A REVIEW OF FLUBENDAZOLE AND ITS POTENTIAL AS A MACROFILARIACIDE

A report submitted to Dr Gary Weil (PI DOLF) - a study supported by the BIII & Melinda Gates Foundation

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Goal: This report covers data on flubendazole reviewed in preparation for preparing a full drug development proposal to the B&M Gates Foundation for the use of the drug for filariasis

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1. INTRODUCTION

Flubendazole is a member of the benzimidazole class of heterocyclic aromatic organic compounds, a class that has been used extensively in a wide range of species. This class includes a range of active anthelminthic compounds that represent arguably the most important collection of anti-worm agents available today.

This report is the result of considering current literature, and of discussions with many experts in this field of work including those at pharmaceutical companies who are, or were, involved with the commercial distribution of this anthelmintic agent. The document does not attempt to review all the literature available but rather to present the main components that are relevant to the idea that this agent may have a major role as a macrofilaricide for improving the mass drug administration activities against river blindness (onchocerciasis) and lymphatic filariasis. It should be noted that some information on this compound was not available due to unavoidable circumstances (e.g. data/information had been destroyed, data not released for consideration by scientists, etc.). However, it is believed that the information that was not available would not have affected the overall conclusions made in this present report and the consequent second B&MG grant submission.

2. BASIC DATA

Chemical Structure:



There are two major metabolites for flubendazole, namely the hydrolysed form (FLU-NH2) and the reduced form (FLU-OH).



Figure 1. Structures of flubendazole (FUZ) and its metabolites, hydrolised flubendazole (FUZ-NH₂) and reduced flubendazole (FUZ-OH).

The three forms, parent and 2 metabolites are active in individuals that are treated with this drug and therefore all three compounds must be considered in the test procedures used to determine the safety of flubendazole if it to be used for filariasis. Two polymorphs of flubendazole exist - flubendazole and flubendazole polymorph B (2011-03-22. CAS : 31430-15-6) with Form B being variably described as being active or inactive.

Flubendazole, as with the benzimidazole group, is a specific inhibitor of microtubule assembly, acting by binding to the heterodimeric subunit, the tubulin molecule. The preferential binding of the benzimidazoles for parasite tubulin is some 100-400 fold greater than that to host tubulin; this fact is an important key to their safe use anthelminthic agents.

Names used for Flubendazole

Flubendazole Fluvermal Tricyclo[5.2.1.0]decan-8-one Tricyclo[5.2.1.0(2,6)] Decan-8-One tricyclo(5.2.1.02,6)decane - 8 - one Flubendazol Tricyclo[5.2.1.02,6]decan-8-one Flubenzimin

Flubendazole



Some minor differences exist in the commercial description of flubendazole.



FLUBENDAZOLE

CAS No. Chemical Name:

Synonyms: CBNumber:

Molecular

Formula:

260°C >100 °C 0-6°C

Formula Weight: MOL File:

37893-02-0 FLUBENDAZOLE

slj0312;cropotex;FLUMOXAL;FLUBENOL;FLUVERNAL;FLUMOXANE;FLUMOXANAL;bayslj0312;FLUBENDAZOL;FLUBENZIMIN CB5669631 C17H10F6N4S

37893-02-0.mol

FLUBENDAZOLE Property

mp:	
Fp:	
storage temp. :	

416.35

FLUBENDAZOLE REVIEW

Substance Name	Flubendazole
Synonyms	 * Flubendazolum [INN-Latin], * (5-(4-Fluorobenzoyl)-1H-benzimidazole-2-yl)carbamic acid methyl ester, * Methyl 5-(p-fluorobenzoyl)-2-benzimidazolecarbamate, * Methyl N-(5-(p-fluorobenzoyl)-2-benzimidazolyl)carbamate, * 2-Benzimidazolecarbamic acid, 5- (p-fluorobenzoyl), methyl ester, * AIDS-084892, * AIDS084892, * Carbamic acid, (5-(4-fluorobenzoyl)-1H-benzimidazol-2-yl)-, methyl ester, * Carbamic acid, [5- (4-fluorobenzoyl)-1H-benzimidazol-2-yl]-, methyl ester, * CCRIS 4480, * EINECS 250-624-4, * Flubendazol [INN-Spanish], * Flubenol, * Flumoxal, * Flumoxane, * Fluvermal, * N-(5-[(4-Fluorophenyl)carbonyl]benzimidazol-2-yl}methyloxymethanamide, * NSC 313680, *NSC313680, * R 17899, * R 17,889,
Chemical Name	methyl N-[5-(4-fluorobenzoyl)-3H-benzoimidazol-2-yl]carbamate
Molecular Formula	C ₁₆ H ₁₂ FN ₃ O ₃
Molecular Weight	313.283
CAS No.	31430-15-6
Chemical Structure	F C C C C C C C C C C C C C C C C C C C

Flubendazole is a synthetic anthelmintic belonging to the benzimidazole carbamates which acts by inhibiting the microtubular assembly in absorptive cells of nematodes. It acts by binding to tubulin, the dimeric subunit protein of the microtubules. It inhibits microtubular assembly in absorptive cells: i.e. of intestinal cells of nematodes. This is shown by disappearance of cytoplasmic microtubules,

Pharmacodynamic
Propertiesaccumulation of secretory granules in the cytoplasm due to a block in their
transport, leading to an impaired coating of the cellular membrane and a decreased
digestion and absorption of nutrients. Irreversible lytic degeneration of the cell, due
to the accumulation of secretory substances (hydrolytic and proteolytic enzymes),
results in the death of the parasite.Flubendazole is a broad-spectrum anthelminthic agent effective against
endoparasites such as gastro-intestinal ascarids, hookworms, whipworms found in
dogs, and active a range of gastrointestinal parasites in pigs and poultry such as
roundworms, and tapeworms, Ascaris suum (large roundworm), gapeworms,
Hyostrongylus rubidus (red stomach worm), Oesophagostomum dentatum (nodular
worm), Trichuris suis (whip worm), Strongyloides ransomi (adult), Metastrongylus
apri (lungworm).Flubendazole is ovicidal.

3. USE IN NON-HUMAN SPECIES

Flubendazole is a very commonly used compound in the veterinary world and is freely available for aquatic, avian and mammalian species, There are very few validated descriptions of adverse reactions to its use and the few described are associated with very high doses or the use of pro-drugs (which have considerably different pharmacology to flubendazole itself or the two metabolites, reduced flubendazole and hydrolyzed flubendazole).

Use with Aquatic Species

In fish it is used for controlling a number of organisms, including hydra, intestinal parasites (Heximata, gill flukes and Camallanus) possible by adsorption through the fish's skin. Regularly used dose levels are 0.5 g of 10% flubendazole in 20 gallons (75 liters) of water. Overall it is reported to be safe at comparatively high levels in its use with fish and the like. No delayed expression of toxicity were observed for 21 d after a 96-h exposure to flubendazole was noted, probably reflecting to the relatively high elimination constants for the chemical.

Use in Birds

Flubendazole is commonly used in the management of avian species. For example commercially available flubendazole-based products are efficacious against three species of helminth parasites of chickens: Ascaridia galli, Heterakis gallinarum and Capillaria spp; flubendazole achieves an overall efficacy of 99.4% for the three parasite species without any adverse side effects.

Flubenvet (2.5%) as a poultry dewormer is used as a medicated feed supplement commonly used to deworm chickens, turkeys and geese. **Flubenvet** is active against mature and immature nematodes of the respiratory and gastrointestinal tract. There are 3 major worms that usually affect chickens - these are roundworms, gapeworms and tapeworms. Flubenvet contains the active ingredient Flubendazole which has no adverse effect on egg laying or hatching. It is the only licenced in feed wormer for chickens currently available in the UK. It is very effective and can prevent a large number of problems and long term damage to your birds. You do not need to withdraw eggs for consumption when it is given at the correct dose and it is simple to administer in food.

The PK/PD aspects of flubendazole have been well studied in birds with acceptable results such as the following example: After oral administration of the

veterinary medicine Flubenol 5% the concentrations of the flubendazole-derived residues were determined by a liquid chromatographic-mass spectrometric method. The highest residue concentrations were obtained for the reduced metabolite. With the therapeutic dose, the maximum mean residue concentrations obtained for this compound in thigh muscle, breast muscle and liver were 312, 288 and 1043 ug/kg, respectively. The values for flubendazole, the parent molecule, were 114, 108 and 108 ug/kg, respectively. The residues of the hydrolysed metabolite were negligible in the sampled muscle tissues. After 24 h of depletion, the sum of the residues of parent and metabolites in muscle tissue still exceeded 50 ug/kg. After 8 d of depletion, flubendazole-derived residues at low concentrations could still be measured in both muscle tissues and liver. Thus flubendazole in this species has a wide tissue distribution, and consequently can reach parasites in different tissues.

Use in Domestic Species

Flubendazole has been used extensively in poultry and swine but lesser so on ruminants. the pharmacology in the latter species has been carried out and the data available for studies that would need to be carried out with bovine onchocerciasis in teh development of the drug for use in filariasis. No major differences appear to be likely with ruminants compared to mono-gastric animals. Little is published concerning this drug's use in small domestic animals (e.g. dogs) but oral reports stated that it was safe and effective.

4. USE IN HUMANS

Flubendazole, as originally formulated, is used to treat intestinal nematodes in humans. Flubendazole is registered and sold in Europe (EMEA) as Fluvermal (Johnson and Johnson, Sante Bea). **Flubendazole** is reported as an ingredient of Fluvermal in the following countries:

- Algeria
- Benin

- Burkina Faso
- Cameroon
- Central African Republic
- Colombia
- Congo
- Cote D'ivoire
- Cyprus
- Ecuador
- Egypt
- France
- Gabon
- Guinea
- Jordan
- Lebanon
- Madagascar
- Mali
- Mauritania
- Niger
- Peru
- Portugal
- Saudi Arabia
- Senegal
- Sudan
- Togo
- United Arab Emirates
- Yemen

The excipients used in Fluvermal are Talc (E553b), Saccharose, Amidon de pomme de terre, Amidon, Sodium laurylsulfate (E487), Magnésium stéarate (E572), Cellulose microcristalline (E460).

A 100mg dose of Fluvermal is most commonly proscribed for treating pinwoms (Enterobius vermiculus)). This is followed by a second dose of 100mg 15-21 days later to ensure reinfection is avoided, as flubendazole does not kill pinworm eggs. 100mg taken 3 times a day for 3 days is effective against larger nematodes, but only marginally effective against tapeworms. Fluvermal is available over the counter for human use in Europe under this brand name, and in 100mg tablets or as a 20mg/ml oral solution.

FLUVERMAL INSERT DATA

The form of flubendazole available for human use

Flubendazole

FLUVERMAL, tablets, 100 mg FLUVERMAL, oral suspension, 20 mg/ml FLUVERMAL, tablets + oral suspension, 100 mg + 20 mg/ml

Contraindications

Fluvermal is contraindicated in persons with a known hypersensitivity to the drug or its components.

Special warnings and special precautions for use

Use in infants < 1 year: as well-documented experience in children below 1 year of age is scarce, Fluvermal should only be given to very young children if their worm infection interferes significantly with their nutritional status and physical development.

The tablets contain lactose. Patients with rare hereditary diseases of galactose intolerance, Lapp's lactose deficiency or malabsorption of glucose-galactose should not use this medicinal product.

The oral suspension contains parabenes that can cause allergic reaction (possibly delayed).

Interaction with other medicaments and other forms of

interaction

None known.

Pregnancy and lactation

Fluvermal has shown embryotoxic and teratogenic activity in one study in rats. No such findings have been reported in teratology studies in the rabbit, mice or other studies in rats.

Experience in humans has not shown any increase in the risk of malformations. Nonetheless, it is better to avoid using the product in pregnant women or in those liable to become pregnant.

It is not known whether flubendazole is excreted in human milk. Therefore caution should be exercised when Fluvermal is administered to nursing women.

Effects on ability to drive and use machines

Fluvermal does not affect the mental alertness or driving ability.

Undesirable effects

Transient abdominal pain and diarrhoea have only rarely been reported, in cases of massive infestation and expulsion of worms.

Hypersensitivity reactions such as exanthema, rash, urticaria and angioedema have rarely been observed.

Overdose

Symptoms

In the event of accidental overdosage, abdominal cramps, nausea, vomiting and diarrhea may occur.

Although the maximum recommended treatment duration of Fluvermal is limited to three days there have been rare reports of liver function disturbances, hepatitis and blood dyscrasias described in patients who were treated for hydatid disease with massive doses for prolonged periods of time.

Treatment

There is no specific antidote. Within the first hour after ingestion, gastric lavage may be performed. Activated charcoal may be given if considered appropriate.

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As described below flubendazole was tested along with other benzimidazoles in humans for its effect on *Onchocerca volvulus* but as the preparation induced SAE's at the injections site it was not pursued further at that time (1981) - although it appeared to be very effective against this filarial pathogen.

5. SAFETY ISSUES

There are known issues with all this group of anthelminthics in terms of safety. The basic reason appears to be related to the effect of these agents in disrupting the tubulin of host cells and consequently the functions related to these structures. There are considerable differences in effects between species which may relate primarily to the pharmacodynamics in these species. However, it has been suggested that the vehicles that the primary compound is delivered in may also be important. The use of this property of binding tubulin in maintaining the health of the host (patient) is considered a potential advantage when used at higher levels. Combined with other cancer inhibiting drugs it is being proposed as as a treatment for certain types of neoplasia. In a study with leukemia and myeloma it was found that flubendazole induced cell death in leukemia and myeloma cell lines and primary patient samples at nano-molar concentrations. Moreover, it delayed tumor growth in leukemia and myeloma xenografts without evidence of toxicity. Mechanistically, flubendazole inhibited tubulin polymerization by binding tubulin at a site distinct from vinblastine. In addition, cells resistant to vinblastine because of overexpression of P-glycoprotein remained fully sensitive to flubendazole, indicating that flubendazole can overcome some forms of vinblastine resistance. Thus it was suggested by these authors that flubendazole is a novel microtubule inhibitor that displays pre-clinical activity in leukemia and myeloma.

An EMEA report (EMEA/CVMP/33128/2006-FINAL July 2006) states that flubendazole has a low acute oral and subcutaneous toxicity. The acute oral LD50 values were greater than 5000 mg/kg bw in mice, rats and guinea pigs. Acute subcutaneous LD50 values were greater than 5000 mg/kg bw in the rat and the mouse and 4679 and 4834 mg/kg bw in male and female guinea pigs, respectively. Any toxicity occurring appears to depend on route and on formulation.

FOOD RESIDUES

Flubendazole is one of the safest of drugs used in food animals in terms of the food residues. Residues of veterinary medicinal products, as defined by the European Union, are "pharmacologically active substances (whether active principles, excipients or degradation products) and their metabolites which remain in foodstuffs obtained from animals to which the veterinary medicinal product in question has been administered". The MRL is the maximum concentration of residue following administration of a veterinary medicine which is legally permitted or acceptable in food under the laws of the EU.

This is particularly important in the poultry industry where benzimidazoles are veterinary drugs widely used for prevention and treatment of parasitic infections. Metabolites of benzimidazoles have been reported in several matrices including eggs. However, to date flubendazole is the only anthelminthic that has an established MRL of 400 μ g kg-1 in egg. A value of 50 μ g kg-1 was established for all other anthelmintics. Thus flubendazole appears to be in terms of residues safer than most other anthelminthic agents.

6. POTENTIAL OF FLUBENDAZOLE AS A MACROFILARICIDE

A safe, field-usable chemotherapeutic agent that will rapidly kill adult filarial worms is urgently needed in tropical medicine. Ivermectin, distributed as Mectizan® by Merck & Co.Inc. has had an enormous impact on two major human filarial infections of developing countries, onchocerciasis and lymphatic filariasis. However, this agent works primary against the microfilarial stage and lacks the ability to rapidly kill the adult parasites. Since the adult worms can survive for many years producing offspring, it has been necessary for control programs to continue drug distribution for more than a decade, i.e., until the adult worms eventually die; a labor-intensive and expensive proposition. Other agents used in filarial control programs, such as diethylcarbamazine and albendazole, may be more effective macrofilaricides than ivermectin, but for various reasons are not suitable, or are unable, to fill the role of a being rapidly-acting macrofilariacide. Thus, a drug, administered once, or at least in multiple doses over a very short period, that safely kills adult filarial worms would be a major contributor to the current efforts to rid the world of filarial infections and the diseases they cause. A field useful agent has typically been required to be administered in an oral dosage form, but a truly safe agent administered by another route, including parenteral approaches, could be acceptable and may even be advantageous.

Given the challenges of discovery and development of agents for human use, a drug as described above is arguably most likely, at least at present, to come from the benzimidazole group of anthelmintics. Although several benzimidazoles are currently employed in human chemotherapy, there are other potential candidate macrofilaricides in other drug classes. However, time is of the essence in finding a new drug for use in ongoing filarial control programs, the first priority is to consider the benzimidazoles as the most likely source of a macrofilaricide. This group has provided many important effective agents for both veterinary and human medicine over the past 50 years, beginning with thiabendazole and now most prominently including albendazole and mebendazole for human parasites and a whole range of agents in veterinary medicine. Benzimidazoles work by interfering with the equilibrium among tubulin subunits, tubulin and microtubules. Not surprisingly, benzimidazoles can affect host tubulin as well as that of the parasites, are typically positive in mammalian cell cytotoxicity assays and cause chromosomal non-disjunction during mitosis. However, the benzimidazole anthelmintics show a differential preference for binding to nematode tubulin compared to mammalian tubulin, an important factor for development of a drug against nematodes in mammals. Benzimidazoles are also anti-fungal agents as well as anthelmintics, a fact that may be important in filarial conditions such as elephantiasis that involve secondary infections often involving fungi; albendazole is one of the two drugs used in the global lymphatic filariasis elimination program.

The most appealing benzimidazole with regard to filarial parasites is flubendazole as it is highly active against filariae in a number of hosts. It has the typical

benzimidazole structure with an added fluorine as the major structural difference from other benzimidazoles. It is a very efficacious macrofilaricide in a variety of experimental animals, with perhaps its most dramatic and relevant action being its ability to completely eliminate adult *Dirofilaria immitis* from dogs after a single injection. Flubendazole is was developed by Janssen in the mid 1970's and is currently licensed in Europe for the use as an anthelmintic in humans for intestinal nematodes (at 5 mg /kg for 3 days). Flubendazole is a potent and efficacious anthelmintic for gastrointestinal nematode infections in swine, poultry and companion animals, as well as against lungworms in swine. It is usually given over 3 days at ~5 mg/kg, but is probably also efficacious even as a single dose at this same rate. In a number of experimental filarial rodent models, flubendazole was found to have essentially 100% efficacy as a macrofilaricide at reasonable doses and schedules. A trial in human onchocerciasis was also carried in Mexico in the early 1980's with promising results. However, wider testing in humans was restricted at that time by problems associated with the route of administration and the relatively unsophisticated carrier agent used at that time, some 39 years ago. In addition, the introduction of ivermectin at about this time lessened the urgency to replace diethylcarbamazine for onchocerciasis control with a new macrofilaricide.

As noted, flubendazole is highly efficacious in various experimental filariasis models, including the feline *Brugia pahangi* model, a host in which it occurs naturally. Efficacy varies with parasite species, location in the host, and host species (Table 1). It should be noted that flubendazole is highly efficacious and potent as a macrofilaricide in these models only when given parenterally (in keeping with its very low oral bioavailability in standard formulations). Given parenterally, flubendazole is arguably the best macrofilaricide tested in animal models. Importantly, no adverse reactions were reported in any of these animal studies. An important observation, relevant to current

problems faced by the global control and elimination programs for human lymphatic filariasis and onchocerciasis, is that in cats and jirds infected with Brugia spp., flubendazole is active against adult worms but poorly active against the microfilarial stage. The significance of this observation lies in the fact that a major problem for filarial control programmes using ivermectin is that individuals co-infected with high levels of circulating Loa loa microfilariae may suffer severe adverse events. Over 124 people have died in the past ten years, usually with signs and symptoms of central nervous system pathology related to microfilarial death. An agent that will kill adult filariae but not microfilariae may be a breakthrough for this important practical problem, which currently limits ivermectin distribution programs in many Africa countries.

Table 1. Summary of lowest effective dose of flubendazole in filarial animal models.						
Parasite	LED ₉₆ × 5 (mg/kg)	LED,,, × 1 (mg/kg)	Ref.			
Jird						
Brugia pahangi [†]	1.5	25	[7]			
B. pahangi*	1.56	ND	[19]			
B. pahangi ^t	2.5	ND	[20]			
B. pahangi*	12.5	ND	[20]			
B. pahangi [*]	201	ND	[21]			
B. pahangi§	101	ND	[22]			
Dipetalonema viteae	100"	ND	[23]			
Acanthocheilonema viteae	1.56	ND	[19]			
Brugia pahangi*	25	ND	[24]			
B. malayi	12.5	50	[24]			
A. viteae	3.1	1.6	[24]			
Litomosoides carinii*	12.5	12.5	[24]			
Onchocerca lienalis ^{††}	100	ND	[25]			
B. pahangi [‡]	ND	100	[7]			
Adult parasites in the peritor Adult parasites in the lymph	neal cavity. atics.					

¹Not titrated, only dose reported.

"Multimammate rat. "Microfilariae transplanted into the skin.

Most of the efficacies reported at these doses in these studies were 100% Efficacy determinations are dependent on the time of necropsy, efficacy is higher (i.e., number of worms observed) in jirds necropsied 8 weeks posttreatment compared with 6 weeks post-treatment. [McCall Pers. Comm.] LED₅₀. Lowest dose that was at least 90% effective.

Following the encouraging findings in rodent models, a study was carried out in Mexico in the early 1980's in which several potential macrofilaricides, including flubendazole, were tested in humans infected with Onchocerca volvulus. This study was terminated early due to problems associated with reactions at the intra-muscular injection site where the flubendazole in its oil-based carrier was administered. Nevertheless, efficacy data on adult O. volvulus worms in surgically removed nodules from these patients suggested that flubendazole is a potent macrofilaricide. At 3 weeks after initiation of treatment (750 mg once per week for 5 weeks), significant degeneration of the adult worms was detected; at 5 weeks (Table 2), there was very effective destruction of the adult worms compared with the other anti-filarial agents.

Status of parasites	2 months post Rx		3 months post Rx	
	DEC	FLUB ⁺	DEC	FLUB
Degenerated adults	12	10	12	27
Intact adult worm	44	11	16	0
Females with empty uteri	6	1	5	0
Females with only oocytes	8	6	14	0
Reduction in dermal microfilariae*	Yes	No	Yes	No

*DEC (100 mg) was administered twice daily for 14 days and 750 mg FLUB was injected intramuscularly once a week for five doses.

*There was no significant ocular or skin pathology related to microfilarial death in those receiving FLUB. The only significant post-FLUB treatment reactions were associated with inflammation at the injection site. Dermal microfilarial loads stayed at pretreatment levels in the FLUB-treated individuals for approximately 6 months. DEC: Diethylcarbamazine; FLUB: Flubendazole; post RX: After last treatment with flubendazole.

'Data taken from [6.10].

As mentioned above, flubendazole is currently registered for human use in Europe for treatment of gut-residing nematodes, an action that does not require efficient uptake into the host's circulation. A challenge for ensuring its suitability for filariasis will be to develop a new formulation that will produce blood and tissue levels sufficient to destroy tissue-resident parasites, such as the filariae. Efficacy against filariae was not observed following oral dosing of flubendazole in any the early animal studies, but it should be noted that none of these studies used any of the new formulation methods now common in the pharmaceutical industry. Encouraging results come from Lanusse's group, which showed that the tissue-residing stage of the cestode, *Ecchinococcus* granulosis, can be killed by an orally administered flubendazole formulated with the now this could greatly enhance the likelihood of developing flubendazole as a suitable macrofilariacide for human filariasis for oral dosing. A hydroxypropyl-β-cyclodextrin formulation might indeed be suitable given the increased degree of bioavailability it provides; for example, it markedly enhances the bioavailability of albendazole, mebendazole and flubendazole. This material is approvable and is a gold-standard reagent for enhancing bioavailability of lipophilic drugs, and can be used in both liquid and solid dosage forms.

As the target infective adult filariae are complex and biochemically resourceful (many nematodes have the ability to switch biochemical pathways when stressed), it is likely that a relatively long duration of exposure to the drug will be needed. This may involve the need for dosing on multiple (e.g., 3-5) days to maintain lethal levels of the

DOLF

agent for the needed period of time. For many nematodes, acute exposure to benzimidazoles has few noticeable effects, even at very high concentrations; this is true for flubendazole in various adult filariids. As the drug acts by disrupting the tubulinmicrotubule equilibrium in cells, leading to cessation of nutrient transport and eventual cell death, these effects take time to become evident. *In vitro* experiments have shown that flubendazole concentrations as low as 100 ng/ml (incubated for 32 hours) disrupt tissue structure in parasitic nematodes in the same clade as filariae.

In addition to pharmacodynamic challenges, there are other hurdles to developing a safe and effective formulation of a drug for the treatment of complicated infections such as lymphatic filariasis and onchocerciasis. A primary concern with the benzimidazoles is safety. As these drugs interfere with microtubules, they have the potential of interfering with host cells, especially during cell division. Thus, the use of drugs such as albendazole is generally contra-indicated for pregnant women; this is likely to apply with a new flubendazole formulation that provides for systemic exposure. However, it should be noted that albendazole has been used very successfully in mass drug programs across the world since 1999, and that inadvertent treatment studies in pregnant women have not detected adverse effects on the unborn. Nevertheless, a major hurdle for a formulation that produces enhanced bioavailable flubendazole will need to be carefully evaluated for embryotoxicity. It may turn out that flubendazole is only useful for filarial infections in males and females outside childbearing ages. However, such a product still would be a most useful advance for control programs.

What will it take to determine if flubendazole is an important answer to the needs of filarial control and elimination programs? Scientifically, it will initially require the determination of the blood and tissue levels needed for macrofilaricidal efficacy; closely related is the need to determine the levels that induce toxicity. Both issues are central to moving forward with development of flubendazole. Based on recent experimental data from animal models, it is highly likely that current modern formulation techniques, including micronization, hydroxypropyl-β-cyclodextrin complexing or another new approach, will be able to provide the blood levels needed to kill adult worms. The testing of newly developed formulations for efficacy against filariae itself poses some challenge. Filarial infections are generally host specific and thus each filariae-host model is to some degree unique in form and properties. Flubendazole in a new formulation should be evaluated in a range of filarial models to encompass all the variations and characteristics of these infections, and to make predictions about the pharmacokinetic parameters likely to be required for efficacy in human infections; this would allow formulations to be evaluated on the basis of pharmacokinetic data rather than efficacy per se, which requires extended periods of time post-treatment. A combination of many disciplines and institutions will be needed, including, as with the pioneering onchocerciasis-ivermectin control program, "public-private partnerships" between the pharmaceutical industry, non-governmental organizations and academic scientists. Drug companies have the expertise needed to develop new formulations and are central to the final production phase needed; field-based expertise (Ministries of Health, NGDO's, academics) are essential for developing a practical field-based mass drug administration intervention and will be important partners in any successful effort.

The benefit of developing a safe and practical agent that needs distribution only once, or perhaps twice, is substantial when compared to what is currently in place, i.e., annual distribution for 8-12 years in filarial control and elimination programs; a highly

effective macrofilaricide would still be important even if a 3-5 daily course of treatment is needed. Financial savings as well as significant savings in terms of health personnel time commitment would be realized.

Flubendazole has great potential as a macrofilaricide. Its reformulation using modern pharmaceutical platforms should be expedited to enable efficacy testing as soon as possible. Although flubendazole faces, as does any new anthelmintic, important challenges with regard to safety and formulation, the potential benefits that could result relatively quickly from a safe, usable formulation of flubendazole make this a top priority for the filarial world today.

FLUBENDAZOLE REFERENCE LIST

This is a list of references that have been considered in this Review Document. Although extensive it does not purport to be a complete list of all references that mention flubendazole.

- Alcantara, A. K., Uylangco, C. V., and Cross, J. H. (1985) An obstinate case of intestinal capillariasis, *Southeast Asian J Trop Med Public Health 16*, 410-413.
- Alders, R. G., Bagnol, B., Young, M. P., Ahlers, C., Brum, E., and Rushton, J. (2007) Challenges and constraints to vaccination in developing countries, *Dev Biol* (*Basel*) 130, 73-82.
- Alders, R. G., and Hutchins, D. R. (1987) Chronic nephritis in a horse, Aust Vet J 64, 151-154.
- Alders, R. G., Landsverk, T., and Shelton, J. N. (1988) Mg2+-dependent adenosine triphosphatase: an enzyme marker for ovine T lymphocytes, *Immunol Cell Biol 66* (*Pt 5-6*), 361-367.
- Alders, R. G., and Press, C. M. (1990) Cellular composition of utero-ovarian lymph during the exogenous gonadotrophin-stimulated oestrous cycle in sheep, *Reprod Fertil Dev 2*, 165-172.
- Alders, R. G., and Shelton, J. N. (1988) Analysis of ovine peripheral blood lymphocyte subsets following Ficoll-Hypaque separation or erythrocyte lysis, *Res Vet Sci 45*, 253-254.
- Alders, R. G., and Shelton, J. N. (1990) Lymphocyte subpopulations in lymph and blood draining from the uterus and ovary in sheep, *J Reprod Immunol 17*, 27-40.

Allemandou, M. (2001) [Toxocariasis], Ann Dermatol Venereol 128, 1377.

Alvarez, L., Moreno, G., Moreno, L., Ceballos, L., Shaw, L., Fairweather, I., and Lanusse, C. (2009) Comparative assessment of albendazole and triclabendazole ovicidal activity on Fasciola hepatica eggs, *Vet Parasitol 164*, 211-216.

- Arru, E., Leoni, A., and Garippa, G. (1989) Morphobiology of Echinococcus granulosus and therapeutic trials in sheep, *J Chemother* 1, 255-256.
- Awadzi, K., Schulz-Key, H., Howells, R. E., Haddock, D. R., and Gilles, H. M. (1982) The chemotherapy of onchocerciasis VIII Levamisole and its combination with the benzimidazoles, *Ann Trop Med Parasitol 76*, 459-473.
- Azhar, M., Lubis, A. S., Siregar, E. S., Alders, R. G., Brum, E., McGrane, J., Morgan, I., and Roeder, P. (2010) Participatory disease surveillance and response in Indonesia: strengthening veterinary services and empowering communities to prevent and control highly pathogenic avian influenza, *Avian Dis 54*, 749-753.
- Baeyens, W., Abdel Fattah, F., and De Moerloose, P. (1986) Chromatographic isolation and mass spectrometric identification of a base-induced mebendazole fluorophor, *Pharmazie 41*, 709-710.
- Baeyens, W., Fattah, F. A., and De Moerloose, P. (1985) Fluorimetric analysis of mebendazole and flubendazole with hydrogen peroxide, *J Pharm Biomed Anal 3*, 397-404.
- Bagheri, H., Simiand, E., Montastruc, J. L., and Magnaval, J. F. (2004) Adverse drug reactions to anthelmintics, *Ann Pharmacother 38*, 383-388.
- Baiandina, D. G., Kazantseva, G. N., Krotov, A. I., Cherniaeva, A. I., and Naidenova, A.
 S. (1986) [Comparative effectiveness of mebendazole and flubendazole in experimental opisthorchiasis in golden hamsters], *Med Parazitol (Mosk)*, 38-39.
- Bakker, J. (1984) [Long-term effects of a deworming program using flubendazole and levamisole on the percentage of condemned livers in slaughtering pigs], *Tijdschr Diergeneeskd 109*, 815-819.
- Baliharova, V., Velik, J., Savlik, M., Szotakova, B., Lamka, J., Tahotna, L., and Skalova,
 L. (2004) The effects of fenbendazole, flubendazole and mebendazole on activities of hepatic cytochromes P450 in pig, *J Vet Pharmacol Ther 27*, 85-90.
- Balizs, G. (1999) Determination of benzimidazole residues using liquid chromatography and tandem mass spectrometry, *J Chromatogr B Biomed Sci Appl 727*, 167-177.
- Bartikova, H., Krizova, V., Lamka, J., Kubicek, V., Skalova, L., and Szotakova, B. (2010) Flubendazole metabolism and biotransformation enzymes activities in healthy sheep and sheep with haemonchosis, *J Vet Pharmacol Ther 33*, 56-62.
- Bartikova, H., Krizova, V., Stepnickova, M., Lamka, J., Kubicek, V., Skalova, L., and Szotakova, B. (2010) Activities of biotransformation enzymes and flubendazole metabolism in lambs (Ovis aries): effect of gender and flubendazole therapy, *Pharmacol Rep 62*, 362-373.

- Bauer, C., and Gerwert, S. (2002) Characteristics of a flubendazole resistant isolate of Oesophagostomum dentatum from Germany, *Vet Parasitol 103*, 89-97.
- Bauer, C., and Gey, A. (1995) Efficacy of six anthelmintics against luminal stages of Baylisascaris procyonis in naturally infected raccoons (Procyon lotor), *Vet Parasitol 60*, 155-159.
- Bauer, C., Taubert, A., and Hermosilla, C. (1999) Efficacy of two flubendazole formulations against Trichuris vulpis in naturally infected dogs, *Vet Rec 145*, 48.
- Baumeister, S., Dennis, R. D., Klunder, R., Schares, G., Zahner, H., and Geyer, E. (1994) Litomosoides carinii: macrofilariae-derived glycolipids--chromatography, serology and potential in the evaluation of anthelminthic efficacy, *Parasite Immunol 16*, 629-641.
- Becquet, R., Poirriez, J., Dei-Cas, E., Dutoit, E., Deblock, S., Abdellatifi, M., and Vernes, A. (1982) [Human trichostrongylosis (apropos of 71 case reports)], Ann Soc Belg Med Trop 62, 139-155.
- Beus, A. (1989) [Comparative study of thiabendazole and mebendazole in strongyloidiasis], *Lijec Vjesn 111*, 98-101.
- Bhopale, G. M., and Bhatnagar, B. S. (1985) The efficacy of some newer broad spectrum anthelmintics against third-stage larvae of Ancylostoma caninum in the mouse, *J Helminthol 59*, 307-311.
- Bhopale, G. M., and Bhatnagar, B. S. (1988) Efficacy of various anthelmintics against third-stage larvae of Ancylostoma caninum in the brain of mice, *J Helminthol 62*, 40-44.
- Boes, J., Kanora, A., Havn, K. T., Christiansen, S., Vestergaard-Nielsen, K., Jacobs, J., and Alban, L. (2010) Effect of Ascaris suum infection on performance of fattening pigs, *Vet Parasitol 172*, 269-276.
- Bories, C., Loiseau, P., Gueyouche, C., and Gayral, P. (1986) [Use of surviving infectious larvae of Dipetalonema dessetae in study and research on filaricidal substances], *J Pharmacol 17*, 301-307.
- Bouree, P., and Jerray, M. (1984) [Imidazoles in larval cestode infections], *Pathol Biol (Paris) 32*, 651-653.
- Bradley, R. E., Guerrero, J., Becker, H. N., Michael, B. F., and Newcomb, K. (1983) Flubendazole: dose range and efficacy studies against common internal parasites of swine, *Am J Vet Res 44*, 1329-1333.
- Bunnag, D., Harinasuta, T., Viravan, C., Jarupakorn, V., Chindanond, D., and Desakorn, V. (1980) Clinical trial of flubendazole on hookworm, Trichuris trichiura and

Ascaris lumbricoides infections, *Southeast Asian J Trop Med Public Health 11*, 363-366.

- Cabrera, B. D., Valdez, E. V., and Go, T. G. (1980) Clinical trials of broad spectrum anthelmintics against soil-transmitted helminthiasis, *Southeast Asian J Trop Med Public Health 11*, 502-506.
- Cacho, C., Turiel, E., and Perez-Conde, C. (2009) Molecularly imprinted polymers: an analytical tool for the determination of benzimidazole compounds in water samples, *Talanta 78*, 1029-1035.
- Campbell, B., Newcomb, K., and Guerrero, J. (1983) Evaluation of the safety of flubendazole premix in swine, *Am J Vet Res 44*, 486-489.
- Caumes, E., and Gentilini, M. (1993) [Treatment of cutaneous ancylostoma larva migrans], *Ann Dermatol Venereol 120*, 571-573.
- Cavier, R., and Notteghem, M. J. (1979) [Pharmacological study of tenicidal properties of two anthelmintic benzimidazole derivatives: mebendazole and flubendazole (author's transl)], *Ann Pharm Fr 37*, 33-36.
- Ceballos, L., Elissondo, C., Sanchez Bruni, S., Confalonieri, A., Denegri, G., Alvarez, L., and Lanusse, C. (2010) Chemoprophylactic activity of flubendazole in cystic echinococcosis, *Chemotherapy 56*, 386-392.
- Ceballos, L., Elissondo, M., Bruni, S. S., Denegri, G., Alvarez, L., and Lanusse, C. (2009) Flubendazole in cystic echinococcosis therapy: pharmaco-parasitological evaluation in mice, *Parasitol Int 58*, 354-358.
- Chevrel, B. (1981) [Treatment of intestinal worms with flubendazole], *Med Chir Dig 10*, 719-723.
- Chung, M. S., Joo, K. H., Quan, F. S., Kwon, H. S., and Cho, S. W. (2001) Efficacy of flubendazole and albendazole against Trichinella spiralis in mice, *Parasite 8*, S195-198.
- Chusattayanond, W., and Denham, D. A. (1984) Chemoprophylactic activity of flubendazole against Brugia pahangi in jirds, *J Parasitol 70*, 191-192.
- Chusattayanond, W., and Denham, D. A. (1984) Induction of host resistance to Brugia pahangi in jirds (Meriones unguiculatus) protected by chemoprophylaxis, *J Helminthol 58*, 245-249.
- Cook, G. C. (1990) Use of benzimidazole chemotherapy in human helminthiases: indications and efficacy, *Parasitol Today 6*, 133-136.

- Cossu, M. L., Carboni, G., Dessanti, A., Porcu, A., Cottu, P., Ena, M. A., Noya, G., and Dettori, G. (1994) [Biology, physiopathology and general aspects of echinococcosis-hydatidosis], *Ann Ital Chir 65*, 625-633.
- Court, J. P. (1982) A diffusion chamber technique for detecting compounds with clinical prophylactic activity against Brugia pahangi, *Tropenmed Parasitol 33*, 83-86.
- Court, J. P., Bianco, A. E., Townson, S., Ham, P. J., and Friedheim, E. (1985) Study on the activity of antiparasitic agents against Onchocerca lienalis third stage larvae in vitro, *Trop Med Parasitol 36*, 117-119.
- Court, J. P., and Lees, G. M. (1983) Improvement of in vitro culture conditions of Brugia pahangi four day old developing larvae for use in an antifilarial drug assay, *Tropenmed Parasitol 34*, 162-168.
- Court, J. P., Stables, J. N., Lees, G. M., Martin-Short, M. R., and Rankin, R. (1988) Dipetalonema viteae and Brugia pahangi transplant infections in gerbils for use in antifilarial screening, *J Helminthol 62*, 1-9.
- Cumino, A. C., Elissondo, M. C., and Denegri, G. M. (2009) Flubendazole interferes with a wide spectrum of cell homeostatic mechanisms in Echinococcus granulosus protoscoleces, *Parasitol Int 58*, 270-277.
- Cvilink, V., Kubicek, V., Nobilis, M., Krizova, V., Szotakova, B., Lamka, J., Varady, M., Kubenova, M., Novotna, R., Gavelova, M., and Skalova, L. (2008)
 Biotransformation of flubendazole and selected model xenobiotics in Haemonchus contortus, *Vet Parasitol 151*, 242-248.
- Cvilink, V., Skalova, L., Szotakova, B., Lamka, J., Kostiainen, R., and Ketola, R. A. (2008) LC-MS-MS identification of albendazole and flubendazole metabolites formed ex vivo by Haemonchus contortus, *Anal Bioanal Chem 391*, 337-343.
- Cvilink, V., Szotakova, B., Vokral, I., Bartikova, H., Lamka, J., and Skalova, L. (2009) Liquid chromatography/mass spectrometric identification of benzimidazole anthelminthics metabolites formed ex vivo by Dicrocoelium dendriticum, *Rapid Commun Mass Spectrom 23*, 2679-2684.
- Dangolla, A., Bjorn, H., Willeberg, P., and Barnes, E. H. (1997) Faecal egg count reduction percentage calculations to detect anthelmintic resistance in Oesophagostomum spp. in pigs, *Vet Parasitol 68*, 127-142.
- Davis, A., Pawlowski, Z. S., and Dixon, H. (1986) Multicentre clinical trials of benzimidazolecarbamates in human echinococcosis, *Bull World Health Organ 64*, 383-388.

- de Burbure, G., Kumar, V., and Vanparijs, O. (1986) Prophylactic activity of flubendazole-medicated feed on Brugia pahangi infection of multimammate rats, *Ann Trop Med Parasitol 80*, 455-457.
- Dangolla, A., Bjorn, H., Willeberg, P., and Barnes, E. H. (1997) Faecal egg count reduction percentage calculations to detect anthelmintic resistance in Oesophagostomum spp. in pigs, *Vet Parasitol 68*, 127-142.
- De Ruyck, H., Daeseleire, E., Grijspeerdt, K., De Ridder, H., Van Renterghem, R., and Huyghebaert, G. (2004) Distribution and depletion of flubendazole and its metabolites in edible tissues of guinea fowl, *Br Poult Sci 45*, 540-549.
- Dec Bronsvoort, B. M., Makepeace, B. L., Renz, A., Tanya, V. N., Fleckenstein, L., Ekale, D., and Trees, A. J. (2008) UMF-078: A modified flubendazole with potent macrofilaricidal activity against Onchocerca ochengi in African cattle, *Parasit Vectors 1*, 18.
- De Keyser, H. (1980) [Individual and stock deworming of swine with flubendazole], *Tierarztl Prax 8*, 163-170.
- De Rosa, F., Stagni, G., and Palumbo, M. (1980) [Therapy of experimental peritoneal hydatidosis in BALB/C mice with mebendazole and flubendazole administered orally], *Ann Sclavo 22*, 831-836.
- De Ruyck, H., Daeseleire, E., Grijspeerdt, K., De Ridder, H., Van Renterghem, R., and Huyghebaert, G. (2001) Determination of flubendazole and its metabolites in eggs and poultry muscle with liquid chromatography-tandem mass spectrometry, *J Agric Food Chem 49*, 610-617.
- De Ruyck, H., Daeseleire, E., Grijspeerdt, K., De Ridder, H., Van Renterghem, R., and Huyghebaert, G. (2004) Distribution and depletion of flubendazole and its metabolites in edible tissues of guinea fowl, *Br Poult Sci 45*, 540-549.
- de Silva, D. G., Lionel, N. D., and Jayatilleka, S. M. (1984) Flubendazole in the treatment of Ascaris lumbricoides and Trichuris trichiura: a comparison of two different regimens with single-dose, *Ceylon Med J 29*, 199-203.
- Dec Bronsvoort, B. M., Makepeace, B. L., Renz, A., Tanya, V. N., Fleckenstein, L., Ekale, D., and Trees, A. J. (2008) UMF-078: A modified flubendazole with potent macrofilaricidal activity against Onchocerca ochengi in African cattle, *Parasit Vectors 1*, 18.
- Denham, D. A. (1980) Anthelmintic properties of flubendazole against Dipetalonema viteae in jirds, *Trans R Soc Trop Med Hyg 74*, 829.

- Denham, D. A., and Brandt, E. (1980) Chemoprophylactic activity of flubendazole against adult Brugia pahangi transplanted into the peritoneal cavity of jirds, *J Parasitol 66*, 933-934.
- Denham, D. A., Samad, R., Cho, S. Y., Suswillo, R. R., and Skippins, S. C. (1979) The anthelmintic effects of flubendazole on Brugia pahangi, *Trans R Soc Trop Med Hyg 73*, 673-676.
- Devaney, E., Howells, R. E., and Smith, G. (1985) Brugia pahangi in the BALB/C mouse: a model for testing filaricidal compounds, *J Helminthol 59*, 95-99.
- Dominguez Vazquez, A., and Rivas Alcala, A. R. (1985) [Recent investigations in the chemotherapy of onchocerciasis], *Salud Publica Mex 27*, 21-30.
- Dominguez-Vazquez, A., Taylor, H. R., Greene, B. M., Ruvalcaba-Macias, A. M., Rivas-Alcala, A. R., Murphy, R. P., and Beltran-Hernandez, F. (1983) Comparison of flubendazole and diethylcarbamazine in treatment of onchocerciasis, *Lancet 1*, 139-143.
- Duong, T. H., Barrabes, A., Renier, M., and Combescot, G. (1980) [Use of fluoromebenzadole in the treatment of opisthorchiasis], *Nouv Presse Med 9*, 256.
- Dusi, G., Gamba, V., and Faggionato, E. (2005) Rapid determination of the antiparasitic drugs flubendazole and febantel in feeds by HPLC with ultraviolet detection, *J Pharm Biomed Anal 38*, 375-379.
- Dutoit, E., Poirriez, J., Leclercq, P., Vernes, A., and Fontaine, G. (1983) [Value of flubendazole in the treatment of toxocariasis], *Presse Med 12*, 2396.
- Dymon, M., and Papir, B. (2004) [Effect of anthelmintic therapy supplemented with glucan in experimental toxocarosis], *Wiad Parazytol 50*, 465-470.
- Dzhabarova, V. I., and Krotov, A. I. (1987) [Experimental chemotherapy of alveococcosis. XI. Comparative efficacy of mebendazole, flubendazole and albendazole], *Med Parazitol (Mosk)*, 15-19.
- El-Enany, N., Belal, F., and Rizk, M. (2003) Polarographic determination of flubendazole in spiked human urine and plasma, *Farmaco 58*, 613-617.
- el-Temsahi, M. M., and el-Mansoury, S. T. (1995) The effect of flubendazole on the course of Trichinella spiralis infection in mice: parasitological study, *J Egypt Soc Parasitol 25*, 453-459.

- Elissondo, M., Ceballos, L., Dopchiz, M., Andresiuk, V., Alvarez, L., Bruni, S. S., Lanusse, C., and Denegri, G. (2007) In vitro and in vivo effects of flubendazole on Echinococcus granulosus metacestodes, *Parasitol Res 100*, 1003-1009.
- Elissondo, M., Dopchiz, M., Ceballos, L., Alvarez, L., Sanchez Bruni, S., Lanusse, C., and Denegri, G. (2006) In vitro effects of flubendazole on Echinococcus granulosus protoscoleces, *Parasitol Res 98*, 317-323.
- Elissondo, M. C., Ceballos, L., Alvarez, L., Sanchez Bruni, S., Lanusse, C., and Denegri, G. (2009) Flubendazole and ivermectin in vitro combination therapy produces a marked effect on Echinococcus granulosus protoscoleces and metacestodes, *Parasitol Res 105*, 835-842.
- Ellrodt, A., Halfon, P., Le Bras, P., Halimi, P., Bouree, P., Desi, M., and Caquet, R. (1987) Multifocal central nervous system lesions in three patients with trichinosis, *Arch Neurol* 44, 432-434.
- Esterre, P., and Jamet, P. (1987) [New antifungal and antiparasitic molecules], *Med Trop* (*Mars*) 47, 47-52.
- Feldmeier, H., Bienzle, U., Dohring, E., and Dietrich, M. (1982) Flubendazole versus mebendazole in intestinal helminthic infections, *Acta Trop 39*, 185-189.
- Fok, E., and Kassai, T. (1998) Toxocara canis infection in the paratenic host: a study on the chemosusceptibility of the somatic larvae in mice, *Vet Parasitol 74*, 243-259.
- Fontan, I., Taieb, A., Klene, C., and Maleville, J. (1986) [Critical review of scabies treatments], *Ann Dermatol Venereol 113*, 593-596.
- Fourestie, V., Bougnoux, M. E., Ancelle, T., Liance, M., Roudot-Thoraval, F., Naga, H., Pairon-Pennachioni, M., Rauss, A., and Lejonc, J. L. (1988) Randomized trial of albendazole versus tiabendazole plus flubendazole during an outbreak of human trichinellosis, *Parasitol Res 75*, 36-41.
- Fournier, J. P., Marty, P., Bernard, E., Leloire, P., Dellamonica, P., and Le Fichoux, Y. (1985) [Spinal hydatidosis treated by albendazole. A propos of 2 cases], *Pathol Biol (Paris) 33*, 611-613.
- Froyman, R., and De Keyser, H. (1983) Flubendazole: safety regarding egg production and reproductive performance of breeder chickens, *Avian Dis 27*, 43-48.
- Franz, M., Zahner, H., and Benten, P. (1990) Fine-structure alterations in female Brugia malayi and Litomosoides carinii after in vivo treatment with flubendazole, *Parasitol Res 76*, 401-405.

- Froyman, R., and De Keyser, H. (1983) Flubendazole: safety regarding egg production and reproductive performance of breeder chickens, *Avian Dis 27*, 43-48.
- Gallais, H., Raoult, D., Xeridat, B., Dumon, H., and Quilici, M. (1983) [Hydatic cyst of the pelvis. Failure of treatment with flubendazole], *Bull Soc Pathol Exot Filiales 76*, 709-712.Gauly, M., Duss, C., and Erhardt, G. (2007) Influence of Ascaridia galli infections and anthelmintic treatments on the behaviour and social ranks of laying hens (Gallus gallus domesticus), *Vet Parasitol 146*, 271-280.
- Gayral, P., Dreyfuss, G., and Gantier, J. C. (1982) [Dipetalonema dessetae in Proechimys oris. II. Evaluation of the model for pharmacologic investigations of antifilarial chemotherapy (author's transl)], *J Pharmacol 13*, 49-63.
- Geary, T. G., Woo, K., McCarthy, J. S., Mackenzie, C. D., Horton, J., Prichard, R. K., de Silva, N. R., Olliaro, P. L., Lazdins-Helds, J. K., Engels, D. A., Bundy, D. A. Unresolved issues in anthelmintic pharmacology for helminthiases of humans. Int J Parasitol. 40, 1-13 (2009).
- Gerwert, S., Failing, K., and Bauer, C. (2002) Prevalence of levamisole and benzimidazole resistance in oesophagostomum populations of pig-breeding farms in North Rhine-Westphalia, Germany, *Parasitol Res 88*, 63-68.
- Ghannad, E., Abbou, C. B., Nottin, R., Hourdebaigt-Larrusse, P., Soulie, J., and Grivaux, M. (1983) [Cardiac hydatidosis], *Sem Hop 59*, 1459-1463.
- Gibson, K. T., and Alders, R. G. (1987) Eosinophilic enterocolitis and dermatitis in two horses, *Equine Vet J 19*, 247-252.
- Gratteri, P., Pinzauti, S., La Porta, E., Mura, P., Papeschi, G., and Santoni, G. (1990) Determination of flubendazole in pharmaceutical dosage forms by differential pulse polarography and UV spectroscopy, *Farmaco 45*, 707-714.
- Guerrero, J., Campbell Seibert, B. P., Newcomb, K. M., Michael, B. F., and McCall, J. W. (1983) Activity of flubendazole against developing stages of Dirofilaria immitis in dogs, *Am J Vet Res 44*, 2405-2406.
- Hanser, E., Mehlhorn, H., Hoeben, D., and Vlaminck, K. (2003) In vitro studies on the effects of flubendazole against Toxocara canis and Ascaris suum, *Parasitol Res 89*, 63-74.
- Harendra de Silva, D. G., Lionel, N. D., Premaratne, U. N., Warnasuriya, N., and Soysa, P. E. (1987) Flubendazole in the treatment of soil transmitted helminths, *Ceylon Med J 32*, 129-132.

- Harrison, J. L., and Alders, R. G. (2010) An assessment of chicken husbandry including Newcastle disease control in rural areas of Chibuto, Mozambique, *Trop Anim Health Prod 42*, 729-736.
- Hauck, R., and Hafez, H. M. (2009) Partial sequence of the beta-tubulin of Histomonas meleagridis and the activity of benzimidazoles against H. meleagridis in vitro, *Parasitol Res 104*, 1183-1189.

Hopkins, A D Ivermectin and onchocerciasis: is it all solved? Eye (Lond). 19, 1057-66 (2005).

- Hutchinson, G. W. (1981) Anthelmintic treatment of prepatent stephanuriasis with flubendazole, levamisole and disophenol and the effects on liver-specific serum enzymes, *Res Vet Sci 30*, 175-180.
- Issar, M., Nagaraja, N. V., Lal, J., Paliwal, J. K., and Gupta, R. C. (1999) Determination of antifilarial compound UMF-078 and its metabolites in plasma by highperformance liquid chromatography, *J Chromatogr B Biomed Sci Appl 724*, 147-155.
- Jacquemin, J. L. (1980) [Rampant linear dermatitis ("larbish"). Resistance to thiabendazole and efficacy of fluoromebendazole], *Nouv Presse Med 9*, 1779.
- Janssen, P. A., and van den Bossche, H. (1982) Treatment of helminthiasis, *Scand J* Infect Dis Suppl 36, 52-57.
- Jerray, M., Benzarti, M., Garrouche, A., Klabi, N., and Hayouni, A. (1992) Hydatid disease of the lungs. Study of 386 cases, *Am Rev Respir Dis 146*, 185-189.
- Juliano, C., Martinotti, M. G., and Cappuccinelli, P. (1985) "In vitro" effect of microtubule inhibitors on Trichomonas vaginalis, *Microbiologica 8*, 31-42.
- Kamaraj, C., Rahuman, A. A., Elango, G., Bagavan, A., and Zahir, A. A. (2010) Anthelmintic activity of botanical extracts against sheep gastrointestinal nematodes, Haemonchus contortus, *Parasitol Res*.
- Kamath, V. R., Menon, S., Bhopale, M. K., Deshpande, V. R., and Renapurkar, D. M. (1987) Experimental chemotherapy of Angiostrongylus cantonensis infection in mice with flubendazole, levamisole, and their combination, *Folia Parasitol* (*Praha*) 34, 87-92.
- Kan, C. A., Keukens, H. J., and Tomassen, M. J. (1998) Flubendazole residues in eggs after oral administration to laying hens: determination with reversed phase liquid chromatography, *Analyst 123*, 2525-2527.

- Kan, S. P. (1983) The anthelmintic effects of flubendazole on Trichuris trichiura and Ascaris lumbricoides, *Trans R Soc Trop Med Hyg 77*, 668-670.
- Kanda, S., and Maki, J. (1992) In vitro observation on egg release by Angiostrongylus cantonensis from rats treated with flubendazole, *Kitasato Arch Exp Med 65*, 155-158.
- Katiyar, S. K., Gordon, V. R., McLaughlin, G. L., and Edlind, T. D. (1994) Antiprotozoal activities of benzimidazoles and correlations with beta-tubulin sequence, *Antimicrob Agents Chemother 38*, 2086-2090.
- Kim, H. J., Lee, D. S., and Kwon, J. H. (2010) Sorption of benzimidazole anthelmintics to dissolved organic matter surrogates and sewage sludge, *Chemosphere 80*, 256-262.
- Krizova, V., Nobilis, M., Pruskova, L., Chladek, J., Szotakova, B., Cvilink, V., Skalova,
 L., and Lamka, J. (2009) Pharmacokinetics of flubendazole and its metabolites in lambs and adult sheep (Ovis aries), *J Vet Pharmacol Ther 32*, 606-612.
- Krotov, A. I., and Dzhabarova, V. I. (1986) [Current status of the chemotherapy of echinococcosis (a review of the literature)], *Med Parazitol (Mosk)*, 82-85.
- Kumar, V., Ceulemans, F., and De Meurichy, W. (1978) Chemotherapy of helminthiasis among wild mammals. IV. Efficacy of flubendazole 5% (R17889) against Trichuris trichiura infection of baboons, Papio hamadryas L, *Acta Zool Pathol Antverp*, 3-9.
- Kumar, V., Ghenim, H., Brandt, J., and Vanparijs, O. (1987) Flubendazole in-feed preparation for prophylaxis of experimental lymphatic filariasis, *Ann Soc Belg Med Trop 67*, 75-77.
- Lamka, J., Vondrejc, M., and Kleca kova, J. (1996) [Effect of flubendazole on Muellerius capillaris in mouflon], *Vet Med (Praha) 41*, 347-350.
- Lassegue, A., Estavoyer, J. M., Minazzi, H., Barale, T., Gillet, M., Vuitton, D., and Miguet, J. P. (1984) [Treatment of human alveolar echinococcosis with flubendazole. Clinical, morphological and immunological study], *Gastroenterol Clin Biol 8*, 314-320.
- Latif, L. A., and Surin, J. (1993) Relationships between the anthelmintic activity of eight derivatives of benzimidazole carbamates against Trichinella spiralis and their chemical structures, *Jpn J Med Sci Biol 46*, 203-214.
- Lelong, M., Wattre, P., Vaudour, G., Bras, C., Bouvier, C., and Drain, J. P. (1986) [What problems does childhood toxocariasis currently pose? Apropos of 6 clinical cases], *Allerg Immunol (Paris) 18*, 23-27.

- Maeda, R., Hayashi, Y., and Shibuya, T. (1988) Basic studies on the laboratory assessment of macrofilaricides using Brugia malayi in the jird, Meriones unguiculatus. 2. Establishment and evaluation of a new method of macrofilaricide assessment, *Jpn J Exp Med 58*, 45-49.
- Mahmoud, M. S., and Moustafa, M. A. (2003) Cystatin capture-dot-enzyme-linked immunosorbent assay for immunodiagnosis and assessment of cure of experimental trichinellosis in mice, *J Egypt Soc Parasitol 33*, 275-290.
- Mak, J. W. (1981) Antifilarial activity of mebendazole and flubendazole on Breinlia booliati, *Trans R Soc Trop Med Hyg 75*, 306-307.
- Maki, J., and Kanda, S. (1992) Higher sensitivity of the developing larvae of Angiostrongylus cantonensis than the adult worms to flubendazole and mebendazole, *Kitasato Arch Exp Med 65*, 131-136.
- Maki, J., and Kanda, S. (1992) Significance of diaphragm sampling for determining larvicidal effect of flubendazole and mebendazole on Trichinella spiralis in mice, *Kitasato Arch Exp Med 65*, 53-56.
- Maki, J., Kondo, A., and Yanagisawa, T. (1983) Effects of alcoholic extract from Ma-Klua (Diospyros mollis) on adults and larvae of the dwarf tapeworm, Hymenolepis nana in mice and on the infectivity of the eggs, *Parasitology 87 (Pt 1)*, 103-111.
- Maki, J., and Yanagisawa, T. (1983) A comparison of the effects of flubendazole and thiabendazole on the larvae of Angiostrongylus cantonensis, Trichinella spiralis, Diphyllobothrium erinacei and Hymenolepis nana in mice, *Parasitology 87 (Pt 3)*, 525-531.
- Maki, J., and Yanagisawa, T. (1985) Larvicidal effect of flubendazole on Angiostrongylus cantonensis in mice with various worm burdens, *J Helminthol 59*, 301-302.
- Maki, J., and Yanagisawa, T. (1985) Anthelmintic effects of bithionol, paromomycin sulphate, flubendazole and mebendazole on mature and immature Hymenolepis nana in mice, *J Helminthol 59*, 211-216.
- Maki, J., and Yanagisawa, T. (1986) Studies on anthelmintic effects of flubendazole and mebendazole on the rat lungworm Angiostrongylus cantonensis in mice and rats, *J Parasitol 72*, 512-516.
- Maki, J., and Yanagisawa, T. (1988) Comparative efficacy of flubendazole and mebendazole on encysted larvae of Trichinella spiralis (USA strain) in the diaphragm of mice and rats, *J Helminthol 62*, 35-39.

- Maki, J., and Yanagisawa, T. (1990) Effect of flubendazole on the number of first-stage larvae of Angiostrongylus cantonensis released in the faeces of treated rats, *J Helminthol 64*, 87-95.
- Marinculic, A., Fajdiga, M., and Durakovic, E. (2001) The efficacy of flubendazole against Trichinella spiralis in swine, *Parasite 8*, S191-194.
- Mate, L., Virkel, G., Lifschitz, A., Ballent, M., and Lanusse, C. (2008) Hepatic and extrahepatic metabolic pathways involved in flubendazole biotransformation in sheep, *Biochem Pharmacol 76*, 773-783.
- Mentz, M. B., and Graeff-Teixeira, C. (2003) Drug trials for treatment of human angiostrongyliasis, *Rev Inst Med Trop Sao Paulo 45*, 179-184.
- Michiels, M., Hendriks, R., Heykants, J., and van den Bossche, H. (1982) The pharmacokinetics of mebendazole and flubendazole in animals and man, *Arch Int Pharmacodyn Ther 256*, 180-191.
- Min, D. Y., Ahn, M. H., and Kim, K. M. (1986) [Effect of flubendazole against Ascaris lumbricoides, Trichocephalus trichiurus and Enterobius vermicularis in infected children], *Kisaengchunghak Chapchi 24*, 12-14.
- Molina, J. M., Goguel, J., Sarfati, C., Chastang, C., Desportes-Livage, I., Michiels, J. F., Maslo, C., Katlama, C., Cotte, L., Leport, C., Raffi, F., Derouin, F., and Modai, J. (1997) Potential efficacy of fumagillin in intestinal microsporidiosis due to Enterocytozoon bieneusi in patients with HIV infection: results of a drug screening study. The French Microsporidiosis Study Group, *AIDS 11*, 1603-1610.
- Molinier, S., Chaudier, B., Kraemer, P., Graffin, B., San, V. V., Imbert, P., Morillon, M., Touze, J. E., and Gras, C. (1998) [Diagnostic and treatment of hypereosinophilia upon return from the tropics: 102 patients], *Med Trop (Mars) 58*, 499-502.
- Moreno, L., Alvarez, L., Mottier, L., Virkel, G., Bruni, S. S., and Lanusse, C. (2004) Integrated pharmacological assessment of flubendazole potential for use in sheep: disposition kinetics, liver metabolism and parasite diffusion ability, *J Vet Pharmacol Ther 27*, 299-308.
- Morris, D. L., and Gould, S. E. (1982) Serum and cyst concentrations of mebendazole and flubendazole in hydatid disease, *Br Med J (Clin Res Ed) 285*, 175.
- Mottier, L., Alvarez, L., and Lanusse, C. (2003) Quantitative chromatographic determination of several benzimidazole anthelmintic molecules in parasite material, *J Chromatogr B Analyt Technol Biomed Life Sci 798*, 117-125.
- Neri, B., Bidolli, G., Felli, M., and Cozzani, R. (2002) Determination of benzimidazole anthelmintics in animal-derived biological matrices, *Ann Chim 92*, 451-456.

- Nessim, N. G., Hassan, S. I., William, S., and el-Baz, H. (2000) Effect of the broad spectrum anthelmintic drug flubendazole upon Schistosoma mansoni experimentally infected mice, *Arzneimittelforschung 50*, 1129-1133.
- Nianjun, H., Cerepnalkoski, L., Nwankwo, J. O., Dews, M., and Landolph, J. R. (1994) Induction of chromosomal aberrations, cytotoxicity, and morphological transformation in mammalian cells by the antiparasitic drug flubendazole and the antineoplastic drug harringtonine, *Fundam Appl Toxicol 22*, 304-313.
- Nobilis, M., Jira, T., Lisa, M., Holcapek, M., Szotakova, B., Lamka, J., and Skalova, L. (2007) Achiral and chiral high-performance liquid chromatographic determination of flubendazole and its metabolites in biomatrices using UV photodiode-array and mass spectrometric detection, *J Chromatogr A 1149*, 112-120.
- Nobilis, M., Vybiralova, Z., Krizova, V., Kubicek, V., Soukupova, M., Lamka, J., Szotakova, B., and Skalova, L. (2008) Sensitive chiral high-performance liquid chromatographic determination of anthelmintic flubendazole and its phase I metabolites in blood plasma using UV photodiode-array and fluorescence detection Application to pharmacokinetic studies in sheep, *J Chromatogr B Analyt Technol Biomed Life Sci 876*, 89-96.
- Nogami, S., Hayashi, Y., Nakamura, M., Maeda, R., and Tanaka, H. (1987) Effect of milbemycin D on Dipetalonema viteae in the hamster and in vitro, *Jpn J Exp Med 57*, 237-239.
- Notteghem, M. J., Leger, N., and Cavier, R. (1979) [Study of fluke-killing activity of flubendazole on Echinostoma caproni Richard, 1964 (author's transl)], *Ann Pharm Fr 37*, 153-156.
- Oh, S. J., Park, J., Lee, M. J., Park, S. Y., Lee, J. H., and Choi, K. (2006) Ecological hazard assessment of major veterinary benzimidazoles: acute and chronic toxicities to aquatic microbes and invertebrates, *Environ Toxicol Chem 25*, 2221-2226.
- Oosterhuis, B., Wetsteyn, J. C., and van Boxtel, C. J. (1984) Liquid chromatography with electrochemical detection for monitoring mebendazole and hydroxymebendazole in echinococcosis patients, *Ther Drug Monit 6*, 215-220.
- Pedrosa, C., Costa, H., Oliveira, G., Romariz, J., and Praca, F. (2005) Anaphylaxis to povidone in a child, *Pediatr Allergy Immunol 16*, 361-362.
- Penot, C., Picot, H., and Lavarde, V. (1978) [Therapeutic trials of a new anti-helminthic in Colombian Amazonia: flubendazole], *Bull Soc Pathol Exot Filiales 71*, 370-375.

- Petersen, M. B., Friis, C., and Bjorn, H. (1997) A new in vitro assay of benzimidazole activity against adult Oesophagostomum dentatum, *Int J Parasitol 27*, 1333-1339.
- Petigny, A., Savoye, B., Vilain, C., Boulez, J., Girard, M., and Prud'hon, M. C. (1980) [Multiple hydatidosis: hepatic, splenic, peritoneal and gynaecologic locations (author's transl)], *Sem Hop 56*, 685-687.
- Petit, A., Duong, T. H., Bremond, J. L., Barrabes, A., Binet, C., Combescot, C., and Leroux, M. E. (1981) [Irregular anti-P1 allo-antibodies and sinensis clonorchiasis], *Rev Fr Transfus Immunohematol 24*, 197-210.
- Piens, M. A., Excler, J. L., Maisonneuve, H., Paillard, B., and Garin, J. P. (1984) [Flubendazole in human Echinococcus granulosus hydatidosis. Preoperative care: parasit-pharmacologic study], *Bull Soc Pathol Exot Filiales 77*, 69-80.
- Prelezov, P. N., and Bauer, C. (2003) Comparative efficacy of flubendazole chewable tablets and a tablet combination of febantel, pyrantel embonate and praziquantel against Trichuris vulpis in experimentally infected dogs, *Dtsch Tierarztl Wochenschr 110*, 419-421.
- Raeymaekers, A. H., Van Gelder, J. L., Roevens, L. F., and Janssen, P. A. (1978) Synthesis and anthelminthic activity of alkyl-(5-acyl-1-H-benzimidazol-2-yl) carbamates, *Arzneimittelforschung 28*, 586-594.
- Ramanathan, S., Nair, N. K., Mansor, S. M., and Navaratnam, V. (1994) Determination of the antifilarial drug UMF-078 and its metabolites UMF-060 and flubendazole in whole blood using high-performance liquid chromatography, *J Chromatogr B Biomed Appl 655*, 269-273.
- Reddy, A. B., Rao, U. R., Chandrashekar, R., Shrivastava, R., and Subrahmanyam, D. (1983) Comparative efficacy of some benzimidazoles and amoscanate (Go.9333) against experimental filarial infections, *Tropenmed Parasitol 34*, 259-262.
- Roche, G., Canton, P., Gerard, A., and Dureux, J. B. (1982) [Treatment of alveolar echinococcosis with flubendazole. Pharmacological study (author's transl)], *Pathol Biol (Paris) 30*, 452-457.
- Rossignol, J. F., and Maisonneuve, H. (1984) Benzimidazoles in the treatment of trichuriasis: a review, *Ann Trop Med Parasitol 78*, 135-144.
- Saimot, A. G. (1981) [Medical treatment for hydatidosis?], *Nouv Presse Med 10*, 3119-3120.
- Saimot, A. G., Meulemans, A., Hay, J. M., Mohler, J., Manuel, C., and Coulaud, J. P. (1981) [Pharmacokinetic study of flubendazole in human hydatid disease caused

by Echinococcus granulosus. Preliminary results (author's transl)], *Nouv Presse Med 10*, 3121-3124.

- Saunders, M., Taubert, A., Dafa'alla, T., and Zahner, H. (2008) Effect of chemotherapeutic treatment on cytokine (IFN-gamma, IL-2, IL-4, IL-5, IL-10) gene transcription in response to specific antigens in Brugia malayi-infected Mastomys coucha, *Parasitol Res 103*, 1163-1176.
- Savlik, M., Fimanova, K., Szotakova, B., Lamka, J., and Skalova, L. (2006) Modulation of porcine biotransformation enzymes by anthelmintic therapy with fenbendazole and flubendazole, *Res Vet Sci 80*, 267-274.
- Savlik, M., Polackova, L., Szotakova, B., Lamka, J., Velik, J., and Skalova, L. (2007) Activities of biotransformation enzymes in pheasant (Phasianus colchicus) and their modulation by in vivo administration of mebendazole and flubendazole, *Res Vet Sci 83*, 20-26.
- Savlik, M., Polaskova, P., Szotakova, B., Lamka, J., and Skalova, L. (2005) The effects of flubendazole and mebendazole on cytochromes P4501A in pheasant hepatocytes, *Res Vet Sci 79*, 139-147.
- Schantz, P. M., Van den Bossche, H., and Eckert, J. (1982) Chemotherapy for larval echinococcosis in animals and humans: report of a workshop, *Z Parasitenkd 67*, 5-26.
- Schenone, H., Galdames, M., Inzunza, E., Jimenez, M., Romero, E., and Bloomfield, R. (1977) [Flubendazole in the treatment of intestinal nematode infections in children (author's transl)], *Bol Chil Parasitol 32*, 85-86.
- Shcherbakov, A. M. (1986) [Optimization of the chemotherapy of echinococcal disease (the current state of the problem and the means for its resolution)], *Med Parazitol (Mosk)*, 57-61.
- Sinsheimer, J. E., Giri, A. K., Osorio, S., Wise, D. S., and Townsend, L. B. (1990) Comparative mutagenicity and genotoxicity of the antiparasitic drugs, mebendazole, flubendazole and flubendazole oxime, *Prog Clin Biol Res 340E*, 225-234.
- Six, R. H., Sture, G. H., Thomas, C. A., Clemence, R. G., Benchaoui, H. A., Boy, M. G., Watson, P., Smith, D. G., Jernigan, A. D., and Rowan, T. G. (2000) Efficacy and safety of selamectin against gastrointestinal nematodes in cats presented as veterinary patients, *Vet Parasitol 91*, 321-331.
- Smedts Walters, H., Bader-Meunier, B., and Dommergues, J. P. (1998) -Hemophagocytic syndrome associated with strongyloidiasis in a child treated for acute lymphoblastic leukemia, *Arch Pediatr 5*, 346-347.

- Snabel, V., DeMeews, T., Varady, M., Nansen, P., Bjorn, H., and Corba, J. (2000) The sexually linked Mpi locus is presumably involved in imidothiazole resistance in Oesophagostomum dentatum parasites, *Parasitol Res 86*, 486-490.
- Sohn, W. M., Kim, H. M., Chung, D. I., and Yee, S. T. (2000) The first human case of Trichinella spiralis infection in Korea, *Korean J Parasitol 38*, 111-115.
- Spagnuolo, P. A., Hu, J., Hurren, R., Wang, X., Gronda, M., Sukhai, M. A., Di Meo, A., Boss, J., Ashali, I., Beheshti Zavareh, R., Fine, N., Simpson, C. D., Sharmeen, S., Rottapel, R., and Schimmer, A. D. (2010) The antihelmintic flubendazole inhibits microtubule function through a mechanism distinct from Vinca alkaloids and displays preclinical activity in leukemia and myeloma, *Blood 115*, 4824-4833.
- Strote, G., Wieland, S., Darge, K., and Comley, J. C. (1990) In vitro assessment of the activity of anthelmintic compounds on adults of Onchocerca volvulus, *Acta Leiden 59*, 285-296.
- Surin, J. (1995) Anti-nematode activity of sixteen compounds against Trichinella spiralis in mice--a possible new screen for macrofilaricides, *Southeast Asian J Trop Med Public Health 26*, 128-134.
- Surin, J., and Denham, D. A. (1990) Comparative susceptibility to anthelmintics of Brugia pahangi in jirds infected by different methods, *J Helminthol 64*, 232-238.
- Szotakova, B., Nobilis, M., Lamka, J., Krizova, V., Savlik, M., and Skalova, L. (2008) Modulation of porcine (Sus scrofa domestica) and pheasant (Phasianus colchicus) carbonyl reducing enzymes by anthelmintic therapy with flubendazole, *Drug Metab Lett 2*, 29-34.
- Tawatsin, A., Sirisinha, S., Vajrasthira, S., Bunnag, D., and Harinasuta, T. (1984) Evaluation of selected anthelmintic compounds for activity against Opisthorchis viverrini, *Tropenmed Parasitol 35*, 85-90.
- Tellez-Giron, E., Ramos, M. C., Dufour, L., Montante, M., Tellez, E., Jr., Rodriguez, J., Gomez Mendez, F., and Mireles, E. (1984) Treatment of neurocysticercosis with flubendazole, *Am J Trop Med Hyg 33*, 627-631.
- Tellez-Giron, E., Ramos, M. C., and Montante, M. (1981) Effect of flubendazole on Cysticercus cellulosae in pigs, *Am J Trop Med Hyg 30*, 135-138.
- Theplertboon, R., Fleckenstein, L., Dzimianski, M. T., and McCall, J. W. (2000) Pharmacokinetics of UMF-078, a candidate antifilarial drug, in infected dogs, *Am J Trop Med Hyg 62*, 86-91.

- Thienpont, D., Vanparijs, O., Hermans, L., and De Roose, P. (1982) Treatment of Trichuris suis infections in pigs with flubendazole, *Vet Rec 110*, 517-520.
- Thienpont, D., Vanparijs, O., Niemegeers, C., and Marsboom, R. (1978) Biological and pharmacological properties of flubendazole, *Arzneimittelforschung 28*, 605-612.
- Tojo, J. L., and Santamarina, M. T. (1998) Oral pharmacological treatments for parasitic diseases of rainbow trout oncorhynchus mykiss. III. Ichthyobodo necator, *Dis Aquat Organ 33*, 195-199.
- Tojo, J. L., and Santamarina, M. T. (1998) Oral pharmacological treatments for parasitic diseases of rainbow trout Oncorhynchus mykiss. II. Gyrodactylus sp, *Dis Aquat Organ 33*, 187-193.
- Tojo, J. L., and Santamarina, M. T. (1998) Oral pharmacological treatments for parasitic diseases of rainbow trout Oncorhynchus mykiss. I: Hexamita salmonis, *Dis Aquat Organ 33*, 51-56.
- Townson, S., Connelly, C., Dobinson, A., and Muller, R. (1987) Drug activity against Onchocerca gutturosa males in vitro: a model for chemotherapeutic research on onchocerciasis, *J Helminthol* 61, 271-281.
- Townson, S., Dobinson, A., Connelly, C., and Muller, R. (1988) Chemotherapy of Onchocerca lienalis microfilariae in mice: a model for the evaluation of novel compounds for the treatment of onchocerciasis, *J Helminthol 62*, 181-194.
- Townson, S., Dobinson, A. R., Townsend, J., Siemienska, J., and Zea-Flores, G. (1990) The effects of ivermectin used in combination with other known antiparasitic drugs on adult Onchocerca gutturosa and O. volvulus in vitro, *Trans R Soc Trop Med Hyg 84*, 411-416.
- Underwood, N. (2009) Supply of Flubenvet 2.5% 240 g medicated premixture, *Vet Rec 165*, 271-272.
- Van De Steene, J. C., and Lambert, W. E. (2008) Comparison of matrix effects in HPLC-MS/MS and UPLC-MS/MS analysis of nine basic pharmaceuticals in surface waters, *J Am Soc Mass Spectrom 19*, 713-718.
- Van De Steene, J. C., and Lambert, W. E. (2008) Validation of a solid-phase extraction and liquid chromatography-electrospray tandem mass spectrometric method for the determination of nine basic pharmaceuticals in wastewater and surface water samples, *J Chromatogr A 1182*, 153-160.
- Van de Steene, J. C., Mortier, K. A., and Lambert, W. E. (2006) Tackling matrix effects during development of a liquid chromatographic-electrospray ionisation tandem

mass spectrometric analysis of nine basic pharmaceuticals in aqueous environmental samples, *J Chromatogr A 1123*, 71-81.

- Van De Steene, J. C., Stove, C. P., and Lambert, W. E. (2010) A field study on 8 pharmaceuticals and 1 pesticide in Belgium: removal rates in waste water treatment plants and occurrence in surface water, *Sci Total Environ 408*, 3448-3453.
- Van den Bossche, H. (1981) A new look at the mode of action of some old and new antifilarial compounds, *Ann Soc Belg Med Trop 61*, 287-296.
- Van Kerckhoven, I., and Kumar, V. (1988) Macrofilaricidal activity of oral flubendazole on Brugia pahangi, *Trans R Soc Trop Med Hyg 82*, 890-891.
- van Krimpen, M. M., Binnendijk, G. P., Borgsteede, F. H., and Gaasenbeek, C. P. (2010) Anthelmintic effects of phytogenic feed additives in Ascaris suum inoculated pigs, *Vet Parasitol 168*, 269-277.
- Vanparijs, O. (1984) Anthelmintic activity of flubendazole in naturally infected geese and the economic importance of deworming, *Avian Dis 28*, 526-531.
- Vanparijs, O. (1986) Chemotherapy of experimental echinococcosis in mice, *Ann Trop Med Parasitol 80*, 601-605.
- Vanparijs, O. (1990) Chemotherapy of experimental Echinococcus multilocularis in jirds, *Parasitol Res 76*, 238-240.
- Vanparijs, O., Hermans, L., and Marsboom, R. (1988) Efficacy of flubendazole against gastrointestinal and lung nematodes in pigs, *Vet Rec 123*, 337-339.
- Vanparijs, O., Hermans, L., and Van der Flaes, L. (1985) Anthelmintic efficacy of flubendazole paste against nematodes and cestodes in dogs and cats, *Am J Vet Res 46*, 2539-2541.
- Varma, T. K., Shinghal, T. N., Saxena, M., and Ahluwalia, S. S. (1990) Studies on the comparative efficacy of mebendazole, flubendazole and niclosamide against human tapeworm infections, *Indian J Public Health 34*, 163-168.
- Velkers, F. C., Dieho, K., Pecher, F. W., Vernooij, J. C., van Eck, J. H., and Landman, W. J. (2011) Efficacy of allicin from garlic against Ascaridia galli infection in chickens, *Poult Sci 90*, 364-368.
- Vetchy, D., Rabiskova, M., and Sucman, E. (2000) [Effect of temperature on drug solubility in complex formation of flubendazole and 2-hydroxypropyl-beta-cyclodextrin], *Ceska Slov Farm 49*, 239-241.

- Vuitton, D. A., Lassegue, A., Miguet, J. P., Herve, P., Barale, T., Seilles, E., and Capron,
 A. (1984) Humoral and cellular immunity in patients with hepatic alveolar
 echinococcosis. A 2 year follow-up with and without flubendazole treatment,
 Parasite Immunol 6, 329-340.
- Ward, S. A., Mihaly, G. W., Tjia, J. F., and Back, D. J. (1985) The effect of some benzimidazoles on the disposition of antipyrine and tolbutamide from the rat isolated perfused liver, *J Pharm Pharmacol 37*, 62-64.
- Watts, S. D., Rapson, E. B., Atkins, A. M., and Lee, D. L. (1982) Inhibition of acetylcholinesterase secretion from Nippostrongylus brasiliensis by benzimidazole anthelmintics, *Biochem Pharmacol 31*, 3035-3040.
- Weber, M., Vespignani, H., Jacquier, P., Gerard, A., Claudon, P., Bracard, S., and Floquet, J. (1988) [Neurological manifestations of alveolar echinococcosis], *Rev Neurol (Paris) 144*, 104-112.
- Weiss, K., Schussler, W., and Porzelt, M. (2008) Sulfamethazine and flubendazole in seepage water after the sprinkling of manured areas, *Chemosphere 72*, 1292-1297.
- William, S., Guirguis, F., and Nessim, N. G. (2003) Effect of simultaneous and/or consecutive administration of the broad spectrum anthelmintic flubendazole together with praziquantel in experimental Schistosoma mansoni infection, *Arzneimittelforschung 53*, 532-537.
- Yangco, B. G., Klein, T. W., Deresinski, S. C., Vickery, A. C., and Craig, C. P. (1981) Flubendazole and mebendazole in the treatment of trichuriasis and other helminthiases, *Clin Ther 4*, 285-290.
- Yoshimura, H. (1987) Teratogenicity of flubendazole in rats, *Toxicology 43*, 133-138.
- Yoshimura, H. (2003) Effect of oral dosing vehicles on the developmental toxicity of flubendazole in rats, *Reprod Toxicol 17*, 377-385.
- Youssef, F. G., Mikhail, E. M., and Mansour, N. S. (1989) Intestinal capillariasis in Egypt: a case report, *Am J Trop Med Hyg 40*, 195-196.
- Zahner, H., and Schares, G. (1993) Experimental chemotherapy of filariasis: comparative evaluation of the efficacy of filaricidal compounds in Mastomys coucha infected with Litomosoides carinii, Acanthocheilonema viteae, Brugia malayi and B. pahangi, *Acta Trop 52*, 221-266.

- Hauck, R., and Hafez, H. M. (2009) Partial sequence of the beta-tubulin of Histomonas meleagridis and the activity of benzimidazoles against H. meleagridis in vitro, *Parasitol Res 104*, 1183-1189.
- Katiyar, S. K., Gordon, V. R., McLaughlin, G. L., and Edlind, T. D. (1994) Antiprotozoal activities of benzimidazoles and correlations with beta-tubulin sequence, *Antimicrob Agents Chemother 38*, 2086-2090.Lassegue, A., Estavoyer, J. M., Minazzi, H., Barale, T., Gillet, M., Vuitton, D., and Miguet, J. P. (1984) [Treatment of human alveolar echinococcosis with flubendazole. Clinical, morphological and immunological study], *Gastroenterol Clin Biol 8*, 314-320.
- Maki, J., and Yanagisawa, T. (1983) A comparison of the effects of flubendazole and thiabendazole on the larvae of Angiostrongylus cantonensis, Trichinella spiralis, Diphyllobothrium erinacei and Hymenolepis nana in mice, *Parasitology 87 (Pt 3)*, 525-531.
- Molina, J. M., Goguel, J., Sarfati, C., Chastang, C., Desportes-Livage, I., Michiels, J. F., Maslo, C., Katlama, C., Cotte, L., Leport, C., Raffi, F., Derouin, F., and Modai, J. (1997) Potential efficacy of fumagillin in intestinal microsporidiosis due to Enterocytozoon bieneusi in patients with HIV infection: results of a drug screening study. The French Microsporidiosis Study Group, *AIDS 11*, 1603-1610.
- Nianjun, H., Cerepnalkoski, L., Nwankwo, J. O., Dews, M., and Landolph, J. R. (1994) Induction of chromosomal aberrations, cytotoxicity, and morphological transformation in mammalian cells by the antiparasitic drug flubendazole and the antineoplastic drug harringtonine, *Fundam Appl Toxicol 22*, 304-313.

Delatour P., Richard Y. The embryotoxic and antimitotic properties of a series of benzimidazoles. Therapie 31, 505-515 (1976).

Lacey, E. The role of the cytoskeletal protein, tubulin, in the mode of action and mechanism of drug resistance to benzimidazoles. Int J Parasitol 7, 885-936 (1988).

Bradley, R. E., Guerrero, J., Becker, H. N., Michael, B. F., Newcomb, K.. Flubendazole: dose range and efficacy studies against common internal parasites of swine. Am J Vet Res. 7, 1329-1333 (1983).

Dominguez-Vasquez A, Taylor HR, Greene BM, Ruvalcaba-Macias AM, Rivas-Alcala AR, Murphy RP, Beltran-Hernandez F. Comparison of flubendazole and diethylcarbamazine in treatment of onchocerciasis. Lancet 1(8317), 139-43 (1983).

Denham, D. A., Samad, R., Cho, S. Y., Suswillo, R. R., Skippins, S. C. The anthelmintic effects of flubendazole on Brugia pahangi. Trans R Soc Trop Med Hyg 73, 673-6. (1979).

Denham D.A., Brandt, E. Chemoprophylactic activity of flubendazole against adult Brugia pahangi transplanted into the peritoneal cavity of jirds. J Parasitol 66, 933-4 (1980).

Mackenzie, C. D., Geary, T. G., Gerlach, J. A. Possible pathogenic pathways in the adverse clinical events seen following ivermectin administration to onchocerciasis patients. Filaria J. 2 (supp 1) S5 (2003).

Mackenzie C.D., Martinez-Palomo A. (1982) unpublished report.

Ceballos L, Elissondo M, Bruni SS, Denegri G, Alvarez L, Lanusse C. Flubendazole in cystic echinococcosis therapy: Pharmaco-parasitological evaluation in mice. Parasitology International. 58, 354-8 (2009).

Challa, R. Ahuja A., Ali J., and Khar R.K. Cyclodextrins in Drug Delivery: An Updated Review. AAPS PharmSciTech 2005; 6, Article 43 (<u>http://www.aapspharmscitech.org</u>) (2005).

Rigter IM, Schipper HG, Koopmans RP, van Kan HJM, Frijlink HW, Kager PA, Guchelaar H-J. Relative bioavailability of three newly developed albendazole formulations: a randomized crossover study with healthy volunteers. Antimicrob. Agents Chemother. 48: 1051-4, (2004).

Lahiani-Skiba M, Coquard A, Bounoure F, Verite P, Arnaud P, Skiba M. Mebendazole complexes with various cyclodextrins: preparation and physicochemical characterization. J. Inclusion Phenom. Macrocyclic Chem. 57: 197-201, (2007).

Pax RA, Williams JF, Guderian RH. In vitro motility of isolated adults and segments of *Onchocerca volvulus, Brugia pahangi* and *Acanthocheilonema viteae*. Tropical Medicine and Parasitology 39 (Supplement): 450-5, (1988).

Satti MZ, VandeWaa EA, Bennett JL, Williams JF, Conder GA, McCall JW. Comparative effects of anthelmintics on motility in vitro of *Onchocerca gutterosa, Brugia pahangi* and *Acanthocheilonema viteae*. Trop. Med. Parasitol.39 (Supplement): 480-3 (1988).

Hanser E, Mehlhorn H, Hoeben D, Vlaminck K. In vitro studies on the effects of flubendazole against *Toxocara canis* and *Ascaris suum*. Parasitology Research 89: 63-74, (2003).

Gyapong, J. O., Chinbuah, M. A., Gyapong, M. Inadvertent exposure of pregnant women to ivermectin and albendazole during mass drug administration for lymphatic

filariasis Trop Med Int Health 8, 1093-1101 (2003).

- Davidse L.C. (1986) Benzimidazole Fungicides: Mechanism of Action and Biological Impact. Annual Review of Phytopathology, Vol. 24: 43-65.
- Lacey E. (1990) Mode of action of benzimidazoles Parasitology Today 6: 112-115
- Martin R.J., , Robertson A.P and Bjorn H. (1997) Target sites of anthelmintics. Parasitology , 114 : 111-124
- Friedman, P.A. and Platzer, E.G. (1980) Interaction of anthelmintic benzimidazoles with *AscarisSuum* embryonic tubulin. Biochim.Blophys. Acta 630,271-278
- Oh, S. J., Park, J., Lee, M. J., Park, S. Y., Lee, J. H., and Choi, K. (2006) Ecological hazard assessment of major veterinary benzimidazoles: acute and chronic toxicities to aquatic microbes and invertebrates, *Environ Toxicol Chem 25*, 2221-2226.
- Sinsheimer, J. E., Giri, A. K., Osorio, S., Wise, D. S., and Townsend, L. B. (1990) Comparative mutagenicity and genotoxicity of the antiparasitic drugs, mebendazole, flubendazole and flubendazole oxime, *Prog Clin Biol Res 340E*, 225-234.
- Six, R. H., Sture, G. H., Thomas, C. A., Clemence, R. G., Benchaoui, H. A., Boy, M. G., Watson, P., Smith, D. G., Jernigan, A. D., and Rowan, T. G. (2000) Efficacy and safety of selamectin against gastrointestinal nematodes in cats presented as veterinary patients, *Vet Parasitol 91*, 321-331.
- Smedts Walters, H., Bader-Meunier, B., and Dommergues, J. P. (1998) -Hemophagocytic syndrome associated with strongyloidiasis in a child treated for acute lymphoblastic leukemia, *Arch Pediatr 5*, 346-347.
- Spagnuolo, P. A., Hu, J., Hurren, R., Wang, X., Gronda, M., Sukhai, M. A., Di Meo, A., Boss, J., Ashali, I., Beheshti Zavareh, R., Fine, N., Simpson, C. D., Sharmeen, S., Rottapel, R., and Schimmer, A. D. (2010) The antihelmintic flubendazole inhibits microtubule function through a mechanism distinct from Vinca alkaloids and displays preclinical activity in leukemia and myeloma, *Blood 115*, 4824-4833.
- Thienpont, D., Vanparijs, O., Niemegeers, C., and Marsboom, R. (1978) Biological and pharmacological properties of flubendazole, *Arzneimittelforschung 28*, 605-612.

Yoshimura, H. (1987) Teratogenicity of flubendazole in rats, *Toxicology 43*, 133-138.

Yoshimura, H. (2003) Effect of oral dosing vehicles on the developmental toxicity of flubendazole in rats, *Reprod Toxicol 17*, 377-385.

8. APPENDICES

8.1

FLUVERMAL INSERT

Flubendazole

FLUVERMAL, tablets, 100 mg FLUVERMAL, oral suspension, 20 mg/ml FLUVERMAL, tablets + oral suspension, 100 mg + 20 mg/ml

Contraindications

Fluvermal is contraindicated in persons with a known hypersensitivity to the drug or its components.

Special warnings and special precautions for use

Use in infants < 1 year: as well-documented experience in children below 1 year of age is scarce, Fluvermal should only be given to very young children if their worm infection interferes significantly with their nutritional status and physical development.

The tablets contain lactose. Patients with rare hereditary diseases of galactose intolerance, Lapp's lactose deficiency or malabsorption of glucose-galactose should not use this medicinal product.

The oral suspension contains parabenes that can cause allergic reaction (possibly delayed).

Interaction with other medicaments and other forms of interaction

None known.

Pregnancy and lactation

Fluvermal has shown embryotoxic and teratogenic activity in one study in rats. No such findings have been reported in teratology studies in the rabbit, mice or other studies in rats.

Experience in humans has not shown any increase in the risk of malformations. Nonetheless, it is better to avoid using the product in pregnant women or in those liable to become pregnant.

It is not known whether flubendazole is excreted in human milk. Therefore caution should be exercised when Fluvermal is administered to nursing women.

Effects on ability to drive and use machines

Fluvermal does not affect the mental alertness or driving ability.

Undesirable effects

Transient abdominal pain and diarrhoea have only rarely been reported, in cases of massive infestation and expulsion of worms.

update_21092009

Hypersensitivity reactions such as exanthema, rash, urticaria and angioedema have rarely been observed.

Overdose

Symptoms

In the event of accidental overdosage, abdominal cramps, nausea, vomiting and diarrhea may occur.

Although the maximum recommended treatment duration of Fluvermal is limited to three days there have been rare reports of liver function disturbances, hepatitis and blood dyscrasias described in patients who were treated for hydatid disease with massive doses for prolonged periods of time.

Treatment

There is no specific antidote. Within the first hour after ingestion, gastric lavage may be performed. Activated charcoal may be given if considered appropriate.

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The European Agency for the Evaluation of Medicinal Products Veterinary Medicines Evaluation Unit

COMMITTEE FOR VETERINARY MEDICINAL PRODUCTS

FLUBENDAZOLE

SUMMARY REPORT (1)

Flubendazole is a benzimidazole anthelmintic which is administered for therapeutic applications in pigs, chickens, turkeys and game birds. The preparations available include tablets, pastes, pellets and premixes. In some countries it is also available as an antheimintic for human use.

Flubendazole was shown to be poorly absorbed from the gastro-intestinal tract with most of the administered drug excreted unchanged in the faeces. It was of low acute toxicity. 2

administered drug excreted unchanged in the faces. It was of low acute toxicity. Duily oral dones of up to 40 mg/kg bw per day of flubendancie were given to dogs for 3 months. Some slight histological changes, which were difficult to interpret, were seen in both the male and fenanle genital tracts. The changes in females were considered that the changes in males (prostatic flurists and not related to treatment. It was also considered that the changes in males (prostatic flurists) were probably not treatment-related. However, as a precautionary measure, and due to the absence of conclusive evidence regarding the activity of the findings, it was agreed that the done level of 2.5 mg/kg bw per day should be regarded as a NOEL for the study. No adverse effects were observed in a 3-month feeding study in which rates were given up to 1600 mg/kg flubendancie in the dist, equivalent to 117-147 mg/kg bw per day.

Flubendazole was tested for mutagenicity in a wide range of *in vitro* and *in vitro* assays, all of which gave negative results. In carcinogenicity studies, rats and mice were fed dietary concentrations of flubendazole timended to provide up to 40 mag/kgb wflubendazole per day; no increase in tumour incidence was observed and there were no other treatment-related effects.

increase in tumour incidence was observed and there were no other treatment-related effects. The reproductive toxicity of flubendazole was investigated in a range of species. No effects on fertility were observed in rats or dogs. In some studies and rats and rabbits, evidence of foetoxicity (increased incidences of abortions and resorptions and reduced foetal weights) was observed at very high dose levels. There was also evidence of teratogenicity in the rat. The results of a published study, in rats, in which the apparent NOEL for teratogenicity, was 10 mg/kg buy erg day were discounted because the flubendazole used had been extracted from a commercial formulation and was of doubtful purity. Instead, it was concluded that a more reliable NOEL for teratogenicity of 40 mg/kg buy per day had been established in a better-documented unpublished study, in rats.

Flubendazole had no significant antimicrobial activity. 3,

4.

It was noted that the JECFA had estimated an ADI of 0-12 $\mu g/kg$ bw per day by applying a safety factor of 200 to the NOEL of 2.5 mg/kg bw per day in the 3-month dog study. The safety factor of 200 was used to take into account the fact that the does were administered 6 days per week only.

It was agreed that the JECFA evaluation was not significantly different from the CVMP Working Group on Safety of Residues interpretation of the data and it was therefore agreed that the JECFA ADI should be adopted.

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5. Carbamate hydrolysis and ketone reduction were the main biotransformation pathways in pigs and the (2-amino-1H-benzimidazo15-yi)-4-fluorophenyi-methanone metabolite was the major drug-related component in pig kidney. However, there was no validated malytical method for the routine determination of this metabolite in tissues and it was therefore agreed that the flubendazote parent compound should be considered the marker residue.

The following provisional MLRs were elaborated :

Chicken: liver 0.5 mg/kg; muscle 0.2 mg/kg; eggs 0.4 mg/kg; Porcine: edible tissues (muscle, liver, kidney, fat) 0.01 mg/kg.

It was calculated that the intake of total extractable flubendazole-derived residues, arising from consumption of animal products containing residues which did not exceed these MRLs, would not exceed the ADI proposed above.

- An analytical method was available for the determination of the parent flubendazole using HPLC with UV detection at 312 nm. The limit of detection of the method was 0.01 mg/kg for all tissues. Validation data for this analytical method was provided only for phesant tissues. 6.
- 7. The following data are required before 31 December 1994:
 - 7.1 Further information on the relationship between extractable flubendazole-related residues and total (extractable & bound) residues in swine tissues;
 - 7.2 Information on whether M7 is the appropriate marker residue in pigs and other species; if it is, a satisfactory analytical method for determination of this metabolite should be developed and fully validated;
 - 7.3 Further information on the relative distributions of parent flubendazole and its metabolites in edible tissues in the chicken after treatment with 60 ppm flubendazole in the feed (the higher recomme inded dose);
 - 7.4 Data for residues in individual animals and for analytical method validation for all pharmacokinetic, metabolism, depletion and distribution studies presented for swine and pharm. oultry;
 - 7.5 Data on quantitative similarities between pharmacokinetic. metabolism, depletion and distribution of flubendazole in chicken and turkey (to allow MRLs to be set for turkey);
 - 7.6 The sensitivity of the analytical methods for the determination of residues should be improved and the methods should be validated for food-producing species (other than pheasants).

flubendazole

> Indications	> Special Precautions	> Mechanism of Action	
> Dosage	> Adverse Drug Reactions	> MIMS Class	
> Contraindications	> Drug Interactions	> ATC Classification	

See related flubendazole information

Abbreviation Index

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Indications	Listed in Dosage.
Dosage	PO Enterobiasis 100 mg as a single dose. Repeat 2-3 wk later if needed. Ascariasis; Hookworm infections; Trichuriasis 100 mg twice daily for 3 days. Click to view flubendazole Dosage by Indications
Contraindications	Pregnancy.
Special Precautions	Lactation. Monitor blood counts and liver function tests regularly during treatment.
Adverse Drug Reactions	GI disturbances; headache, dizziness; allergic reactions; raised liver enzyme values; alopoecia; bone marrow suppression.
Drug Interactions	Enzyme inducers eg phenytoin or carbamazepine; and inhibitors eg cimetidine.
Mechanism of Action	For details of the mechanism of action, pharmacology and pharmacokinetics and toxicology click to view flubendazole
MIMS Class	Anthelmintics
ATC Classification	P02CA05 - Flubendazole ; Belongs to the class of benzimidazole derivative agents. Used as antinematodal.

	FLUBENDAZOLE Barakat Brei market
Brand name:	FLUBENDAZOLE Barakat
Formula:	Tablets 🥝
Categories:	Gastrointestinal System Drugs
Code:	7-20
Composition:	Flubendazole 100mg
Detailed Information:	Composition:
	Each tablet contains: flubendazole IOOmg
	Properties:
	Flubendazol, a benzimidazole carbamate anthelmintic, is an analogue of mebendazole it is
	active against most nematodes and other worms, Flubendazol acts by inhibition or
	destruction of cytoplasmic microtubules in the worm's intestinal or absorptive cells leading
	to death of the worm within several days.
	Indication:
	Flubendazol is used in the treatment of single or mixed infections caused by: Enterobiasis,
	ascariasis, hookworm, and trichuriasis
	Contraindications:
	Hypersensitivity to the drug Pregnancy and Lactation
	Side effects:
	Flubendazol is well tolerated but may cause: transient abdominal pain, diarrhea, headache,
	allergic reactions and raised liver enzyme values with the high doses.
	Precautions:
	function being manifered. No special diet or layotiums
	Decade and administration:
	East the treatment of exterebiasis
	Adults and children: I00mass a single dose repeated if necessary after 2.2 weeks
	Auurts and children. Toomgas a single dose repeated in necessary after 2-3 weeks.
	For ascanasis, nook worm, and inchunasis, roong twice daily for 5 days



Veterinary Drug Residues in Food

Updated up to the 34th Session of the Codex Alimentarius Commission (2011)

VETERINARY DRUG DETAILS

👬 Flubendazole

Functional Class

O Anthelmintic agent

Parch JECFA

Click the above link to access the relevant JECFA residue monograph(s)

Maximum Residue Limits for Flubendazole							
Species	Tissue	MRL	Year of Adoption	Note			
Pig	Muscle	10 µg/kg	1995				
Pig	Liver	10 µg/kg	1995				
Poultry	Liver	500 µg/kg	1995				
Poultry	Muscle	200 µg/kg	1995				
Poultry	Eggs	400 µg/kg	1995				

printer-friendly version

CODEX Veterinary Drugs Home \odot Veterinary Drugs \odot Search \bigcirc Functional Classes Glossary

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EMEA/CVMP/33128/2006-FINAL July 2006

COMMITTEE FOR MEDICINAL PRODUCTS FOR VETERINARY USE

FLUBENDAZOLE (extrapolation to poultry)

SUMMARY REPORT (4)

 Flubendazole is a benzimidazole anthelmintic. It is the fluoro- analogue of mebendazole and has many similar properties. Is administered orally to pigs, chickens and game birds.

Flubendazole is currently entered into Annex I of Council Regulation (EEC) No. 2377/90 for turkey, chicken, game birds and porcine species as follows:

Pharmacologically active substance(s)	Marker residue	Animal species	MRLs	Target tissues	Other provisions
Flubendazole	Sum of flubendazole and (2-amino-1 <i>H</i> - benzimidazole-5-yl) (4-fluorophenyl)- methanone	Chicken, turkey, game birds and porcine	50 μg/kg 50 μg/kg 400 μg/kg 300 μg/kg	Muscle Skin + fat Liver Kidney	
	Flubendazole	Chicken	400 μg/kg	Eggs	

- 2. A request was submitted to the EMEA for the extrapolation of the existing entry in Annex I of Council Regulation (EEC) No. 2377/90 for turkey, chicken and game birds. The scientific justification for this extension was assessed taking into account the Note for Guidance on Risk Analysis Approach for Residues of Veterinary Medicinal Products in Food of Animal Origin (EMEA/CVMP/187/00-FINAL). Based on the approach explained in this guideline the CVMP considered whether the extrapolation to poultry would be possible.
- In setting the ADI in the original assessment of flubendazole, the data summarised in the paragraphs below were considered:
- 4. Flubendazole had low oral bioavailability in the rat, dog and the target species. In rats, the half-life for plasma elimination was around 6 hours. In all species, more than 50% of the administered dose was excreted in the faeces as unchanged flubendazole. The absorbed portion of the drug was rapidly metabolised so that concentrations of the parent drug in the blood and urine were very low. The urine contained a mixture of metabolites. The main metabolic pathways were the same in all the species studied and involved reduction of the ketone functional group and hydrolysis of the carbamate moiety.
- 5. Flubendazole was of low acute oral and subcutaneous toxicity. The acute oral LD50 values were greater than 5000 mg/kg bw in mice, rats and guinea pigs. Acute subcutaneous LD50 values were greater than 5000 mg/kg bw in the rat and the mouse and 4679 and 4834 mg/kg bw in male and female guinea pigs, respectively. The substance was more toxic when administered intraperitoneally with an acute intraperitoneal LD50 of 528 and 434 mg/kg bw in male and female rats, respectively.

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8.8 WHO REPORT 832 VETERINARY DRUG RESIDUES

This report contains the collective views of an international group of experts and does not necessarily represent the decisions or the stated policy of the World Health Organization or of the Food and Agriculture Organization of the United Nations

WHO Technical Report Series

832

EVALUATION OF CERTAIN VETERINARY DRUG RESIDUES IN FOOD

Fortieth report of the Joint FAO/WHO Expert Committee on Food Additives





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v

closantel, with the remaining residue consisting of 3- and 5-monoiodoclosantel. No evidence for the existence of other metabolic pathways was reported. In cattle liver, approximately 10% of the radioactivity was unchanged closantel, and 40–77% was accounted for by 3-monoiodoclosantel. In faeces, 6% of the radioactivity was identified as a sulfate conjugate of a closantel derivative.

Residue depletion of closantel from plasma parallels that from the edible tissues. Within a given species, there is a reasonably constant tissue: plasma ratio which is independent of time. The tissue: plasma ratios in liver and in fat are, respectively, approximately four times and 12 times as large in cattle as in sheep. The tissue: plasma ratios for muscle and kidney in sheep and cattle are comparable.

Maximum Residue Limits

Taking into account this information and the specific residue data discussed on pages 8-10, the Committee recommended amended MRLs for closantel in sheep (Table 2) and new MRLs for cattle (Table 3). The recommendations for cattle are based on the studies in which an oral dose of closantel of 10 mg per kg of body weight and an intramuscular dose of 2.5 mg per kg of body weight were given; from these the Committee calculated that the theoretical maximum daily intake of closantel residues at 42 days withdrawal time would be below the ADI of 1.8 mg for a 60-kg person. It should be noted that the manufacturer recommends an oral dose of 5 mg per kg of body weight for cattle. Residues from such an oral dose will be lower than those given in Table 3.

3.1.2 *Flubendazole*

Flubendazole had not been previously reviewed by the Committee. The compound is used as an anthelminthic in pigs and poultry. It belongs to the group of benzimidazole carbamates.

Toxicological data

A substantial database was available for assessment, including data on kinetics and metabolism, acute toxicity, short-term and long-term toxicity, reproductive and developmental toxicity, and genotoxicity.

The absorption, metabolism, and excretion of flubendazole have been studied using radiolabelled drug. Flubendazole is poorly absorbed and is metabolized in a qualitatively similar way in all species studied. More than 50% of the ingested drug is eliminated unchanged in the faeces. The absorbed drug is rapidly metabolized, so that levels of parent drug in the blood and urine are extremely low. The main site of metabolism is the liver, and major metabolic pathways are carbamate hydrolysis and ketone reduction. It seems probable that flubendazole undergoes enterohepatic circulation.

Single oral doses of flubendazole were slightly toxic to experimental animals, the median lethal dose (LD_{50}) being greater than 5000 mg per kg of body weight in mice, rats, and guinea-pigs.

Table 2 Recommended MRLs for closantel in sheep^a

Tissue	Observed residue	(mg/kg parent drug)	Estimated daily intake	^{b, c} (mg closantel equivalents)	Recommended MRL (µg/kg parent drug)	Theoretical maximum daily intake ^c (mg closantel equivalents)
	Oral dose ^d	Intramuscular dose ^e	Oral dose ^d	Intramuscular dose ^e		
Muscle	< 0.4	1.1	0.12	0.33	1500	0.45
Liver	0.8 (1.14) ^r	0.7 (1.17) ^g	0.11	0.12	1500 (2500) ^g	0.25
Kidney	0.7	1.2	0.04	0.06	5000	0.25
Fat	0.7	0.4	0.04	0.02	2000	0.10
Total			0.31	0.53		1.05

^a Based on concentrations at 28 days withdrawal time. For the original discussion of residue data for sheep, see Annex 1, reference 91.

^b Calculated from the observed residue levels.

^c Based on a daily intake of 0.5 kg of meat made up of 0.3 kg of muscle, 0.1 kg of liver, 0.05 kg of kidney, and 0.05 kg of fat.

^d 10 mg/kg of body weight.

^e 5 mg/kg of body weight.

¹ Estimate of total residues; after oral administration, closantel accounted for 70% of the total residues in liver.

9 Estimate of total residues; after intramuscular administration, closantel accounted for 60% of the total residues in liver.

Tissue	Observed residue	e (mg/kg parent drug)	Estimated daily intake	^{a, b} (mg closantel equivalents)		Theoretical maximum daily intake ^b (mg closantel equivalents)
	Oral dose ^c	Intramuscular dose ^d	Oral dose ^c	Intramuscular dose ^d	Recommended MRL (µg/kg parent drug)	
Muscle	0.19	0.29	0.06	0.09	1 000	0.30
Liver	0.16 (1.6) ^e	0.56 (5.6) ^e	0.16	0.56	1 000 (10 000)⁰	1.00
Kidney	0.83 (1.0) ^f	1.39 (1.7) ^f	0.05	0.09	3000 (3750) ^f	0.19
Fat	0.7 (1.0) ^g	2.36 (3.4) ^g	0.05	0.17	3000 (4290) ^g	0.21
Total			0.32	0.91		1.70

Table 3 Recommended MRLs for closantel in cattle

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^a Calculated from the observed residue levels.

^b Based on a daily intake of 0.5 kg of meat made up of 0.3 kg of muscle, 0.1 kg of liver, 0.05 kg of kidney, and 0.05 kg of fat.

° 10 mg/kg of body weight, 28 days withdrawal time.

^d 2.5 mg/kg of body weight, 42 days withdrawal time.

^e Estimate of total residues; closantel accounted for 10% of the total residues in liver.

^f Estimate of total residues; closantel accounted for 80% of the total residues in kidney.

⁹ Estimate of total residues; closantel accounted for 70% of the total residues in fat.

Flubendazole was given orally in gelatin capsules to dogs at doses of 2.5, 10, or 40 mg per kg of body weight per day, 6 days a week for 3 months. Some atrophic changes and congestion of the epididymis were observed in the male genital tract at doses of 10 and 40 mg per kg of body weight per day, and atrophic changes occurred in the female genital tract at all doses. The changes in the female genital tract were considered to be within normal limits for dogs of the age of those used in the study. On histological examination of male sex organs, changes in the testes could not be clearly associated with flubendazole treatment. The findings in male dogs may not be compound-related, but because of the lack of conclusive evidence as to the cause of these changes, the Committee concluded that the no-observed-effect level (NOEL) was 2.5 mg per kg of body weight per day.

Carcinogenicity studies were performed in mice and rats at doses up to 30 and 20 mg per kg of body weight per day, respectively; no treatment-related effects were observed. There was no treatment-related increase in any type of neoplasm. The Committee was of the opinion that flubendazole had no carcinogenic potential at the highest doses administered in these studies.

The results from a range of *in vitro* and *in vivo* genotoxicity tests were all negative.

The Committee considered data from reproduction, embryotoxicity, and teratogenicity studies. Studies in mice, rabbits, and pigs were negative. Flubendazole was extensively studied in segmented reproduction studies in rats performed as required for human drug regulation purposes and accepted by the Committee in lieu of a multigeneration reproduction study. In several rat developmental studies, doses of up to 40 and 160 mg per kg of body weight per day, given on gestation days 6-15, did not produce any embryotoxic or teratogenic effects. In a rat teratogenicity study published in 1987, using material extracted from a commercial preparation, gross skeletal and internal fetal malformations were recorded at doses of 40 and 160 mg per kg of body weight per day. The NOEL in this study was 10 mg per kg of body weight per day.

An ADI of 0-12 μ g per kg of body weight was established for flubendazole, based on the NOEL of 2.5 mg per kg of body weight per day in the 3-month study in dogs and a safety factor of 200. This safety factor was used by the Committee to take account of the fact that the doses were administered only 6 days per week in this study, the precise consequences of which could not be assessed.

The Committee noted that the ADI also provided a safety margin corresponding to a factor of about 1000 with respect to the NOEL of 10 mg per kg of body weight per day derived from the rat teratogenicity study. Furthermore, the Committee considered that further carcinogenicity studies would not be required, since the highest dose used in the negative studies that it had evaluated exceeded the ADI by a factor of approximately 2000.

Residue data

The Committee considered data on the metabolism of flubendazole and the depletion of flubendazole residues from the edible tissues of pigs, and on the depletion of flubendazole residues from the edible tissues and eggs of laying hens.

When pigs or poultry are treated with flubendazole, the tissue with the highest residue concentration and slowest depletion rate is the liver. The major metabolite in pig liver is (2-amino-1H-benzimidazol-5-yl)-4-fluorophenylmethanone, which is found at a much higher concentration than parent flubendazole. Residue concentrations are higher and more persistent in egg yolk than in egg white.

Pigs. A residue-depletion study was conducted using 18 feeder pigs given a dose of 1.5 mg per kg of body weight of $[^{14}C]$ flubendazole daily for 5 days (Table 4). Total residue concentrations were highest in liver throughout the 30-day withdrawal period.

Three male pigs received flubendazole at 30 mg per kg of body weight in the feed for 5 consecutive days. Flubendazole levels measured by high-performance liquid chromatography (HPLC) were less than 0.01mg/kg (the sensitivity limit of the method) in plasma, liver, kidney, muscle, and fat at withdrawal times of 16, 30, and 54 hours.

A similar residue study was conducted using single oral doses of 5 mg per kg of body weight in three groups of five male pigs. Tissues and plasma were analysed using a radioimmunoassay with quantification limits of 1 µg/kg in plasma and 5 µg/kg in tissues. Animals were slaughtered in groups of five at 24, 72, and 168 hours after dosing. At 24 hours withdrawal time, the tissues contained 7-12 µg/kg of parent flubendazole. All residues were below the detection limit at 72 hours.

In another study, seven sows were treated with 30 mg per kg of body weight flubendazole in the diet for 10 consecutive days. The sows were

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Withdrawal time	Muscle	Liver	Kidney	Fat
6 hours	262	3865	2678	212
5 days	35.5	1863	435	50.1
10 days	10.5	529	78.2	16.3
16 days	8.66	433	76.6	15.6
23 days	8.67	194	49.9	13.5
30 days	2.51	106	22.5	3.38

Table 4 Total residues (μg of flubendazole equivalents per kg) in tissues of pigs given [¹⁴C]flubendazole in the diet at 1.5 mg per kg of body weight for 5 days^a

^a For each withdrawal time, values are means for three pigs.

1	4
	7

Table 5 Total residues (flubendazole equivalents) in tissues and plasma of laying hens given [¹⁴C]flubendazole in the diet at 30 mg per kg of body weight for 6 days^a

Withdrawal time (days)	Plasma (mg/l)	Muscle (mg/kg)	Liver (mg/kg)	Kidney (mg/kg)	Fat (mg/kg)
1	0.007	≤ 0.01	0.210	0.080	≤ 0.01
2	0.005	\leq 0.01	0.146	0.054	\leq 0.01
4	0.002	≤ 0.01	0.069	0.010	\leq 0.01
7	0.001	\leq 0.01	0.073	\leq 0.01	\leq 0.01
11	\leq 0.001	\leq 0.01	0.030	\leq 0.01	\leq 0.01
14	≤ 0.001	\leq 0.01	0.016	\leq 0.01	\leq 0.01

^a For liver and kidney at 1 day withdrawal time, values are means for three animals; all other values are means for four animals.

slaughtered 7 days after the last treatment with flubendazole. Mean levels of flubendazole measured by HPLC were 59, 67, 13, and $33 \mu g/kg$ for liver, kidney, muscle, and fat, respectively.

Poultry. A total of 28 laying hens received [¹⁴C]flubendazole at a dose equivalent to 30 mg per kg of body weight in food for 6 consecutive days. At all withdrawal times tested from 1 to 14 days after treatment, the concentration of radioactive equivalents of flubendazole in blood and plasma was less than $0.01 \,\mu$ g/ml, which suggests that absorption was poor. After total radioactivity levels reached a steady state in 5–6 days, eggs contained an average of $0.12 \,\text{mg}$ of flubendazole equivalents per kg. Radioactivity in the yolks ($0.34 \,\text{mg/kg}$) was much higher than in the egg white ($0.02 \,\text{mg/kg}$). The highest observed levels of radioactivity in individual tissues, calculated in terms of flubendazole equivalents, were $0.21 \,\text{mg/kg}$ in liver and $0.08 \,\text{mg/kg}$ in kidney 24 hours after the last dose. Table 5 shows residues in plasma and tissue for various withdrawal times.

When chickens were treated with flubendazole at 60 mg per kg of body weight for 7 days, residues were detectable in egg yolk for 11 days after treatment ended. Residue levels were higher in yolk than in white. Eggs and tissues were analysed by an HPLC method sensitive to 0.01 mg/kg. Of the tissues, liver had the greatest amount of residue at zero withdrawal time, although flubendazole could not be detected in any tissue by 6 and 7 days withdrawal time. The residue data are summarized in Table 6.

Methods of analysis for residues in tissues

For the studies described on pages 14-15, plasma and tissue levels of flubendazole in pigs were measured by radioimmunoassay or by an HPLC method with ultraviolet detection at 313 or 254 nm, which is sensitive to 0.01 or 0.02 mg/kg, respectively. Another HPLC method has been developed for flubendazole, with ultraviolet detection at 254 nm, that gives excellent separation between flubendazole and the major metabolite

flubendazole in the diet at 60 mg per kg of body weight for 7 days					
Withdrawal time (days)	Egg yolk	Egg white	Muscle	Kidney	Liver
0	0.592	0.036	0.079	0.173	0.198
4	NM	NM	0.071	0.236	0.200
6	NM	NM	\leq 0.01	\leq 0.01	\leq 0.01
7	0.318	\leq 0.01	\leq 0.01	\leq 0.01	\leq 0.01
11	0.019	< 0.01	NM	NM	NM

Table 6 of objekene aiver

 ≤ 0.01 ΝM

NM: not measured

11

28

resulting from carbamate hydrolysis. However, the method described applies to the analysis of pure substances and does not include extraction procedures for tissues.

NM

 ≤ 0.01

NM

 ≤ 0.01

 ≤ 0.01

An HPLC method that has detection limits of 20-50 µg/kg has been developed for simultaneously determining eight benzimidazoles in tissue. This method might be suitable for measuring flubendazole and the major metabolite found in pig tissue, (2-amino-1H-benzimidazol-5-yl)-4-fluorophenylmethanone. Typical recoveries from spiked samples (0.1 mg/kg) were above 70% for flubendazole in liver, kidney, and muscle.

Maximum Residue Limits

0.019

ΝM

In reaching its decision on MRLs, the Committee took into account the following points:

- An ADI of 0-12 µg per kg of body weight was established. This would result in a maximum ADI of $720 \,\mu g$ for a 60-kg person.
- The marker residue is the parent drug for all tissues and for eggs.
- The total daily intake of flubendazole-related residues in food would be about 620 µg (see Table 7), if assumed to be accounted for by pig tissue and eggs at zero withdrawal time, and calculated on the basis of the data presented in Table 4 and the study in chickens treated with a dose of 30 mg per kg of body weight.

Eggs. The daily intake of flubendazole-related residues will probably remain below the ADI even when flubendazole is given at 60 mg per kg of body weight, although this dose produces a much higher concentration of residues in eggs. The argument that increased doses of flubendazole will not increase residue levels because of the drug's low systemic availability appears not to be valid for eggs: the levels of parent flubendazole in egg yolk found in the study with 60 mg per kg of body weight are double the residue levels of all flubendazole-related residues found in the study with 30 mg per kg of body weight.

An MRL for whole egg of $400 \,\mu g/kg$ of parent flubendazole is recommended.

Table 7

Estimated total daily intake of flubendazole-related residues in food

Tissue	Observed residue (mg/kg parent drug equivalents)	Estimated daily intake ^a (µg parent drug equivalents)
Pig tissue		
Muscle	0.262 ^b	79
Liver	3.865 ^b	386
Kidney	2.678 ^b	134
Fat	0.212 ^b	11
Eggs	0.12 °	12
Total		622

^a Calculated from the observed residue levels. Based on a daily intake of 0.5 kg of meat (made up of 0.3 kg of muscle, 0.1 kg of liver, 0.05 kg of kidney, and 0.05 kg of fat) and 0.1 kg of eggs.

^b Based on concentrations at 6 hours withdrawal time in a study in which pigs received an oral dose of [¹⁴C]flubendazole at 1.5 mg per kg of body weight daily for 5 days.

^c Average concentration detected in eggs on day 6 of a study in which laying hens received [¹⁴C]flubendazole in the diet at 30 mg per kg of body weight for 7 days.

Poultry. As no withdrawal period is required for poultry, parent flubendazole is an adequate marker residue. MRLs of 500 and 200 μ g/kg are recommended for parent flubendazole in poultry liver and muscle, respectively.

Pigs. Although edible tissues from pigs require no withdrawal period from a human food safety perspective, a withdrawal period based on good practice in the use of veterinary drugs has been applied.

Parent flubendazole is the only analyte available as the marker residue for pig liver. Methods are available for determining flubendazole, and the residue data indicate that misuse can be detected by monitoring for parent flubendazole in pig tissue.

An MRL of $10 \,\mu$ g/kg is recommended for the parent compound in pig liver and muscle.

3.1.3 Ivermectin

Ivermectin (a mixture of $\geq 80\% 22,23$ -dihydroavermectin B_{1a} (H_2B_{1a}) and $\leq 20\% 22,23$ -dihydroavermectin B_{1b} (H_2B_{1b})) had previously been evaluated at the thirty-sixth meeting of the Committee (Annex 1, reference 91), when an ADI of 0–0.2 µg per kg of body weight was established, based on a NOEL of 0.1 mg per kg of body weight per day for maternal toxicity in the CF₁ mouse and a safety factor of 500.

Toxicological data

The Committee reappraised the developmental toxicity of ivermectin and