

# ***Using Bone Imaging and Mechanical Testing for Skeletal Phenotyping***

WU Musculoskeletal Research Center  
Musculoskeletal Structure & Strength Core

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July 13, 2018



# ???

- I am studying a genetic mutation. Does my mouse have a skeletal phenotype?
- I treated rats with drug XYZ. What was the effect on their bones?
  - any evidence of increased bone formation?
  - ... increased resorption?
  - are the bones stronger?
- What outcomes do I need to measure?
- How do I integrate microCT and mechanical testing outcomes?

# *Structure & Strength*

- Imaging → Structure
  - how much bone is there? how is it distributed?
  - density, morphology (geometry)
  - whole-bone, cortical, cancellous
- Mechanical Testing → Strength
  - stiffness, strength, toughness
  - length scale
    - large: ‘structural’ (whole-bone, organ)
    - small: ‘material’ (tissue, e.g., cortical bone)

# Imaging → Structure

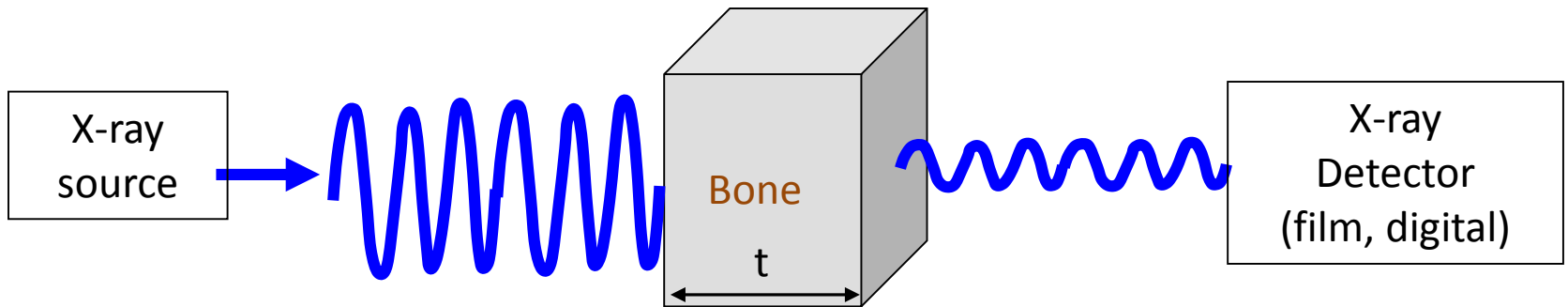
# *X-Ray Based Imaging*



<https://theconversation.com/curious-kids-is-x-ray-vision-possible-90393>

# *X-Ray Based Imaging*

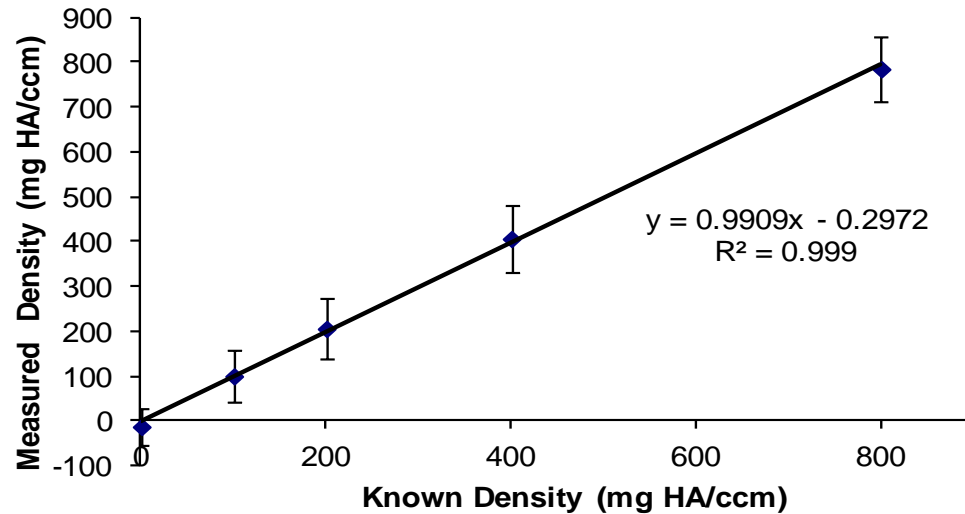
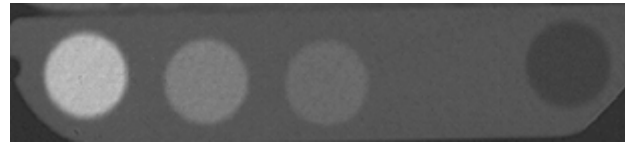
- Based on x-ray absorption/attenuation



- Attenuation depends on thickness ( $t$ ), mineral density

# *X-Ray Based Imaging*

- Can calibrate x-ray absorption to equivalent density of hydroxyapatite (HA) phantom



MicroCT density  
calibration

# Radiography

- Outcomes: qualitative phenotype, spontaneous fractures, fracture callus



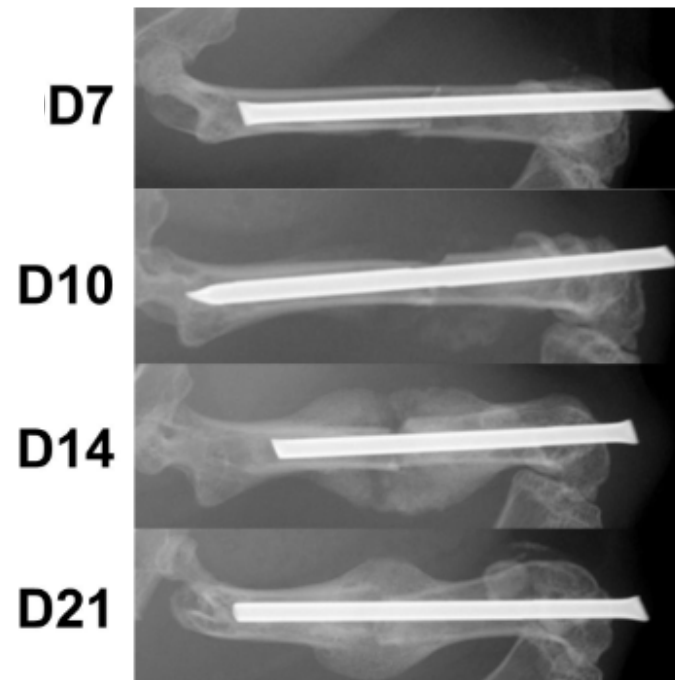
Faxitron

*Mouse hindlimbs*



*Karuppaiah, Ornitz et al (2016)*

*Mouse femur fracture*



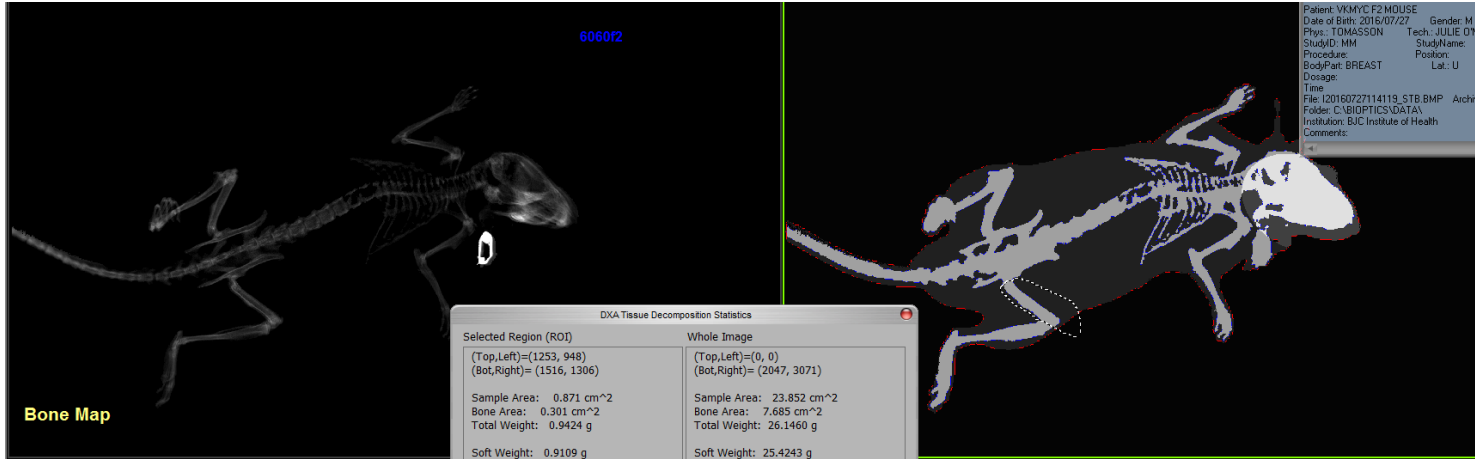
*Liu, McKenzie, Silva et al. (2017)*



# DXA, Dual-Energy X-ray Absorptiometry



Faxitron



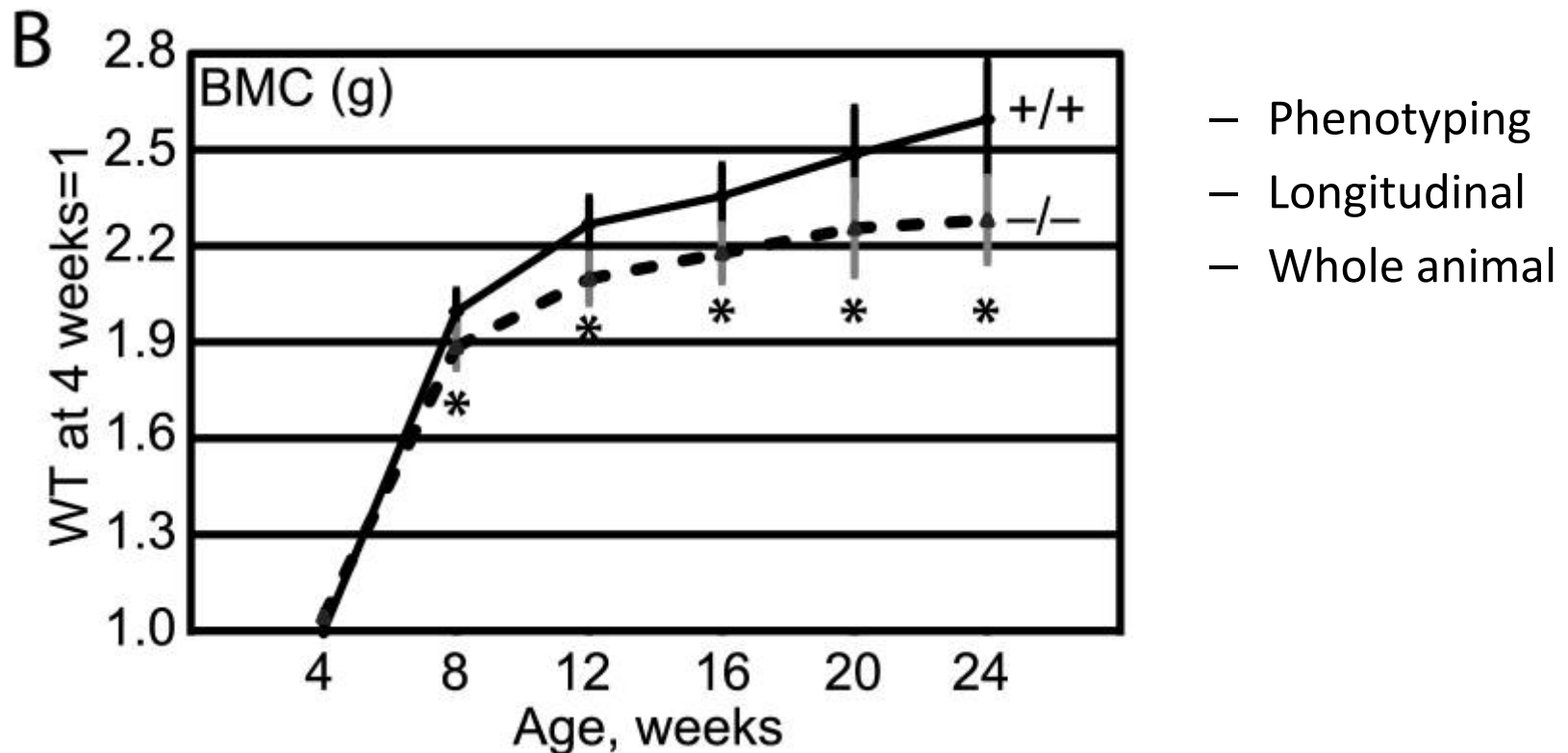
## Outcomes:

- BMC (g), aBMD (g/cm<sup>2</sup>), %fat (caveat)

## Uses:

- Whole animal
- Phenotyping
- Systemic intervention
- Longitudinal studies

*DXA shows decreased post-natal accrual of bone mass (BMC) in MAGP-deficient mice*



*Craft, Mecham et al., 2010*

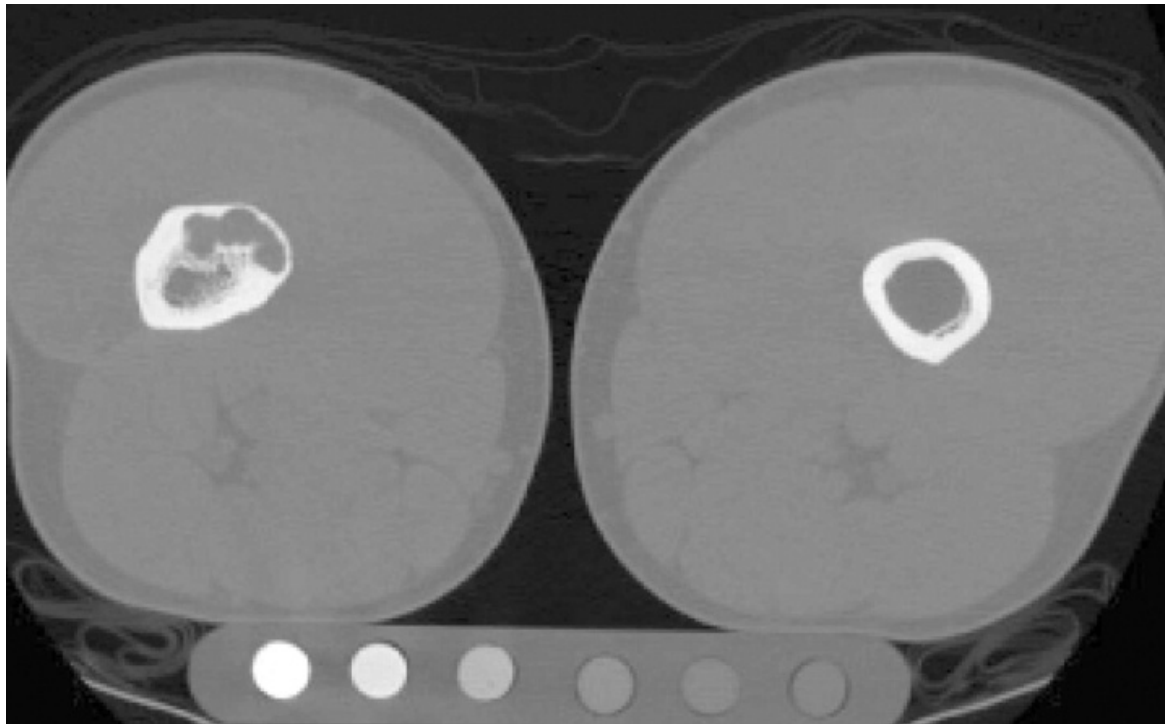
# X-Ray Computed Tomography (CT)



Cat Scan

# *QCT – Quantitative Computed Tomography*

- grayscale image reflects local attenuation, i.e., “mineral density map” [whiter = denser]
- morphology & density



## Guidelines for Assessment of Bone Microstructure in Rodents Using Micro-Computed Tomography

Mary L Bouxsein,<sup>1</sup> Stephen K Boyd,<sup>2</sup> Blaine A Christiansen,<sup>1</sup> Robert E Guldberg,<sup>3</sup> Karl J Jepsen,<sup>4</sup> and Ralph Müller<sup>5</sup>

Journal of Bone and Mineral Research, Vol. 25, No. 7, July 2010, pp 1468–1486



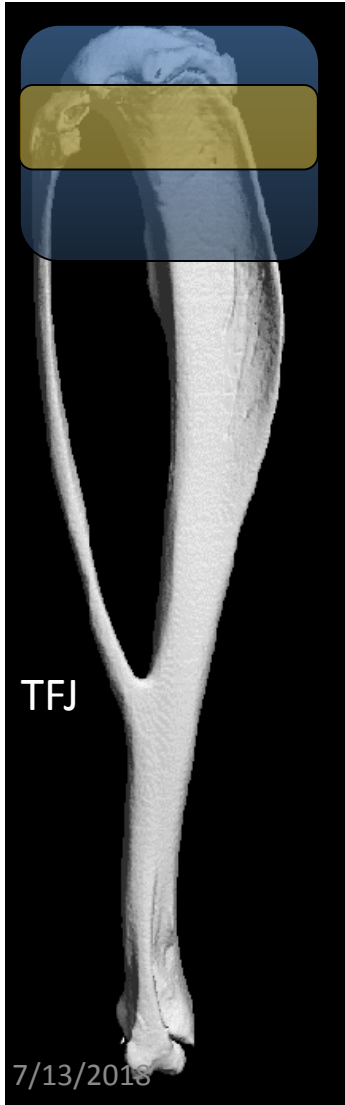
*Specimens ('microCT')*



*Specimens or  
Animals  
(‘VivaCT’)*

# microCT – regions of interest

Mouse tibia



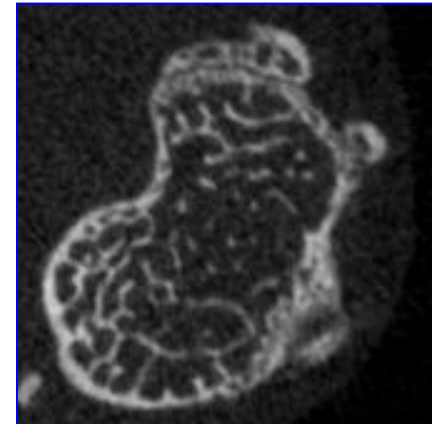
## Cancellous/Trabecular/Metaphyseal

### Scan region

- e.g., 300 slices = 4.8 mm\*

### Analysis region

- landmark (eg, growth plate)
- e.g., 100 slices = 1.6 mm\*



\*assumes 16 um voxel size

# *microCT – regions of interest*



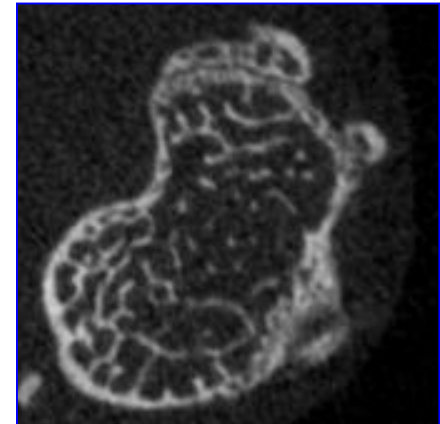
## Cancellous/Trabecular/Metaphyseal

### Scan region

- e.g., 300 slices = 4.8 mm\*

### Analysis region

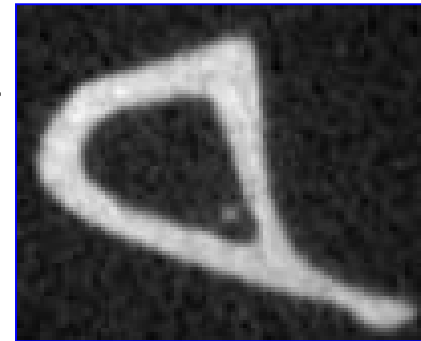
- landmark (eg, growth plate)
- e.g., 100 slices = 1.6 mm\*



## Cortical/Diaphyseal

### Scan region = Analysis region

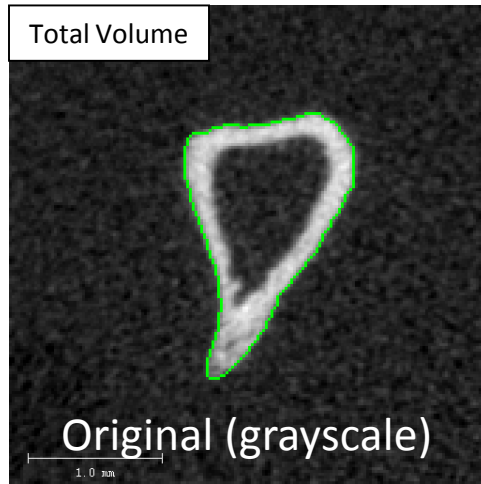
- Mid-point (50% of length), or X mm from TFJ
- e.g., 50 slices = 0.8 mm\*



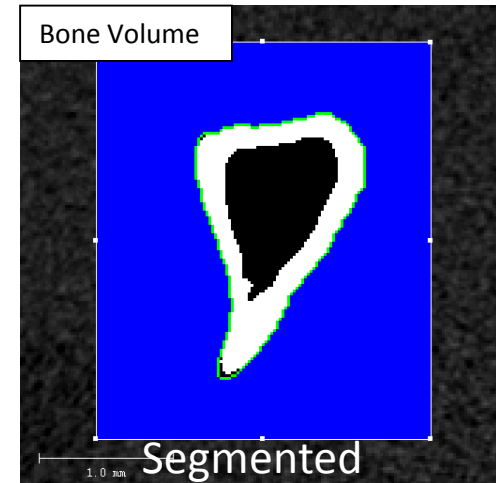
\*assumes 16 um voxel size

# *microCT – cortical analysis*

Contour



Threshold/  
Segment



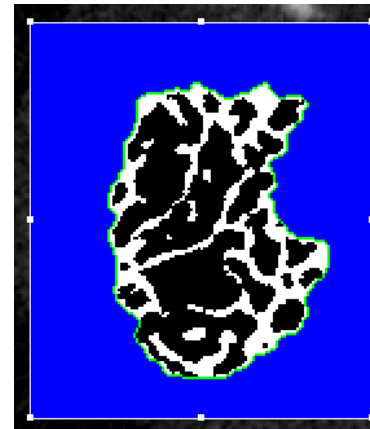
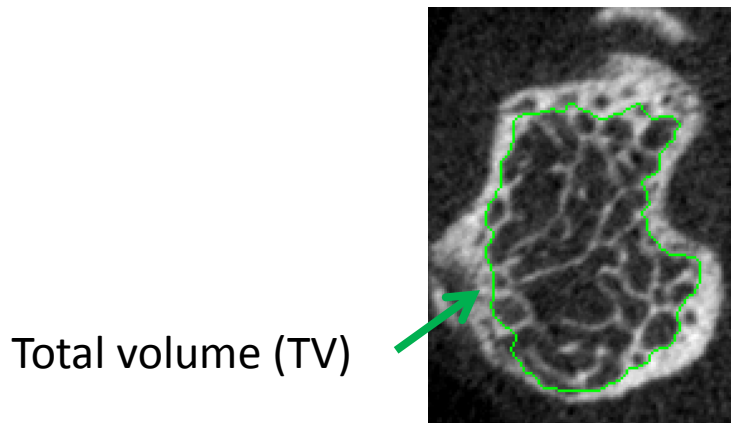
Binary: white=bone  
black=not bone

## Cortical Outcomes

- Tt.Ar, Total area (or volume) – How big is the bone? → periosteal apposition
- Ma.Ar, Marrow area → resorption
- Ct.Ar, Cortical bone area – How much bone is there? → formation & resorption
- Ct.Th, Cortical thickness → formation & resorption
- J, Area moment of inertia → How is bone distributed? Resists bending loads.
- TMD, Tissue mineral density (“mean2”) → ~mineralization



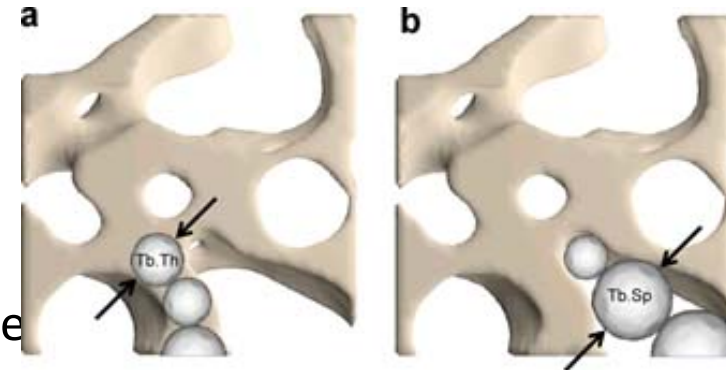
# *microCT – cancellous analysis*



Bone volume (BV) = white

- Cancellous Outcomes\*

- Bone volume fraction (BV/TV) – What fraction of marrow cavity is filled with bone?
- Trabecular thickness (Tb.Th)
- Separation (Tb.Sp)
- Trabecular Number (Tb.N)
- Density, vBMD (“mean1”) – Should correlate

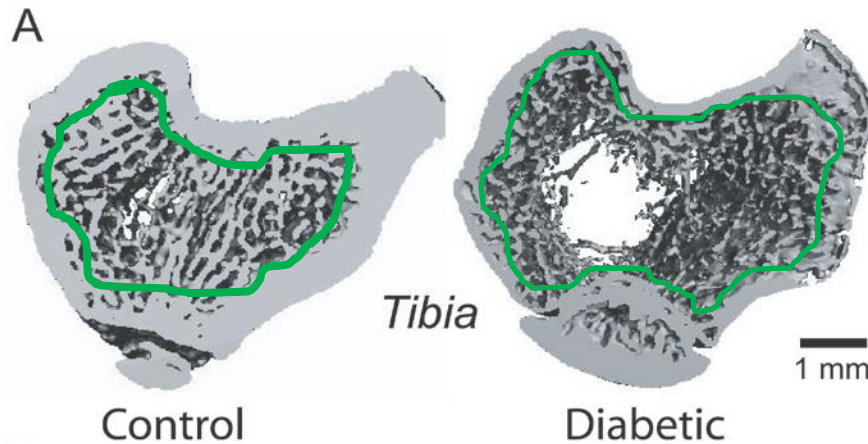


*Bouxsein et al. 2010*

\* Use Direct Method values; prefix “VOX”; do not use Plate Model values; prefix “TRI”

17

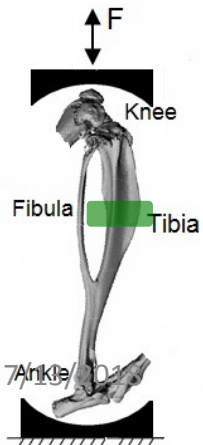
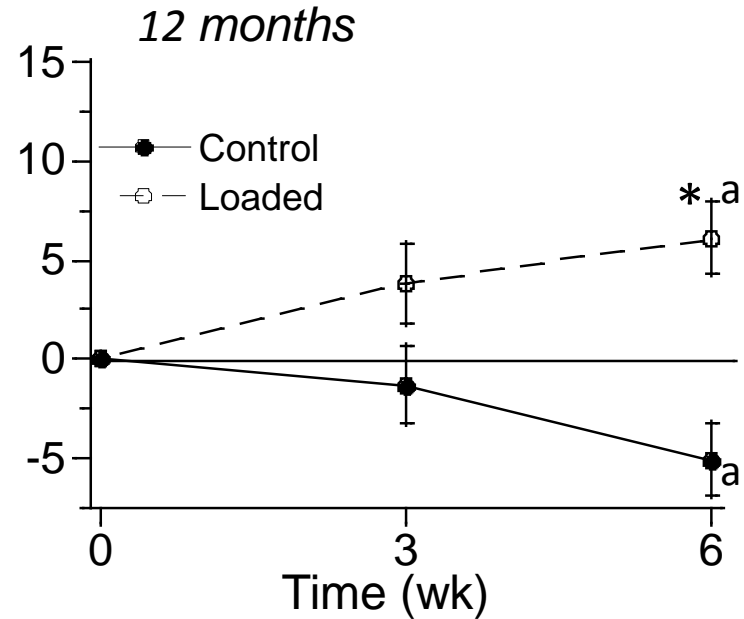
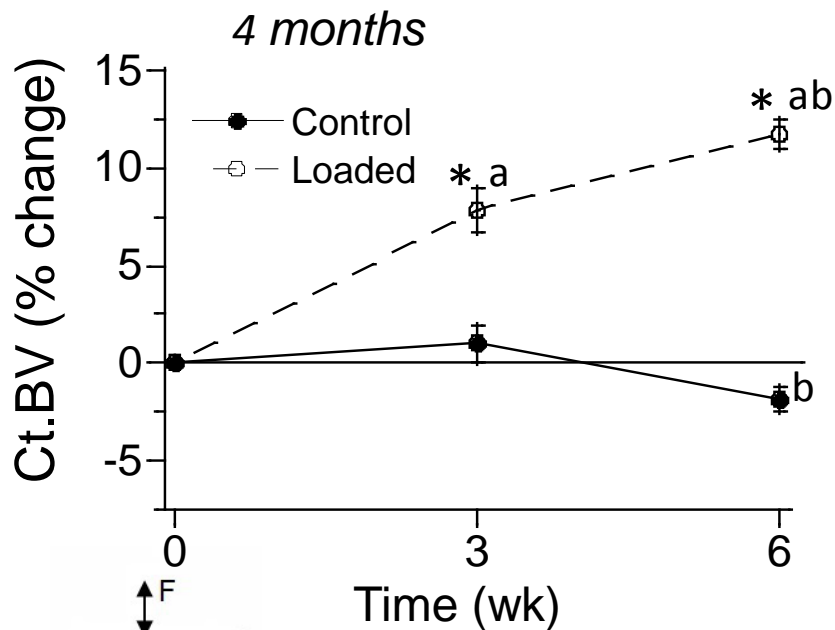
# Specimen microCT reveals osteopenia in T1D diabetic rats



	Control (n = 9)	Diabetic (n = 8)
Parameter		
Tb.BV/TV (mm <sup>3</sup> /mm <sup>3</sup> )	0.37 ± 0.04	0.12 * ± 0.04
Tb.Th (mm)	0.11 ± 0.01	0.07 * ± 0.01
Tb.N (1/mm)	4.4 ± 0.2	2.6 * ± 0.5

*Silva et al. 2009*

# In Vivo CT: Mechanical Loading Stimulates Cortical Bone Accrual (in Age-Dependent Manner)

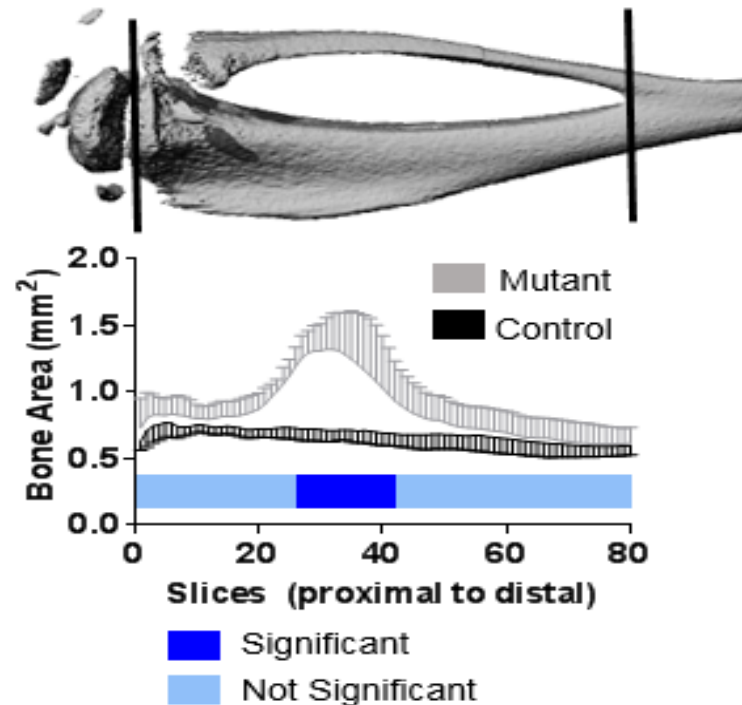


BALB/c  
female mice

\*  $p < 0.05$ , Control vs. Loaded  
a,b:  $p < 0.05$  vs. 0, 3 wks

*Silva et al., 2012*

# New Method: Comparing morphology along bone length



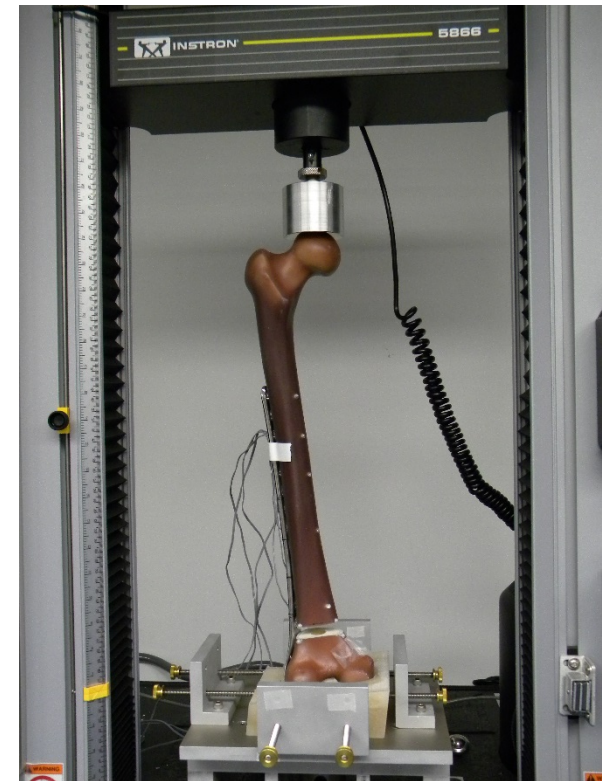
Jenny McKenzie,  
Dan Leib, Dave Ornitz

**Figure 5 – Prelim Data.** Bone area analysis along the length of the tibia revealed a spatially-dependent difference between mutant and control mice. Curves depict mean  $\pm$  SD from n=3-5/group. Statistical significance ( $p < 0.05$ ) is depicted in color bar, and was assessed by t-test with multiple comparison correction.

# Mechanical Testing → Strength

# *How to measure bone mechanical properties?*

- Anderson “The Spider” Silva measures bone strength
- MRC Core measures bone strength



[www.mmamania.com](http://www.mmamania.com)

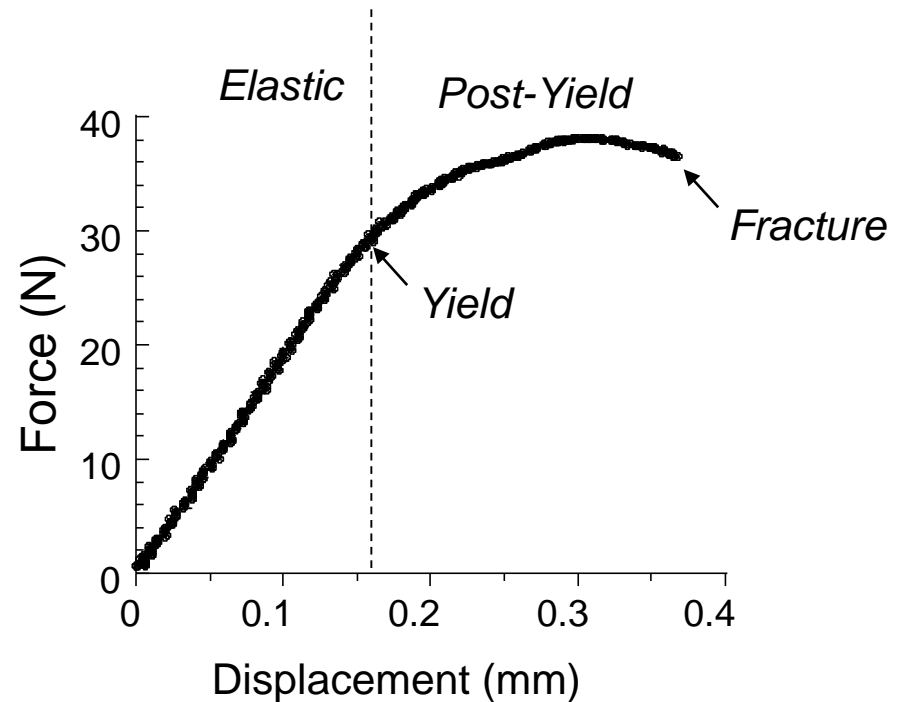
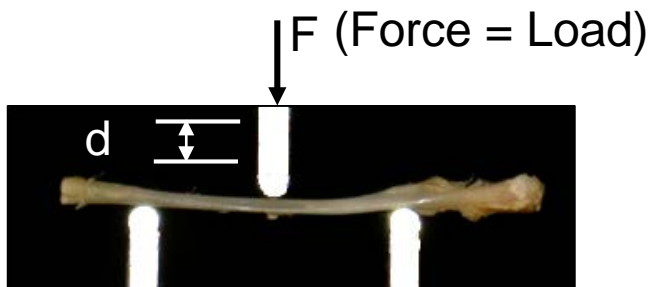
Instron mechanical testing machine

# **Establishing Biomechanical Mechanisms in Mouse Models: Practical Guidelines for Systematically Evaluating Phenotypic Changes in the Diaphyses of Long Bones**

Karl J Jepsen,<sup>1</sup> Matthew J Silva,<sup>2</sup> Deepak Vashishth,<sup>3</sup> X Edward Guo,<sup>4</sup> and Marjolein CH van der Meulen<sup>5</sup>  
Journal of Bone and Mineral Research, Vol. 30, No. 6, June 2015, pp 951–966

# Mechanical Testing: Force-Displacement Plot

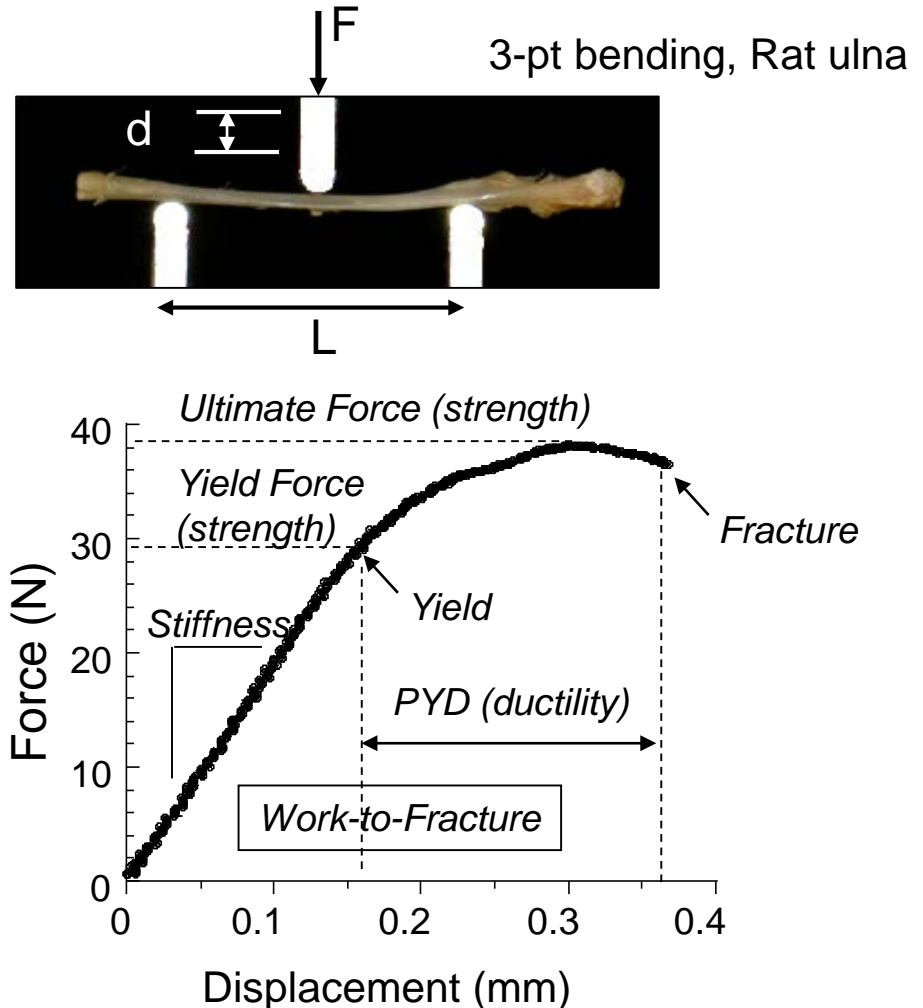
3-pt bending, Rat ulna



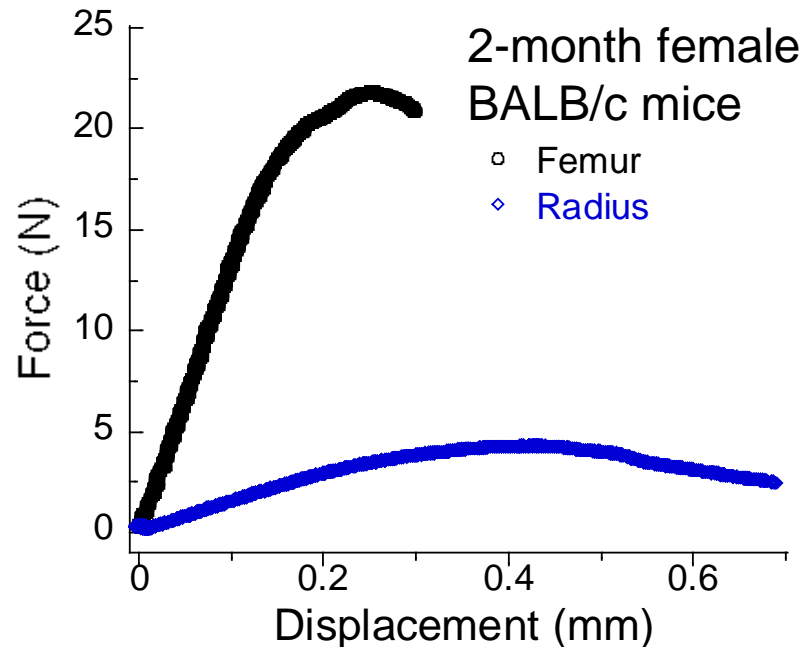


# Mechanical Properties

- Stiffness (N/mm)
- Strength
  - Yield Force (N)
  - Ultimate (Max.) Force (N)
- Post-yield displacement (ductility) (mm)
- Work-to-fracture (N\*mm)



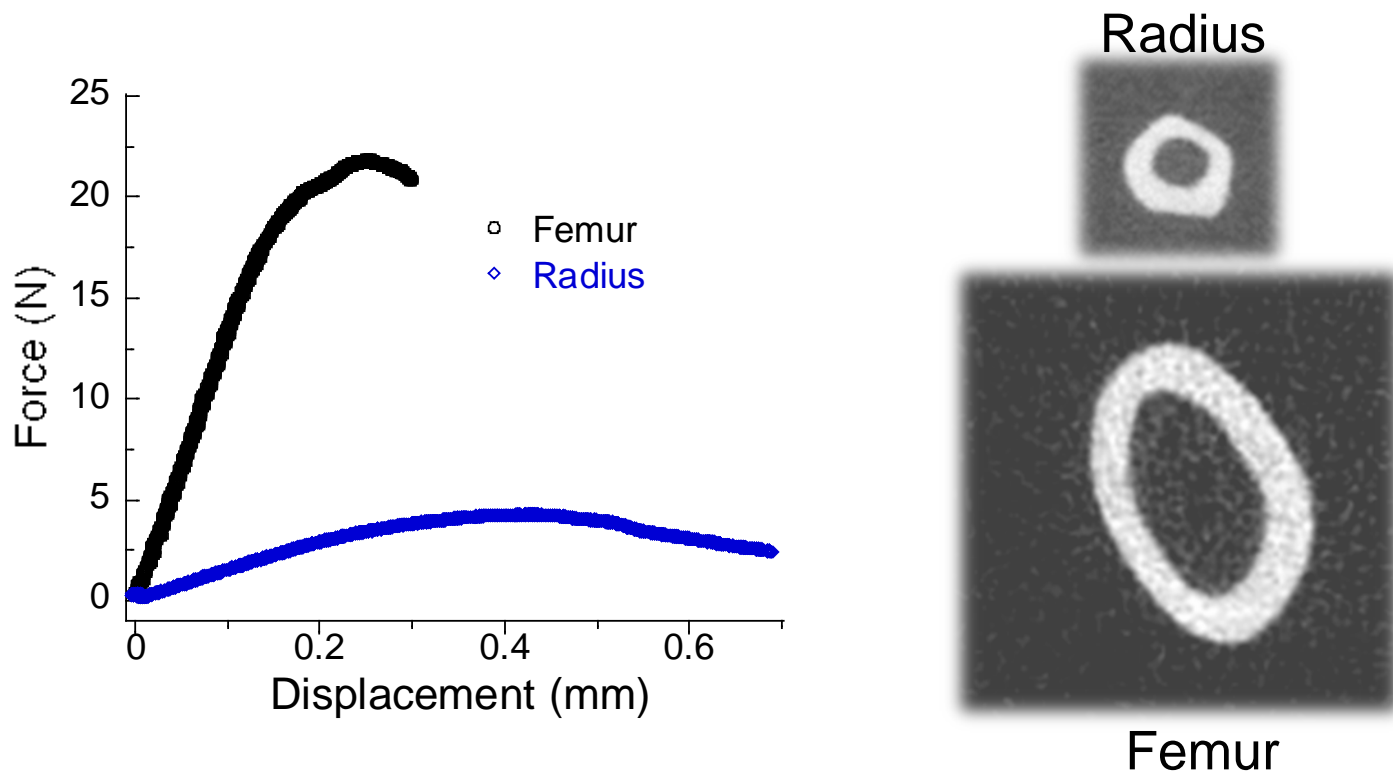
# *Mechanical properties*



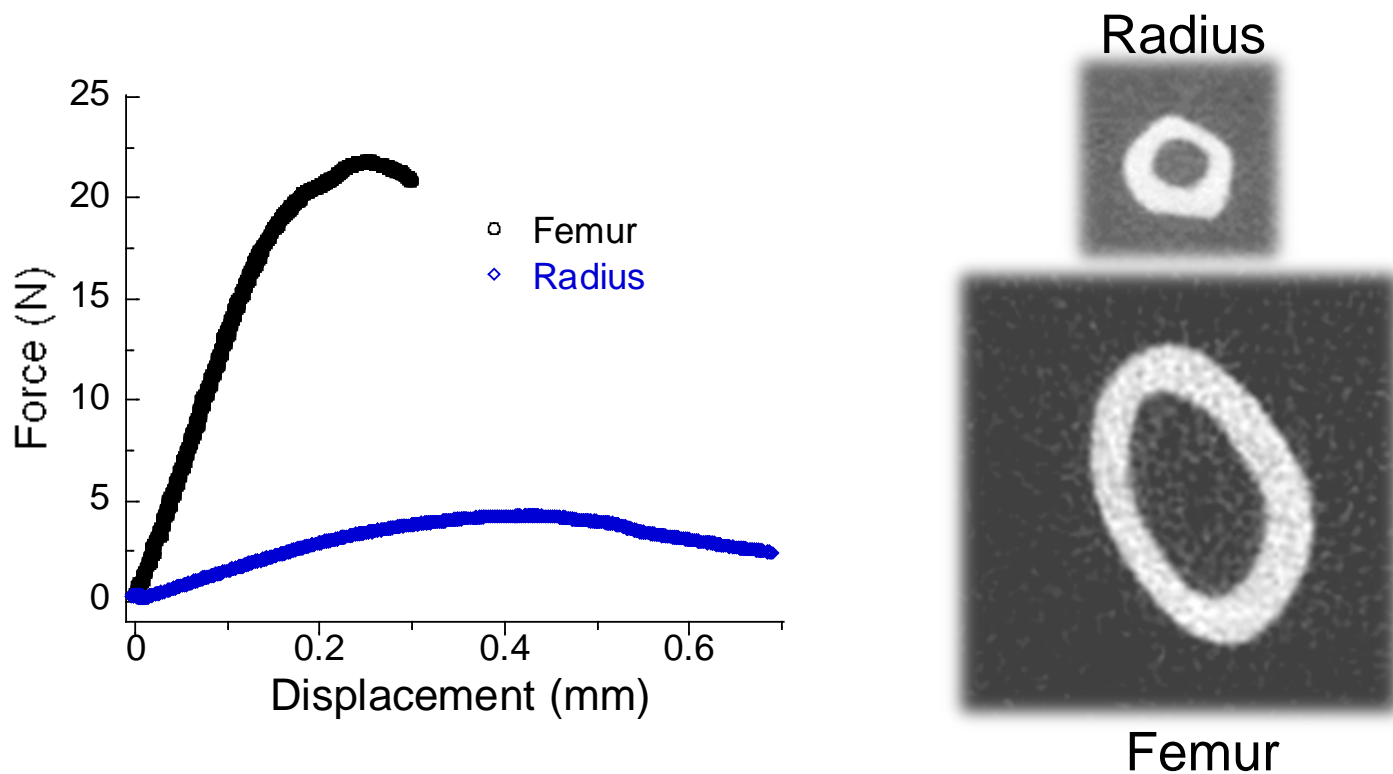
## Femur vs. Radius:

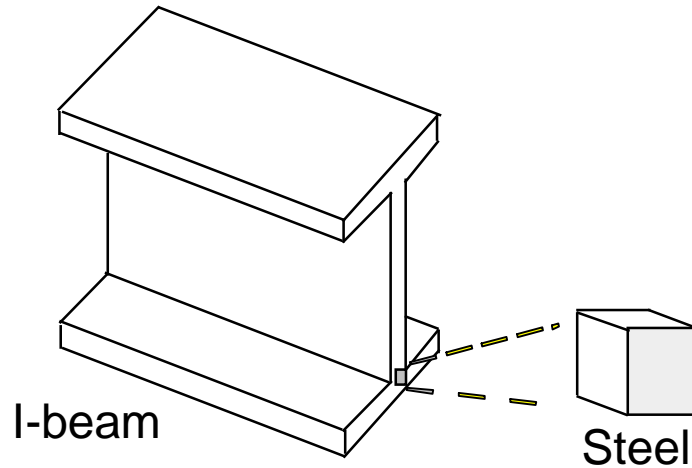
- stiffer
- stronger
- less ductile
- requires more energy to fracture

# *Why is femur stiffer and stronger than radius?*



# *Whole-bone properties depend on morphology & material*



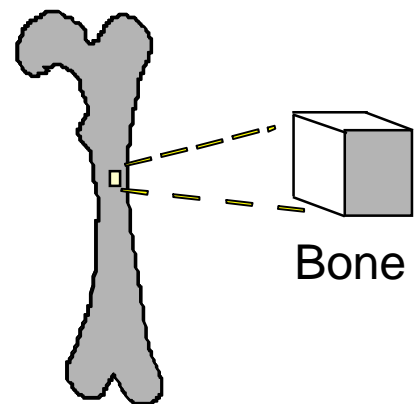


## Larger:

- Structure
- Whole-Bone

## Smaller:

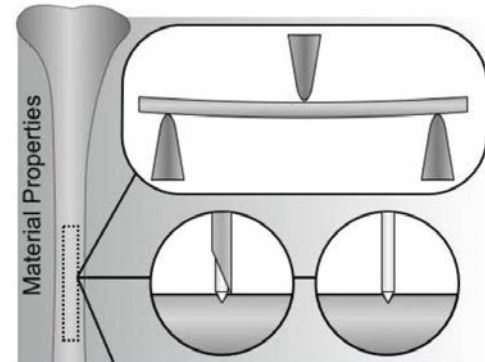
- Material
- Tissue



Femur  
(whole-bone)

# *Material Properties*

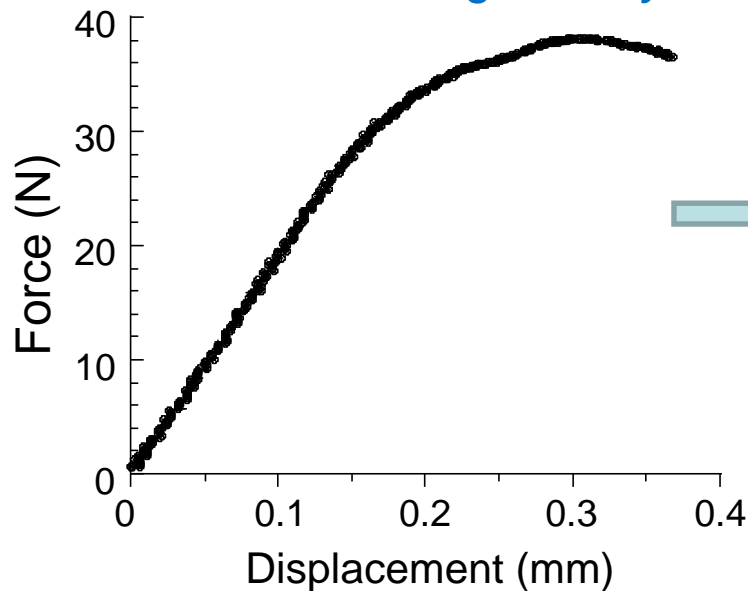
- Estimate from whole-bone mechanical test and bone size (microCT)
  - Whole-bone strength: Ultimate (Max.) Force (N)
  - Material strength: Ultimate Stress ( $\text{N}/\text{mm}^2$ )
- Measure with tissue-level test
  - Bending a small plank of bone
  - Microindentation



# Estimating Material Properties

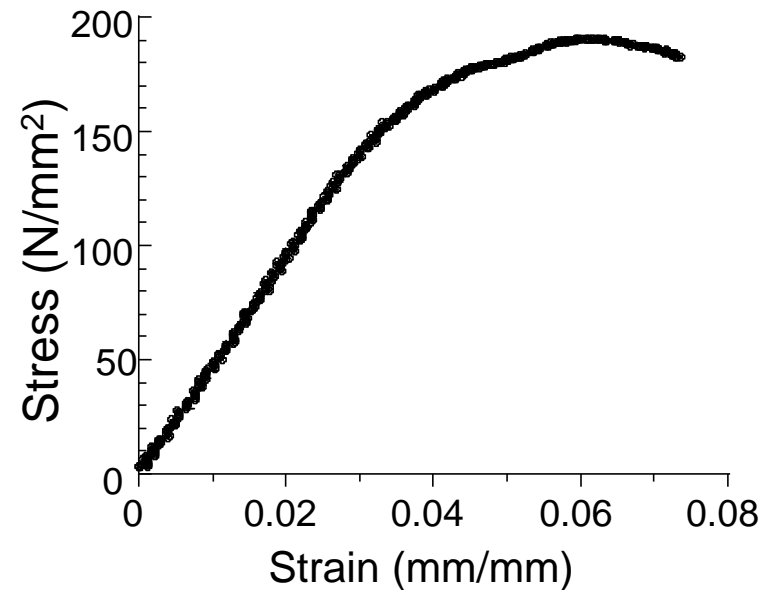
Measured Whole-bone behavior

*Depends on bone size, material  
and test geometry*



Estimated Material behavior

*Depends only on material*



**Table 1.** Terminology Used for Whole-Bone and Tissue-Level Mechanical Properties

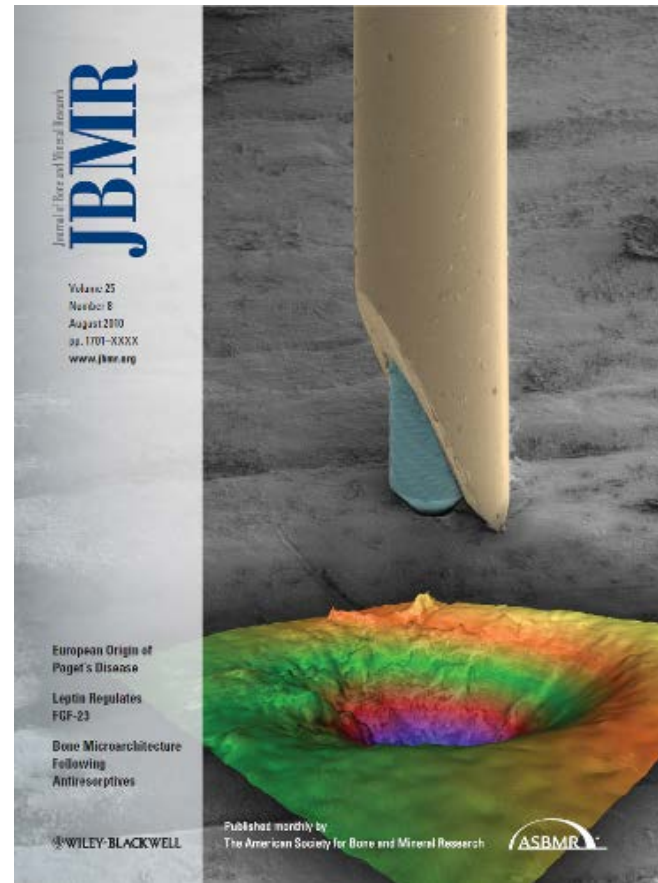
Whole-bone mechanical properties [units]	Tissue-level mechanical properties [units]
Stiffness [N/mm]	Elastic modulus (or tissue-level stiffness) [ $\text{N}/\text{mm}^2 = \text{MPa}$ ]
Maximum load (or whole-bone strength) <sup>a</sup> [N]	Ultimate stress (or tissue-level strength) [ $\text{N}/\text{mm}^2 = \text{MPa}$ ]
Postyield displacement [mm]	Postyield strain [mm/mm (a dimensionless ratio)]
Work-to-fracture [Nmm]	Toughness (or modulus of toughness) [ $\text{N}/\text{mm}^2 = \text{MPa}$ ]

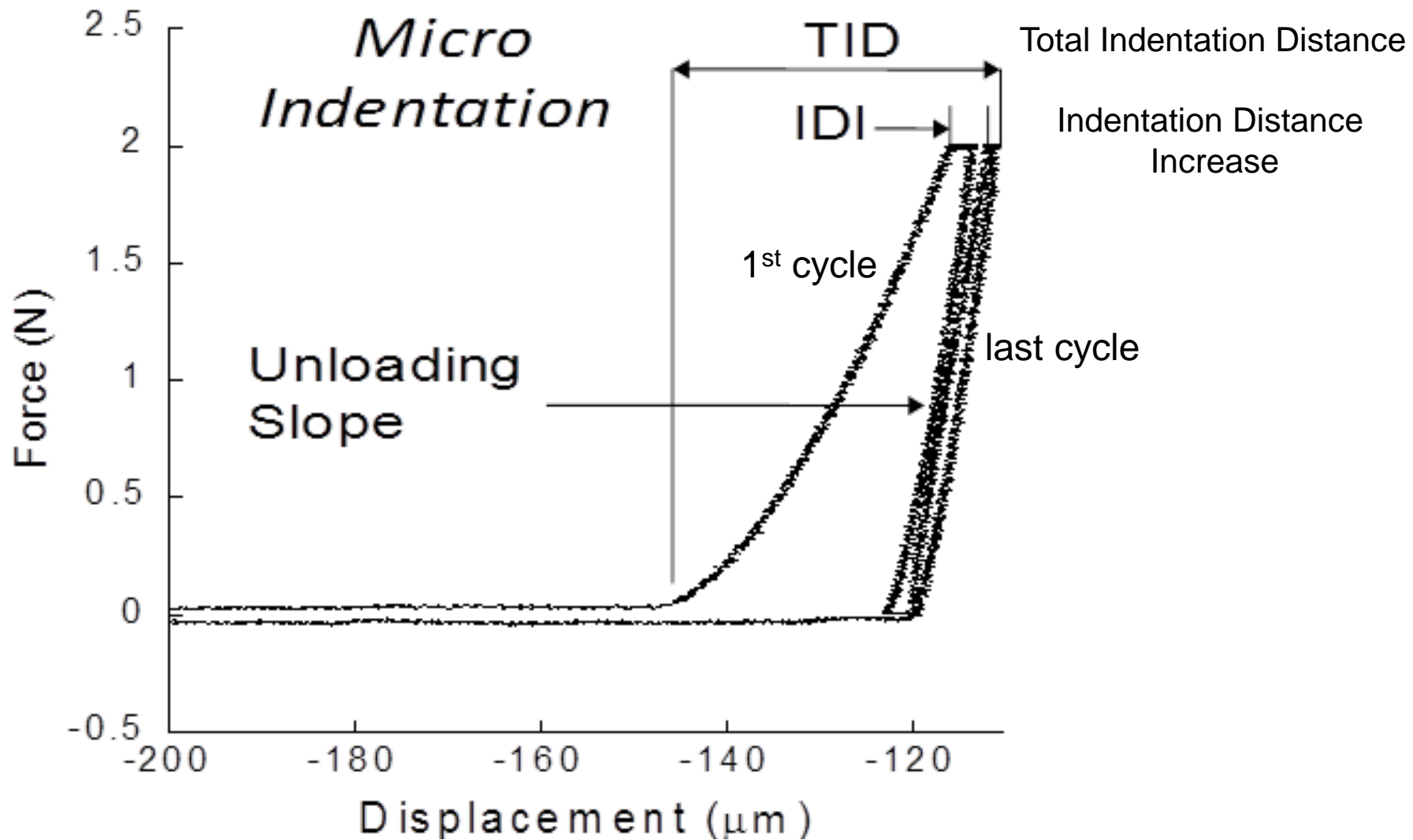
<sup>a</sup>“Load” and “force” can often be used interchangeably; to be precise, “load” is the more general term and may refer to “force” (push or pull) or “moment” (bend or torque).

Jepsen et al., JBMR 2015



# Microindentation (*BioDent*)

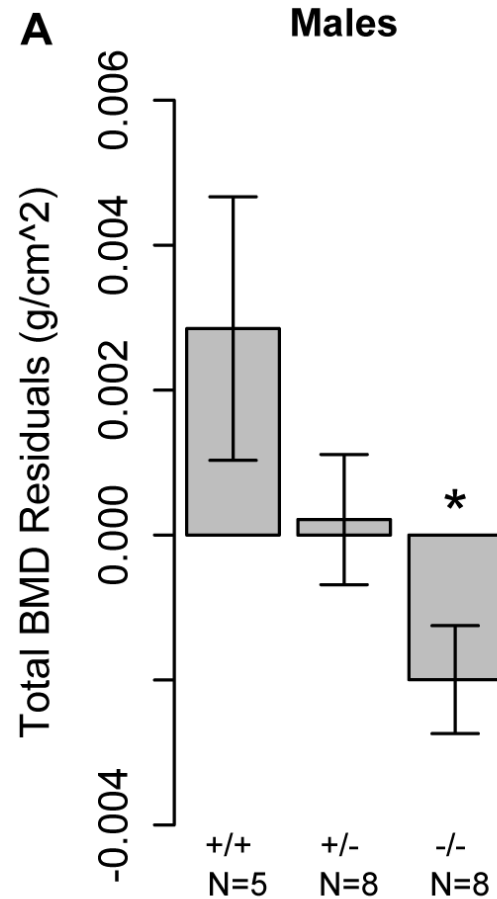




# Case Study 1: Asxl2 null mice

Wei Zou, Steve Teitelbaum

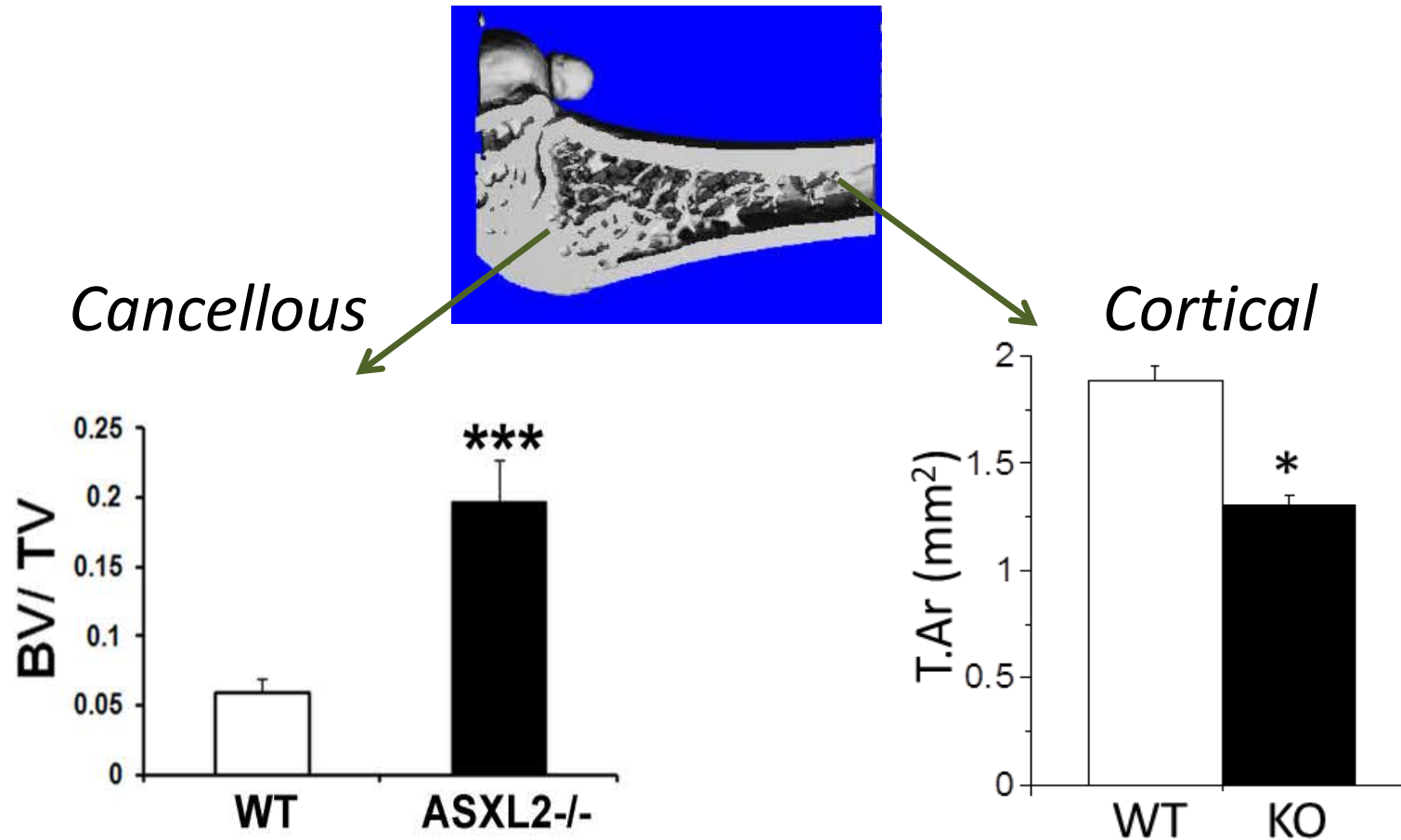
# *Asxl2*-null mice have low BMD



age- and body weight-adjusted

*Farber et al, 2011*

# *Asxl2*-null mice have more cancellous bone, but less cortical bone



Izawa, et al., *Cell Rep*, 2015

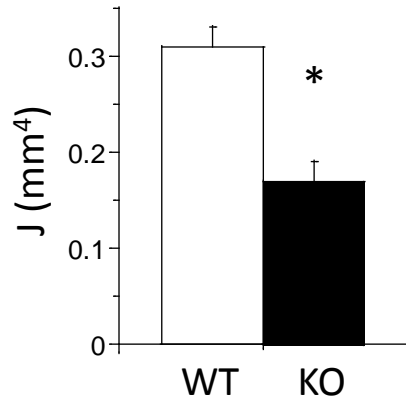
7/13/2018

MJ Silva, Washington University  
Musculoskeletal Research Center

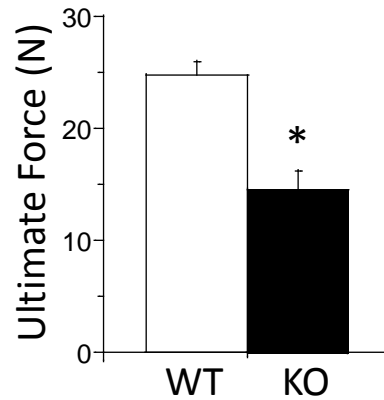
37

# *Asxl2*-null femurs: small & weak bones, normal material strength

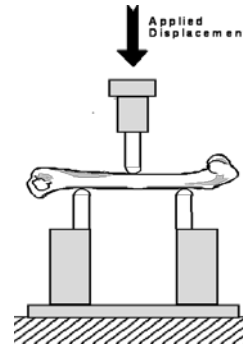
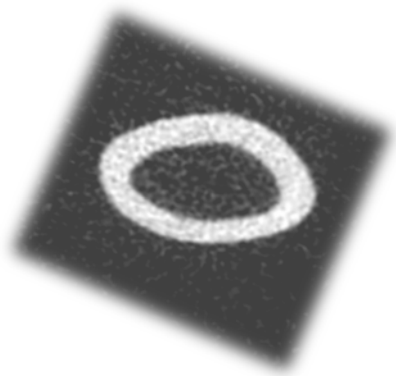
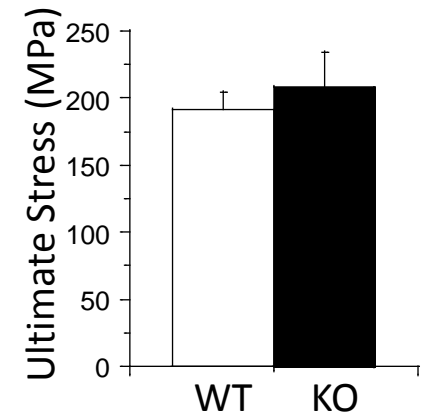
Moment of Inertia



Whole-bone strength



Material strength



## *Asxl2-null mice - Summary*

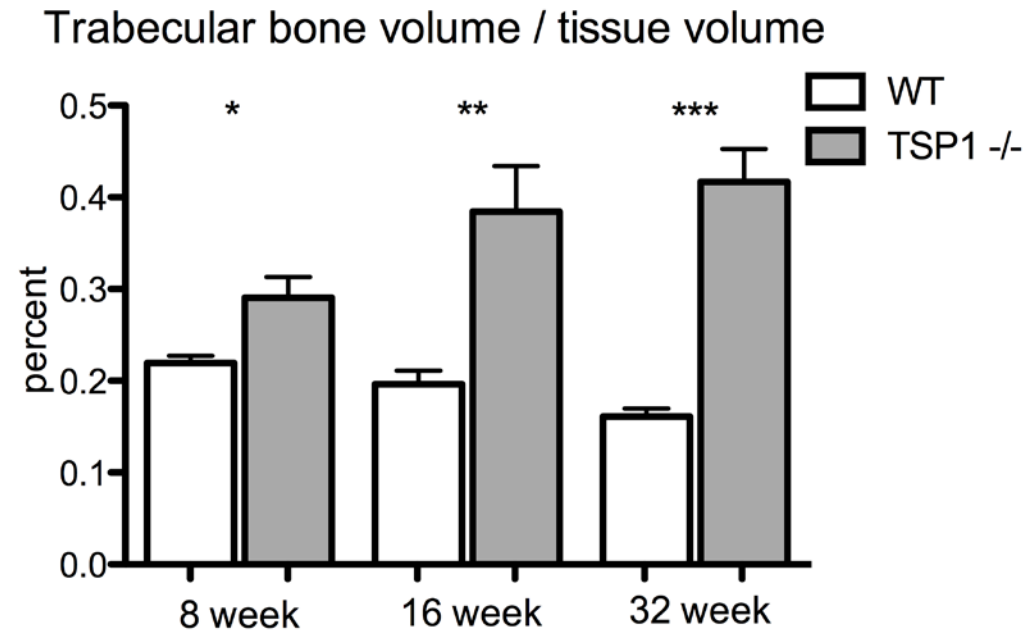
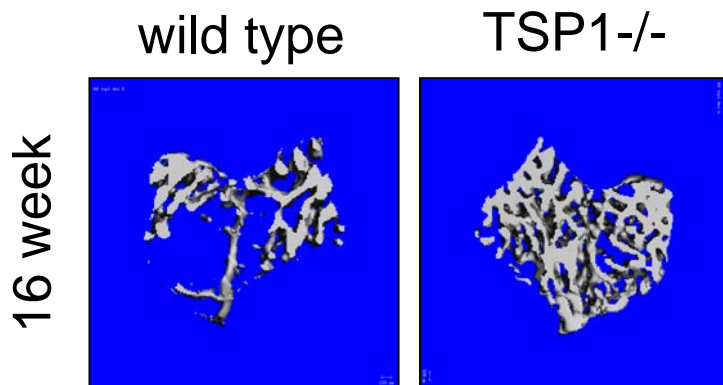
- Increased trabecular bone (osteoclast defect)
  - Decreased bone size & whole-body BMD
- Bones are weaker in proportion to their smaller size
  - Normal material properties

# Case Study 2: Tsp1 null mice

Sarah Amend, Kathy Weilbaecher

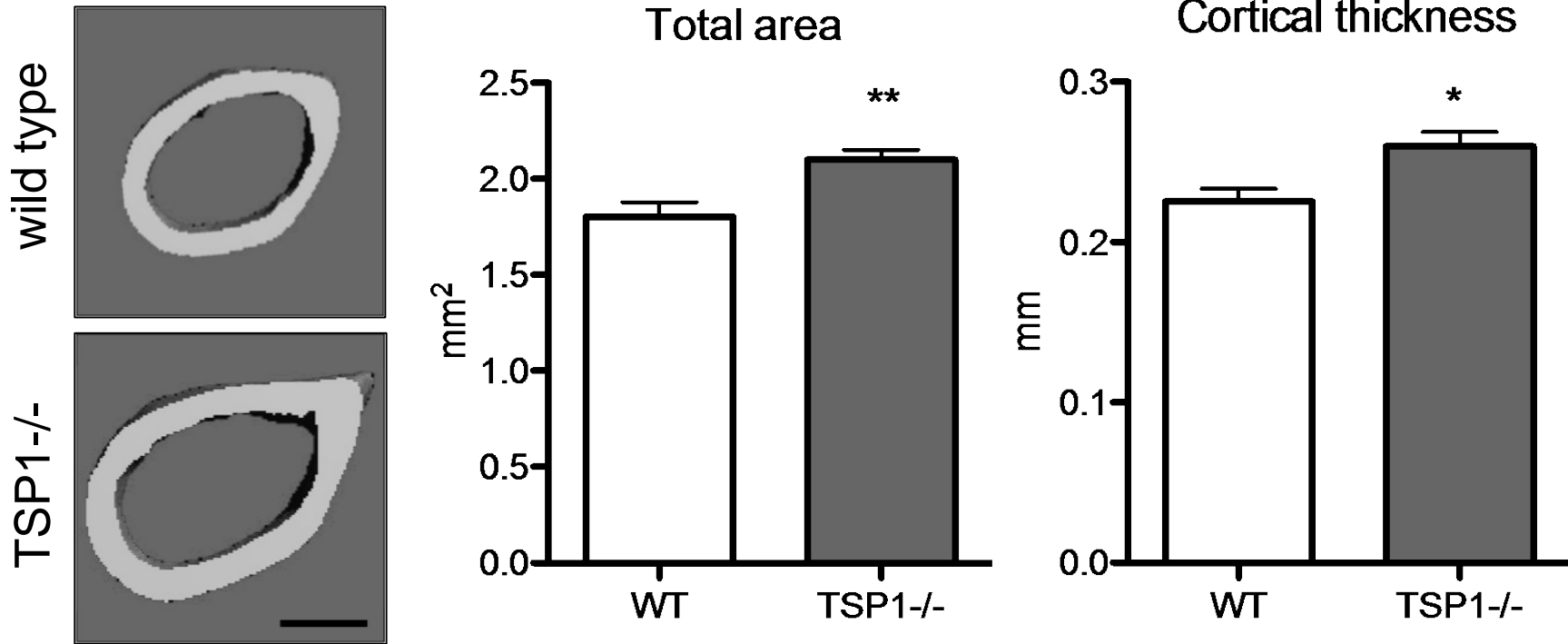


# *Tsp1*<sup>-/-</sup> mice have increased trabecular bone



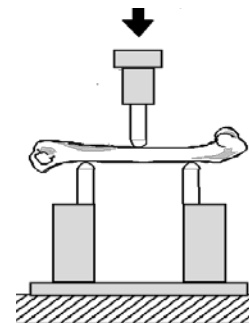
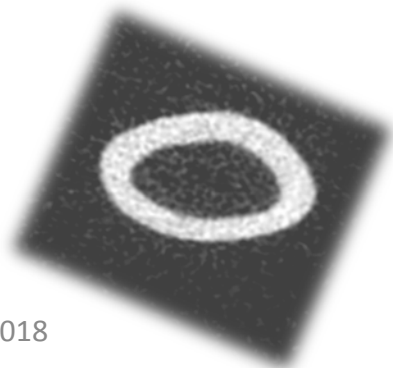
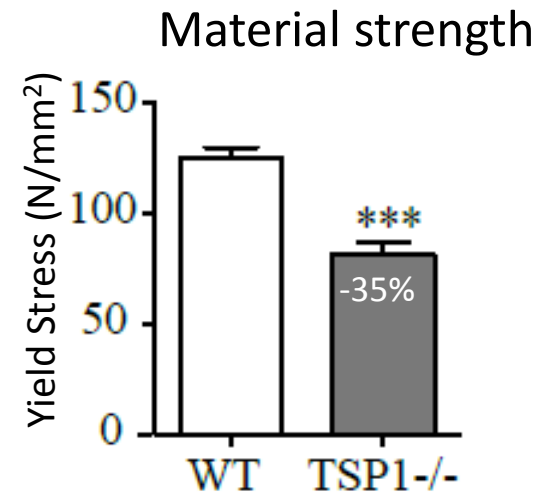
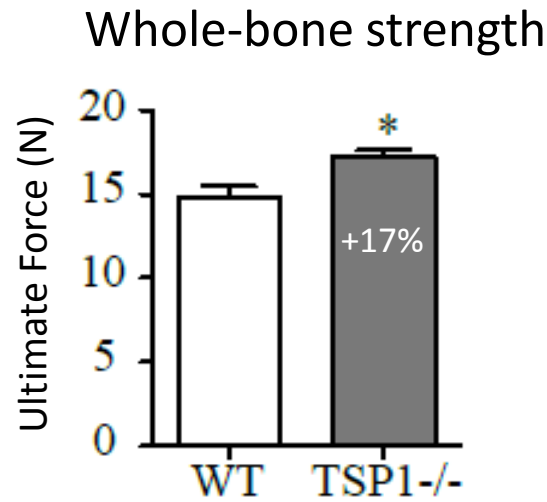
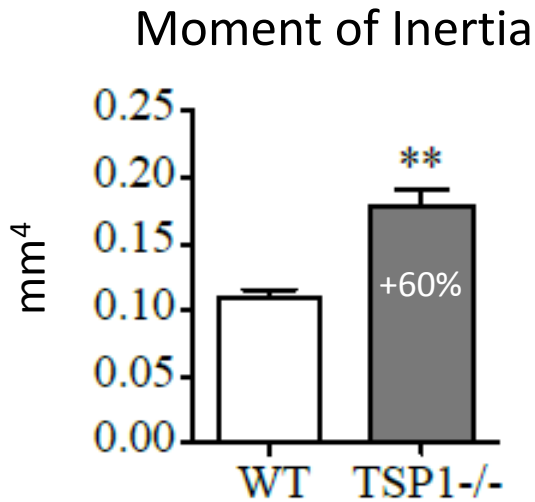
Amend et al., JBMR, 2015

# *Tsp1*<sup>-/-</sup> mice have larger bones



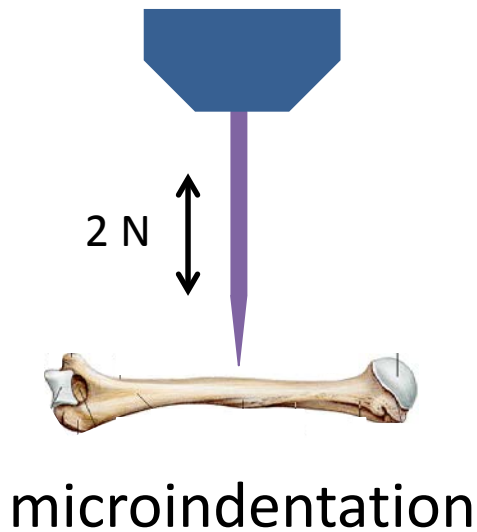
Amend et al., JBMR, 2015

# *Tsp1-null femurs: larger, moderately stronger, ...weaker material*

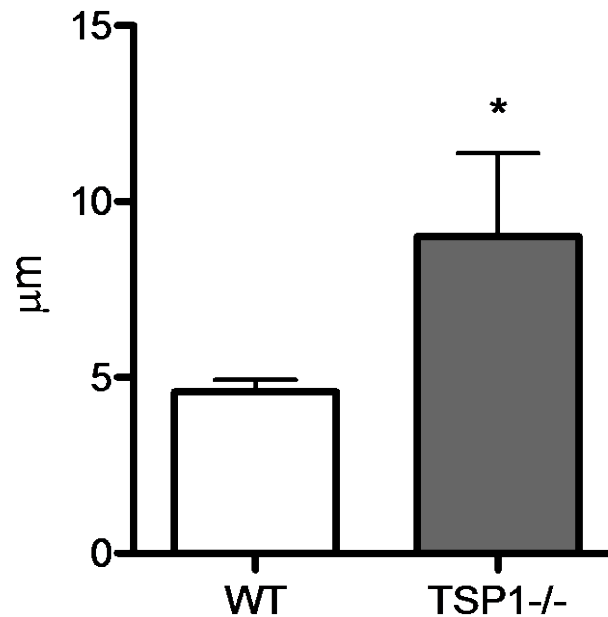


Amend et al., JBMR, 2015

# *Tsp1-null bone: damages more easily at material level*



Indentation distance increase



Amend et al., JBMR, 2015

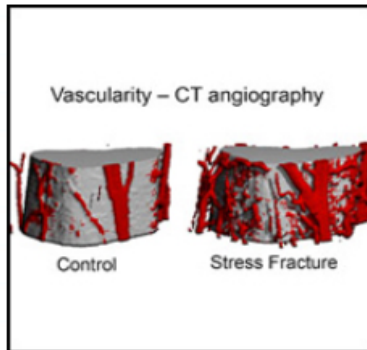
# *TSP1-null mice: Summary*

- Increased trabecular bone
- Increased cortical bone size
- Bones are moderately stronger – disproportionate to size
  - Impaired material properties

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## Structure and Strength Core



### Overview



The MRC is partnering with the Wash U Center for Cellular Imaging (WUCCI) to fund a Just-in-time/Microgrant program to support new imaging projects related to musculoskeletal research. [Read more...](#)

Structure and strength are perhaps the most relevant properties when assessing functional outcomes in animal models related to musculoskeletal biology and medicine. The **objectives** of the **Musculoskeletal Structure and Strength Core** are: 1) to increase access to existing resources for densitometry, imaging, and mechanical testing; 2) to enhance expert oversight and quality control; 3) to provide training and enrichment opportunities related to core services; and 4) to foster new interactions and enhance existing interactions between members of the Research Base of the Center for Musculoskeletal Research at Washington University.

**Consultation:** Contact the Core Director or any of our Associate Directors or Technical Staff to discuss your project. We can advise you on outcomes you may consider for a musculoskeletal structure / biomechanics experiment, and what we can do to help you obtain these outcomes. If you are in the planning stages or submitting a proposal, we can help with study design a priori. If you are starting your experiment, we can provide training or do the work for you. If you have collected your data, we can help with data analysis and interpretation.

**Imaging:** We support the use of x-ray based imaging of musculoskeletal structures and tissues from animal models (mouse to canine) generated by Research Base investigators. Available techniques include plane radiography, dual-energy x-ray absorptiometry (DXA), peripheral quantitative computed tomography (pQCT), and micro-computed tomography (microCT). Importantly, each of these techniques is available for in vivo as well as post mortem imaging.

**Mechanical Testing:** We perform mechanical testing to assess the functional properties of musculoskeletal tissues and structures from animal models (mouse to canine) generated by Research Base investigators. Available "standard" testing

#### Director

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#### Associate Directors

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#### Administrative Coordinator

Orthopaedic Surgery

### SERVICES

[Just in Time Funding](#)[Overview](#)[Fee Schedule and Forms](#)[Core B News](#)[MRC-WUCCI JIT/Microgrant Program](#)[Technical Notes and Presentations](#)[Project Gallery](#)[Publications](#)[Image Library](#)

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