

# **Transgenic Construct Submission Form**

PI: \_\_\_\_\_

Construct name (short w/ no unusual characters): \_\_\_\_\_

Contact person: \_\_\_\_\_

Contact person info: Phone #1: \_\_\_\_\_ Phone #2: \_\_\_\_\_

FAX: \_\_\_\_\_ pager: \_\_\_\_\_

Email: \_\_\_\_\_

Date Submitted: \_\_\_\_\_

**SERVICE DESCRIPTION:** Transgenes will be injected into the pronuclei of fertilized, viable murine oocytes isolated from a set of 10-15 females (depending on strain requested for each session). Mouse strains available for injection are C57BL/6J (inbred), or C57Bl/6xCBA (hybrid). Other strains can be used for an increased fee. DNA from each resulting mouse will be screened by PCR to detect transgenic founder animals. To date, we have averaged 10% transgenic founder animals per live births, with significant variation between constructs. Before injection, you will need to provide some information about the transgene, a genotyping assay protocol, and some administrative details. Contact Mia Wallace after completing the form. Mia is at 314.747.4554, or [mia@wustl.edu](mailto:mia@wustl.edu). We will arrange a meeting with you to turn over the DNA and PCR primers, and to review the construct information. All facility services are performed in the order received and kept confidential.

Please provide the information requested below. If you have any special circumstances or requirements (e.g. a detrimental phenotype or requirement for a different mouse strain), contact us to make arrangements. We are happy to provide specialized services.

Which strain of mice for injections?

FVB/N (inbred) \_\_\_\_\_

C57Bl/6xCBA (hybrid) \_\_\_\_\_

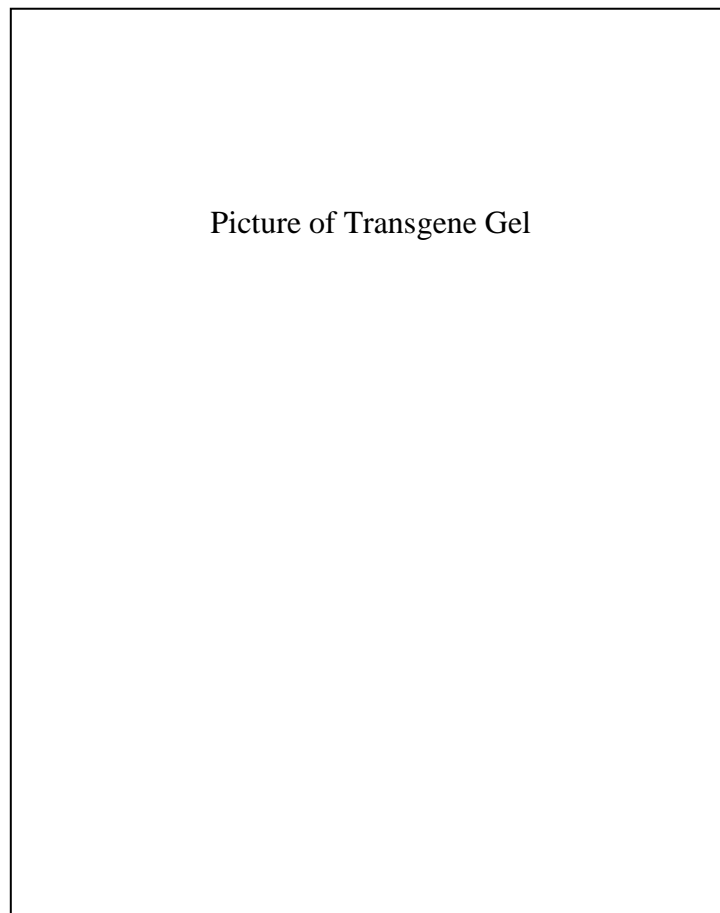
C57Bl6/NJ (inbred) \_\_\_\_\_

**BACKGROUND:** A brief description of the scientific rationale for producing the construct comprehensible to those outside the immediate field, including the expected sites of expression and phenotype.

**GENE STRUCTURE:** Diagram the transgene construct and label salient features, including the promoter, reporter, stop and start sites for transcription, stop and start sites for translation, intron/exon boundaries, and size of each sequence.



**CONSTRUCT PREPARATION PROTOCOL:** The construct should be prepared for injection using the following protocol: <http://mgc.wustl.edu/Protocols.aspx#xgene>. If you prefer a different method, please contact [mia@wustl.edu](mailto:mia@wustl.edu) in advance. BAC constructs should be supplied in high salt buffer. Please contact [mia@wustl.edu](mailto:mia@wustl.edu) in advance of all BAC submissions for information on construct preparation. The construct should be freed from vector sequences with a restriction digest resulting in fragments that are readily separated by agarose gel electrophoresis. Digest enough material to yield 1µg of construct after purification (yields are typically 25-50%). Please submit your DNA sample in TE or injection buffer (10mM TRIS, pH=7.4, 0.1mM EDTA). We will need at least 500ng of DNA for injection, and this should be provided in a tube labeled with the construct name and DNA concentration. The construct name **should be short, alphanumeric, and not contain Greek or other unusual characters, subscripts or superscripts.**



**GENOTYPING:** Before injecting the transgene, you will need a set of PCR primers suitable for genotyping the resulting animals. We use a PCR genotyping protocol that works for all transgenes [Stratman and Simon (*Transgenic Res.* 12, 521-522 (2003))]. This protocol requires primers that are 30 nucleotides in length, and we have proven primer sets for many transgenes (please see [Services / PCR page](#)). If your transgene contains any of these sequences, you do not need to provide PCR primers. If your transgene does not contain these sequences, you should design primers as indicated on the [Protocols page](#). All primers are 30-mers with ~50% GC content and produce an amplicon of 100-400 base pairs. We will pick the primers if you provide us with the transgene sequence. If you provide your own primers you will need at least 50µL of a 100µM solution for each 30-mer. Please indicate either in-house primers or transgene specific primer sequences below.

Proven MGC Primer Set (please see Primer Pairs): \_\_\_\_\_

Primers Provided (please give primer names): \_\_\_\_\_ amplicon size

#1 \_\_\_\_\_

Sequence: \_\_\_\_\_

#2 \_\_\_\_\_

Sequence: \_\_\_\_\_

## BILLING INFORMATION

PI: \_\_\_\_\_

Department/Division & Dept. # \_\_\_\_\_

PI signature: \_\_\_\_\_

Bill to fund (number)\*: \_\_\_\_\_

Accounting contact (name): \_\_\_\_\_

\* Investigators who expect to receive a subsidy from dedicated Core grants, please check the appropriate box below and fill out the required additional forms. The additional forms for Digestive Diseases Research Core Center ([DDRCC](#)), Diabetes Research Center ([DRC](#)), the Children's Discovery Institute ([CDI](#)), and the WashU Musculoskeletal Research Center ([MRC](#)) investigators can be found on the respective websites. Approval of the project by the Core Director is required for subsidy. Subsidy cannot be guaranteed without approval BEFORE the service is performed.

DDRCC

DRC

CDI

**ANIMAL TRANSFER:** We will automatically transfer the transgenic founder animals to you at weaning age unless you instruct us otherwise. Before injection, we will need an Animal Studies Committee protocol number for your project, and a location to transfer the mice. The ASC protocol need only be for the analysis of the animals - the Core Facility has ASC approval for the procedures used to generate the animals. At the time of transfer, we will notify you by email to expect the animals in your barrier room. Please allow approximately 2 weeks after weaning for animals to be moved; we will not transfer any mice until their health screen has come back negative for pathogens.

ASC number: \_\_\_\_\_ Expiration Date: \_\_\_\_\_

Transfer animals to building: \_\_\_\_\_ Room #: \_\_\_\_\_