes of General Medical Sciences (NIGMS) of the National Institutes of Health (NIH) under award number R35GM119659 and through an Institutional Development Award (IDeA) from NIH NIGMS under grant number U54GM104941. The Delaware INBRE program, with a grant from NIGMS (P20-GM103446) from the NIH, also supported this project. The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH.

#### REFERENCES

- 1. Kennedy LC, Bickford LR, Lewinski NA, Coughlin AJ, Hu Y, Day ES, West JL, Drezek RA. A new era for cancer treatment: gold-nanoparticlemediated thermal therapies. *Small* 2011, 7:169–183.
- 2. Guo L, Yan D, Yang D, Li Y, Wang X, Zalewski O. Combinatorial photothermal and immuno cancer therapy using chitosan-coated hollow copper sulfide nanoparticles. *ACS Nano* 2015, 8:5670–5681.
- 3. Hashida Y, Tanaka H, Zhou S, Kawakami S, Yamashita F, Murakami T, Umeyama T, Imahori H, Hashida M. Photothermal ablation of tumor cells using a single-walled carbon nanotube-peptide composite. *J Control Release* 2014, 173:58–66.
- 4. Li M, Teh C, Ang CY, Tan SY, Luo Z, Qu Q, Zhang Y, Korzh V, Zhao Y. Near-infrared lightabsorptive stealth liposomes for localized photothermal ablation of tumors combined with chemotherapy. *Adv Funct Mater* 2015, 25:5602–5610.
- 5. Maeda H, Wu J, Sawa T, Matsumura Y, Hori K. Tumor vascular permeability and the EPR effect in

macromolecular therapeutics: a review. J Control Release 2000, 65:271–284.

- 6. Blaney SM. Vascular-targeted photothermal therapy of an orthotopic murine glioma model; 2011.
- Nanospectra Biosciences Inc. Pilot study of aurolase(tm) therapy in refractory and/or recurrent tumors of the head and neck. Available at: https:// clinicaltrials.gov/ct2/show/NCT00848042?term= auroshell&rank=2. (Accessed May 23, 2016).
- Nanospectra Biosciences Inc. Efficacy study of aurolase therapy in subjects with primary and/or metastatic lung tumors. Available at: https://clinicaltrials. gov/ct2/show/NCT01679470?term=auroshell&rank= 3. (Accessed May 23, 2016).
- 9. Nanospectra Biosciences Inc. MRI/US fusion imaging and biopsy in combination with nanoparticle directed focal therapy for ablation of prostate tissue. Available at: https://clinicaltrials.gov/ct2/show/NCT02680535? term=auroshell&rank=1. (Accessed September 28, 2016).

- Stone J, Jackson S, Wright D. Biological applications of gold nanorods. WIREs Nanomed Nanobiotechnol 2011, 3:100–109.
- 11. Skrabalak SE, Chen J, Au L, Lu X, Li X, Xia Y. Gold nanocages for biomedical applications. *Adv Mater* 2007, 19:3177–3184.
- Dam DHM, Culver KSB, Kandela I, Lee RC, Chandra K, Lee H, Mantis C, Ugolkov A, Mazar AP, Odom TW. Biodistribution and in vivo toxicity of aptamer-loaded gold nanostars. *Nanomed Nanotechnol Biol Med* 2015, 11:671–679.
- 13. Cole JR, Mirin NA, Knight MW, Goodrich GP, Halas NJ. Photothermal efficiencies of nanoshells and nanorods for clinical therapeutic applications. *J Phys Chem C* 2009, 113:12090–12094.
- 14. Wang Y, Black K, Leuhmann H, Li W, Zhang Y, Cai X, Wan D, Liu S-Y, Li M, Kim P, et al. A comparison study of gold nanohexapods, nanorods, and nanocages for photothermal cancer treatment. *ACS Nano* 2013, 7:2068–2077.
- 15. Abadeer NS, Murphy CJ. Recent progress in cancer thermal therapy using gold nanoparticles. J Phys Chem C 2016, 120:4691–4716.
- Lowery AR, Gobin AM, Day ES, Halas NJ, West JL. Immunonanoshells for targeted photothermal ablation of tumor cells. *Int J Nanomedicine* 2006, 1:149–154.
- 17. Day ES, Thompson PA, Zhang L, Lewinski NA, Ahmed N, Drezek RA, Blaney SM, West JL. Nano-shell-mediated photothermal therapy improves survival in a murine glioma model. *J Neurooncol* 2011, 104:55–63.
- 18. Huang X, El-Sayed IH, Qian W, El-Sayed MA. Cancer cell imaging and photothermal therapy in the near-infrared region by using gold nanorods. *J Am Chem Soc* 2006, 128:2115–2120.
- 19. Mackey MA, Ali MRK, Austin LA, Near RD, El-sayed MA. The most effective gold nanorod size for plasmonic photothermal therapy: theory and in vitro experiments. *JPhys Chem B*2014,118:1319–1326.
- 20. Wang S, Xu H, Ye J. Plasmonic rod-in-shell nanoparticles for photothermal therapy. *Phys Chem Chem Phys* 2014, 16:12275–12281.
- 21. Ding X, Liow CH, Zhang M, Huang R, Li C, Shen H, Liu M, Zou Y, Gao N, Zhang Z, et al. Surface plasmon resonance enhanced light absorption and photothermal therapy in the second near-infrared window. *J Am Chem Soc* 2014, 136:15684–15693.
- 22. Tchounwou C, Sinha SS, Viraka Nellore BP, Pramanik A, Kanchanapally R, Jones S, Chavva SR, Ray PC. Hybrid theranostic platform for second near-IR window light triggered selective two-photon imaging and photothermal killing of targeted melanoma cells. ACS Appl Mater Interfaces 2015, 7:20649–20656.

- 23. Jiang K, Smith DA, Pinchuk A. Size-dependent photothermal conversion efficiencies of plasmonically heated gold nanoparticles. *J Phys Chem C* 2013, 117:27073–27080.
- 24. Qin Z, Wang Y, Randrianalisoa J, Raeesi V, Chan WCW, Lipiński W, Bischof JC. Quantitative comparison of photothermal heat generation between gold nanospheres and nanorods. *Sci. Rep.* 2016, 6:29836; doi: 10.1038/srep29836.
- 25. Huang K, Ma H, Liu J, Huo S, Kumar A, Wei T, Zhang X, Jin S, Gan Y, Wang PC, et al. Sizedependent localization and penetration of ultrasmall gold nanoparticles in cancer cells, multicellular spheroids, and tumors in vivo. *ACS Nano* 2012, 6:4483–4493.
- Perrault SD, Walkey C, Jennings T, Fischer HC, Chan WCW. Mediating tumor targeting efficiency of nanoparticles through design. *Nano Lett* 2009, 9:1909–1915.
- 27. Prabhakar U, Maeda HK, Jain R, Sevick-Muraca EM, Zamboni W, Farokhzad OC, Barry ST, Gabizon A, Grodzinski P, Blakey DC. Challenges and key considerations of the enhanced permeability and retention effect for nanomedicine drug delivery in oncology. *Cancer Res* 2013, 73:2412–2417.
- 28. Raeesi V, Chan WCW. Improving nanoparticle diffusion through tumor collagen matrix by photo-thermal gold nanorods. *Nanoscale* 2016, 8:12524–12530.
- 29. Park J-H, von Maltzahn G, Xu MJ, Fogal V, Kotamraju VR, Ruoslahti E, Bhatia SN, Sailor MJ. Cooperative nanomaterial system to sensitize, target, and treat tumors. *Proc Natl Acad Sci U S A* 2010, 107:981–986.
- 30. Chen H, Zhang X, Dai S, Ma Y, Cui S, Achilefu S, Gu Y. Multifunctional gold nanostar conjugates for tumor imaging and combined photothermal and chemo-therapy. *Theranostics* 2013, 3:633–649.
- 31. Yavuz MS, Cheng Y, Chen J, Cobley CM, Zhang Q, Rycenga M, Xie J, Kim C, Song KH, Schwartz AG, et al. Gold nanocages covered by smart polymers for controlled release with near-infrared light. *Nat Mater* 2009, 8:935–939.
- 32. Kim S, Chen YS, Luke G, Emelianov S. In-vivo ultrasound and photoacoustic image-guided photothermal cancer therapy using silica-coated gold nanorods. *IEEE Trans Ultrason Ferroelectr Freq Control* 2014, 61:891–897.
- 33. Rotello VM. The role of surface functionality in determining nanoparticle cytotoxicity. *Acc Chem Res* 2013, 46:681–691.
- Tagami T, Taki M, Ozeki T. Nanomaterials in pharmacology. *Methods Pharmacol Toxicol* 2016, 39:333–347.
- 35. Alkilany AM, Murphy CJ. Toxicity and cellular uptake of gold nanoparticles: what we have learned so far? *J Nanopart Res* 2010, 12:2313–2333.

- Monopoli MP, Aberg C, Salvati A, Dawson KA. Biomolecular coronas provide the biological identity of nanosized materials. *Nat Nanotechnol* 2012, 7:779–786.
- 37. Dai Q, Walkey C, Chan WCW. Polyethylene glycol backfilling mitigates the negative impact of the protein corona on nanoparticle cell targeting. *Angew Chem Int Ed Engl* 2014, 53:5093–5096.
- Walkey CD, Chan WCW. Understanding and controlling the interaction of nanomaterials with proteins in a physiological environment. *Chem Soc Rev* 2012, 41:2780–2799.
- 39. Stern JM, Solomonov VVK, Sazykina E, Schwartz JA, Gad SC, Goodrich GP. Initial evaluation of the safety of nanoshell-directed photothermal therapy in the treatment of prostate disease. *Int J Toxicol* 2015, 35:38–46.
- 40. Gormley AJ, Larson N, Banisadr A, Robinson R, Frazier N, Ray A, Ghandehari H. Plasmonic photo-thermal therapy increases the tumor mass penetration of HPMA copolymers. *J Control Release* 2013, 166:130–138.
- 41. Gormley AJ, Greish K, Ray A, Robinson R, Gustafson JA, Ghandehari H. Gold nanorod mediated plasmonic photothermal therapy: a tool to enhance macromolecular delivery. *Int J Pharm* 2011, 415:315–318.
- 42. Fay BL, Melamed JR, Day ES. Nanoshell-mediated photothermal therapy can enhance chemotherapy in inflammatory breast cancer cells. *Int J Nanomedicine* 2015, 10:6931–6941.
- 43. Bernardi RJ, Lowery AR, Thompson PA, Blaney SM, West JL. Immunonanoshells for targeted photothermal ablation in medulloblastoma and glioma: an in vitro evaluation using human cell lines. *J Neurooncol* 2008, 86:165–172.
- 44. Melamed JR, Edelstein RS, Day ES. Elucidating the fundamental mechanisms of cell death triggered by photothermal therapy. *ACS Nano* 2015, 9:6–11.
- 45. Pérez-Hernández M, Del Pino P, Mitchell SG, Moros M, Stepien G, Pelaz B, Parak WJ, Gálvez EM, Pardo J, De La Fuente JM. Dissecting the molecular mechanism of apoptosis during photothermal therapy using gold nanoprisms. ACS Nano 2015, 9:52–61.
- 46. Fadok VA, Bratton DL, Konowal A, Freed PW, Westcott JY, Henson PM. Macrophages that have ingested apoptotic cells in vitro inhibit proinflammatory cytokine production through autocrine/paracrine mechanisms involving TGF-beta, PGE2, and PAF. *J Clin Invest* 1998, 101:890–898.
- 47. Issels RD. Hyperthermia adds to chemotherapy. *Eur J Cancer* 2008, 44:2546–2554.
- Park H, Yang J, Lee J, Haam S, Choi IH, Yoo KH. Multifunctional nanoparticles for combined doxorubicin and photothermal treatments. ACS Nano 2009, 3:2919–2926.

- 49. Hahn GM, Braun J, Har-Kedar I. Thermochemotherapy: synergism between hyperthermia (42-43 degrees) and adriamycin (or bleomycin) in mammalian cell inactivation. *Proc Natl Acad Sci U S A* 1975, 72:937–940.
- Hauck TS, Jennings TL, Yatsenko T, Kumaradas JC, Chan WCW. Enhancing the toxicity of cancer chemotherapeutics with gold nanorod hyperthermia. *Adv Mater* 2008, 20:3832–3838.
- 51. Park J, Park J, Ju EJ, Park SS, Choi J, Lee JH, Lee KJ, Shin SH, Ko EJ, Park I, et al. Multifunctional hollow gold nanoparticles designed for triple combination therapy and CT imaging. *J Control Release* 2015, 207:77–85.
- 52. Wang Z, Chen Z, Liu Z, Shi P, Dong K, Ju E, Ren J, Qu X. A multi-stimuli responsive gold nanocagehyaluronic platform for targeted photothermal and chemotherapy. *Biomaterials* 2014, 35:9678–9688.
- 53. Strong LE, West JL. Hydrogel-coated near infrared absorbing nanoshells as light-responsive drug delivery vehicles. ACS Biomater Sci Eng 2015, 1:685–692.
- 54. Brann T, Patel D, Chauhan R, James KT, Bates PJ, Malik MT, Keynton RS, Toole MGO. Gold nanoplates as cancer-targeted photothermal actuators for drug delivery and triggered release. J Nanomater 2016, 2016:11 pages. doi:10.1155/2016/2036029.
- 55. Tao Y, Ju E, Liu Z, Dong K, Ren J, Qu X. Engineered, self-assembled near-infrared photothermal agents for combined tumor immunotherapy and chemo-photothermal therapy. *Biomaterials* 2014, 35:6646–6656.
- 56. Eloy JO, Petrilli R, Lopez RFV, Lee RJ. Stimuliresponsive nanoparticles for siRNA delivery. *Curr Pharm Des* 2015, 21:4131–4144.
- 57. Hannon GJ. RNA interference. *Nature* 2002, 418:244–251.
- Kim HJ, Kim A, Miyata K, Kataoka K. Recent progress in development of siRNA delivery vehicles for cancer therapy. *Adv Drug Deliv Rev* 2016, 104:61–77.
- 59. Jensen SA, Day ES, Ko CH, Hurley LA, Luciano JP, Kouri FM, Merkel TJ, Luthi AJ, Patel PC, Cutler JI, et al. Spherical nucleic acid nanoparticle conjugates as an RNAi-based therapy for glioblastoma. *Sci Transl Med* 2013, 5:209ra152.
- Giljohann DA, Seferos DS, Prigodich AE, Patel PC, Mirkin CA. Gene regulation with polyvalent siRNA—nanoparticle conjugates. *JACS Commun* 2009, 131:2072–2073.
- 61. Kouri FM, Hurley LA, Daniel WL, Day ES, Hua Y, Hao L, Peng CY, Merkel TJ, Queisser MA, Ritner C, et al. MiR-182 integrates apoptosis, growth, and differentiation programs in glioblastoma. *Genes Dev* 2015, 29:732–745.

- 62. Braun GB, Pallaoro A, Wu G, Missirlis D, Zasadzinski JA, Tirrell M, Reich NO. Laser-activated gene silencing via gold nanoshell-siRNA conjugates. *ACS Nano* 2009, 3:2007–2015.
- 63. Barhoumi A, Huschka R, Bardhan R, Knight MW, Halas NJ. Light-induced release of DNA from plasmon-resonant nanoparticles: towards lightcontrolled gene therapy. *Chem Phys Lett* 2009, 482:171–179.
- 64. Jones MR, Millstone JE, Giljohann DA, Seferos DS, Young KL, Mirkin CA. Plasmonically controlled nucleic acid dehybridization with gold nanoprisms. *Chemphyschem* 2009, 10:1461–1465.
- 65. Lu W, Zhang G, Zhang R, Ii LGF, Huang Q, Gelovani JG, Li C. Tumor site-specific silencing of NF-κB p65 by targeted hollow gold nanospheremediated photothermal transfection. *Cancer Res* 2010, 70:3177–3189.
- 66. Huschka R, Barhoumi A, Liu Q, Roth J a, Ji L, Halas NJ. Gene silencing by gold nanoshell-mediated delivery and laser-triggered release of antisense oligonucleotide and siRNA. ACS Nano 2012, 6:7681–7691.
- 67. Poon L, Zandberg W, Hsiao D, Erno Z, Sen D, Gates BD, Branda NR. Photothermal release of single-stranded DNA from the surface of gold nanoparticles through controlled denaturating and AU-S bond breaking. ACS Nano 2010, 4:6395–6403.
- 68. Lee SE, Liu GL, Kim F, Lee LP. Remote optical switch for localized and selective control of gene interference. *Nano Lett* 2009, 9:562–570.
- 69. Takahashi H, Niidome Y, Yamada S. Controlled release of plasmid DNA from gold nanorods induced by pulsed near-infrared light. *ChemComm* 2005, 17:2247–2249. doi 10.1039/b500337g
- Wijaya A, Schaffer SB, Pallares IG, Hamad-Schifferli K. Selective release of multiple DNA oligonucleotides from gold nanorods. ACS Nano 2009, 3:80–86.
- Kim J, Kim J, Jeong C, Kim WJ. Synergistic nanomedicine by combined gene and photothermal therapy. *Adv Drug Deliv Rev* 2016, 98:99–112.
- 72. Zhou F, Nordquist RE, Chen WR. Photonics immunotherapy — a novel strategy for cancer treatment. *J Innov Opt Health Sci* 2016, 9:1–11.
- 73. Almeida JPM, Figueroa ER, Drezek RA. Gold nanoparticle mediated cancer immunotherapy. *Nanomed NBM* 2014, 10:503–514.
- 74. Bear AS, Kennedy LC, Young JK, Perna SK, Mattos Almeida JP, Lin AY, Eckels PC, Drezek RA, Foster AE. Elimination of metastatic melanoma using gold nanoshell-enabled photothermal therapy and adoptive T cell transfer. *PLoS One* 2013, 8:e69073.
- 75. Wang C, Xu L, Liang C, Xiang J, Peng R, Liu Z. Immunological responses triggered by photothermal

therapy with carbon nanotubes in combination with anti-CTLA-4 therapy to inhibit cancer metastasis. *Adv Mater* 2014, 26:8154–8162.

- 76. Andersson H, Kim Y-S, O'Neill B, Shi Z-Z, Serda R. HSP70 promoter-driven activation of gene expression for immunotherapy using gold nanorods and near infrared light. *Vaccines* 2014, 2:216–227.
- 77. Burga RA, Patel S, Bollard CM, Y Cruz CR, Fernandes R. Conjugating prussian blue nanoparticles onto antigen-specific T cells as a combined nanoim-munotherapy. *Nanomedicine* 2016, 11:1759–1767.
- Munn DH, Bronte V. Immune suppressive mechanisms in the tumor microenvironment. *Curr Opin Immunol* 2016, 39:1–6.
- 79. Choi MR, Stanton-Maxey KJ, Stanley JK, Levin CS, Bardhan R, Akin D, Badve S, Sturgis J, Robinson JP, Bashir R, et al. A cellular trojan horse for delivery of therapeutic nanoparticles into tumors. *Nano Lett* 2007, 7:3759–3765.
- 80. Zhang J, Xia F, Yang Y, Yue C, Zhang C, Yang Y, Ma L, Alfranca G, Liu Y, Hou Y, et al. Human CIK cells loaded with gold nanoprisms as theranostic platform for targeted photoacoustic imaging and enhanced immuno-photothermal combined therapy. *Nanoscale Res Lett* 2016, 8:109–124.
- 81. Immuno-photothermal E. Human CIK cells loaded with gold nanoprisms as theranostic platform for targeted photoacoustic imaging and enhanced immunophotothermal combined therapy. *Nano Biomed Eng* 2016, 8:109–124.
- Kennedy LC, Bear AS, Young JK, Lewinski NA, Kim J, Foster AE, Drezek RA. T cells enhance gold nanoparticle delivery to tumors in vivo. *Nanoscale Res Lett* 2011, 6:283.
- Gao S, Zhang L, Wang G, Yang K, Chen M, Tian R, Ma Q, Zhu L. Hybrid graphene/Au activatable theranostic agent for multimodalities imaging guided enhanced photothermal therapy. *Biomaterials* 2016, 79:36–45.
- 84. Liu Y, Ashton JR, Moding EJ, Yuan H, Register JK, Fales AM, Choi J, Whitley MJ, Zhao X, Qi Y, et al. A plasmonic gold nanostar theranostic probe for in vivo tumor imaging and photothermal therapy. *Theranostics* 2015, 5:946–960.
- 85. Coughlin AJ, Ananta JS, Deng N, Larina IV, Decuzzi P, West JL. Gadolinium-conjugated gold nanoshells for multimodal diagnostic imaging and photothermal cancer therapy. *Small* 2014, 10:556–565.
- Loo C, Lin A, Hirsch L, Lee M-H, Barton J, Halas N, West J, Drezek R. Nanoshell-enabled photonics-based imaging and therapy of cancer. *Technol Cancer Res Treat* 2004, 3:33–40.
- 87. Gobin AM, Lee MH, Halas NJ, James WD, Drezek RA, West JL. Near-infrared resonant

nanoshells for combined optical imaging and photothermal cancer therapy. *Nano Lett* 2007, 7:1929–1934.

- 88. Huang P, Lin J, Li W, Rong P, Wang Z, Wang S, Wang X, Sun X, Aronova M, Niu G, et al. Biodegradable gold nanovesicles with an ultrastrong plasmonic coupling effect for photoacoustic imaging and photothermal therapy. *Angew Chem Int Ed* 2013, 52:13958–13964.
- 89. Tucker-Schwartz JM, Meyer TA, Patil CA, Duvall CL, Skala MC. In vivo photothermal optical coherence tomography of gold nanorod contrast agents. *Biomed Opt Express* 2012, 3:2881.
- 90. Li W, Chen X. Gold nanoparticles for photoacoustic imaging. *Nanomedicine (Lond)* 2015, 10:299–320.
- 91. Luke GP, Bashyam A, Homan KA, Makhija S, Chen Y, Emelianov SY. Silica-coated gold nanoplates as stable photoacoustic contrast agents for sentinel lymph node imaging. *Nanotechnology* 2013, 24:455101.
- 92. Luke GP, Myers JN, Emelianov SY, Sokolov K. Sentinel lymph node biopsy revisited: ultrasound-guided photoacoustic detection of micrometastases using molecularly targeted plasmonic nanosensors. *Cancer Res* 2014, 74:5397–5408.
- 93. Lu W, Melancon MP, Xiong C, Huang Q, Elliott A, Song S, Zhang R, Flores LG, Gelovani JG, Wang LV, et al. Effects of photoacoustic imaging and photothermal ablation therapy mediated by targeted hollow gold nanospheres in an orthotopic mouse xenograft model of glioma. *Cancer Res* 2011, 71:6116–6121.
- 94. Melancon MP, Elliott AM, Shetty A, Huang Q, Stafford RJ, Li C. Near-infrared light modulated photothermal effect increases vascular perfusion and enhances polymeric drug delivery. *J Control Release* 2011, 156:265–272.
- 95. Rylander M, Stafford J, Hazle J, Whitney J, Diller K. Heat shock protein expression and temperature distribution in prostate tumours treated with laser irradiation and nanoshells. *Int J Hyperthermia* 2011, 27:791–801.
- 96. Stauffer P, Craciunescu O, Maccarini P, Wyatt C, Arunachalam K, Arabe O, Stakhursky V, Li Z, Soher B, MacFall J, et al. Clinical utility of magnetic resonance thermal imaging (MRTI) for realtime guidance of deep hyperthermia. *Proc SPIE* 2009. doi:10.1117/12.812188
- 97. Sommer G, Bouley D, Gill H, Daniel B, Pauly KB, Diederich C. Focal ablation of prostate cancer: four roles for magnetic resonance imaging guidance. *Can J Urol* 2013, 20:6672–6681.
- 98. Vakoc BJ, Fukumura D, Jain RK, Bouma BE. Cancer imaging by optical coherence tomography: preclinical

progress and clinical potential. Nat Rev Cancer 2012, 12:363–368.

- Al-Mujaini A, Wali UK, Azeem S. Optical coherence tomography: clinical applications in medical practice. *Oman Med J* 2013, 28:86–91.
- 100. Adhi M, Duker JS. Optical coherence tomography current and future applications. *Curr Opin Ophthalmol* 2013, 24:213–221.
- 101. Valluru K, Wilson K, Willmann J. Photoacoustic imaging in oncology: translational preclinical and early clinical experience. *Radiology* 2016, 280(2):332-349.
- 102. Kharlamov AN, Gabinsky JL. Plasmonic photothermic and stem cell therapy of atherosclerotic plaque as a novel nanotool for angioplasty and artery remodeling. *Rejuvenation Res* 2012, 15:222–230.
- 103. Yeager D, Chen YS, Litovsky S, Emelianov S. Intravascular photoacoustics for image-guidance and temperature monitoring during plasmonic photothermal therapy of atherosclerotic plaques: a feasibility study. *Theranostics* 2014, 4:36–46.
- 104. Khantamat O, Li CH, Yu F, Jamison AC, Shih WC, Cai C, Lee TR. Gold nanoshell-decorated silicone surfaces for the near-infrared (NIR) photothermal destruction of the pathogenic bacterium *E. faecalis.* ACS Appl Mater Interfaces 2015, 7:3981–3993.
- 105. Millenbaugh NJ, Baskin JB, DeSilva MN, Elliott WR, Glickman RD. Photothermal killing of *Staphylococcus aureus* using antibody-targeted gold nanoparticles. *Int J Nanomedicine* 2015, 10:1953–1960.
- 106. Neumann O, Urban AS, Day J, Lal S, Nordlander P, Halas NJ. Solar vapor generation enabled by nanoparticles. *ACS Nano* 2013, 7:42–49.
- 107. Neumann O, Feronti C, Neumann AD, Dong A, Schell K, Lu B, Kim E, Quinn M, Thompson S, Grady N, et al. Compact solar autoclave based on steam generation using broadband light-harvesting nanoparticles. *Proc Natl Acad Sci U S A* 2013, 110:11677–11681.
- 108. Ibrahim IM, Ali HR. Treatment of natural mammary gland tumors in canines and felines using gold nanorods-assisted plasmonic photothermal therapy to induce tumor apoptosis. *Int J Nanomedicine* 2016, 11:4849–4863.
- 109. Schwartz JA, Shetty AM, Price RE, Stafford RJ, Wang JC, Uthamanthil RK, Pham K, McNichols RJ, Coleman CL, Payne JD. Feasibility study of particleassisted laser ablation of brain tumors in orthotopic canine model. *Cancer Res* 2009, 69:1659–1667.
- 110. Pedrosa P, Vinhas R, Fernandes A, Baptista P. Gold nanotheranostics: proof-of-concept or clinical tool? *Nanomaterials* 2015, 5:1853–1879.