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Main Figures: 6

Supplementary Figures: 7

Supplementary Tables: 0

Supplementary Videos: 0

Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read [Reporting Life Sciences Research](#).

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

► Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).
- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.
- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.
- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.
- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

		TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #	
example 1a	one-way ANOVA	Fig. legend	9, 9, 10, 15	mice from at least 3 litters/group	Methods para 8	error bars are mean +/- SEM	Fig. legend	p = 0.044	Fig. legend	F(3, 36) = 2.97	Fig. legend	
example results, para 6	unpaired t-test	Results para 6	15	slices from 10 mice	Results para 6	error bars are mean +/- SEM	Results para 6	p = 0.0006	Results para 6	t(28) = 2.808	Results para 6	

		TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #	
+ -	3b	Fisher's exact test, two-sided	Fig. legend, methods para 11	255	neurons from 2 monkeys, control condition	Fig. legend, results para 3	odds ratio	Fig. legend	As shown in Fig. S6a	Fig. legend		
+ -	3c	Bootstrapping	Fig. legend, results para 4	255	neurons from 2 monkeys, control condition	Fig. legend, results para 3	one-sided p value	Fig. legend	As shown in Fig. 3c	Fig. legend		
+ -	3e	Fisher's exact test, two-sided	Fig. legend, methods para 11	504	neurons from 2 monkeys, experimental condition	Fig. legend, results para 3	odds ratio	Fig. legend	As shown in Fig. S6b	Fig. legend		
+ -	3f	Bootstrapping	Fig. legend, results para 5	504	neurons from 2 monkeys, experimental condition	Fig. legend, results para 3	one-sided p value	Fig. legend	As shown in Fig. 3f	Fig. legend		
+ -	4a	Fisher's exact test, two-sided	Results para 8	759	neurons from 2 monkeys	Fig. legend	cell counts	Fig. legend	p = 0.11	Results para 8		
+ -	4b	Fisher's exact test, two-sided	Results para 8	457	neurons from 2 monkeys	Fig. legend	cell counts	Fig. legend	p = 0.10	Results para 8		
+ -	5b	Fisher's exact test, two-sided	Fig. legend, results para 10	255	neurons from 2 monkeys, control condition	Fig. legend	odds ratio	Fig. legend	As shown in Fig. S6c	Fig. legend		
+ -	5d	Fisher's exact test, two-sided	Fig. legend, results para 10	504	neurons from 2 monkeys, experimental condition	Fig. legend	odds ratio	Fig. legend	As shown in Fig. S6d	Fig. legend		
+ -	5e	Breslow-Day test	Fig. legend, results para 10	759	neurons from 2 monkeys	Fig. legend	p value	Fig. legend	As shown in Fig. 5e	Fig. legend	d.f. = 1; Breslow-Day statistic are shown in Fig. S7	
+ -	6a	Fisher's exact test, two-sided	Fig. legend	90	neurons from 2 monkeys, experimental condition	Fig. legend	cell counts	Fig. legend	p = 5.8e-5	Fig. legend		
+ -	6b	Fisher's exact test, two-sided	Fig. legend	66	neurons from 2 monkeys, experimental condition	Fig. legend	cell counts	Fig. legend	p = 2.8e-7	Fig. legend		

+	-	6c	Fisher's exact test, two-sided	Fig. legend	54	neurons from 2 monkeys, control condition	Fig. legend	cell counts	Fig. legend	$p = 4.2e-8$	Fig. legend		
+	-	6d	Fisher's exact test, two-sided	Fig. legend	37	neurons from 2 monkeys, control condition	Fig. legend	cell counts	Fig. legend	$p = 2.7e-7$	Fig. legend		
+	-	6e	Fisher's exact test, two-sided	Fig. legend	31	neurons from monkey V, A:B,C:A condition	Fig. legend	cell counts	Fig. legend	$p = 0.018$	Fig. legend		
+	-	6f	Fisher's exact test, two-sided	Fig. legend	12	neurons from monkey V, A:B,C:A condition	Fig. legend	cell counts	Fig. legend	$p = 0.0013$	Fig. legend		
+	-	S1	Kolmogorov-Smirnov test	Fig. legend	455	neurons from 2 monkeys	Fig. legend	cell counts	Fig. legend	$p = 0.83$	Fig. legend		
+	-	S2b	Fisher's exact test, two-sided	Fig. legend	553	neuronal responses from 2 monkeys, control condition	Fig. legend	odds ratio	Fig. legend	As shown in Fig. S6e	Fig. legend		
+	-	S2c	Bootstrapping	Fig. legend	553	neuronal responses from 2 monkeys, control condition	Fig. legend	one-sided p value	Fig. legend	As shown in Fig. S2c	Fig. legend		
+	-	S2e	Fisher's exact test, two-sided	Fig. legend	1171	neuronal responses from 2 monkeys, experimental condition	Fig. legend	odds ratio	Fig. legend	As shown in Fig. S6f	Fig. legend		
+	-	S2f	Bootstrapping	Fig. legend	1171	neuronal responses from 2 monkeys, experimental condition	Fig. legend	one-sided p value	Fig. legend	As shown in Fig. S2f	Fig. legend		
+	-	S3b	Wilcoxon signed rank test	Fig. legend	2841	neuronal responses from 2 monkeys, experimental condition	Fig. legend	neuronal response counts	Fig. legend	$p = 5.5e-176$	Fig. legend		
+	-	S3d	Wilcoxon signed rank test	Fig. legend	1713	neuronal responses from 2 monkeys, control condition	Fig. legend	neuronal response counts	Fig. legend	$p = 1.0e-167$	Fig. legend		
+	-	S3f	Wilcoxon signed rank test	Fig. legend	990	neuronal responses from 2 monkeys, experimental condition	Fig. legend	neuronal response counts	Fig. legend	$p = 3.1e-26$	Fig. legend		
+	-	S3h	Wilcoxon signed rank test	Fig. legend	597	neuronal responses from 2 monkeys, control condition	Fig. legend	neuronal response counts	Fig. legend	$p = 2.5e-38$	Fig. legend		
+	-	S4b	Fisher's exact test, two-sided	Fig. legend	353	neurons from monkey V, experimental condition	Fig. legend	odds ratio	Fig. legend	As shown in Fig. S6g	Fig. legend		
+	-	S4c	Bootstrapping	Fig. legend	353	neurons from monkey V, experimental condition	Fig. legend	one-sided p value	Fig. legend	As shown in Fig. S4c	Fig. legend		
+	-	S4e	Fisher's exact test, two-sided	Fig. legend	151	neurons from monkey C, experimental condition	Fig. legend	odds ratio	Fig. legend	As shown in Fig. S6h	Fig. legend		
+	-	S4f	Bootstrapping	Fig. legend	151	neurons from monkey C, experimental condition	Fig. legend	one-sided p value	Fig. legend	As shown in Fig. S4f	Fig. legend		

+ -	S5a	Fisher's exact test, two-sided	Fig. legend	210	neurons from 2 monkeys	Fig. legend	cell counts	Fig. legend	p = 0.13	Fig. legend		
+ -	S5b	Fisher's exact test, two-sided	Fig. legend	196	neurons from 2 monkeys	Fig. legend	cell counts	Fig. legend	p = 0.10	Fig. legend		
+ -	S6a	Fisher's exact test, two-sided	Fig. legend	255	neurons from 2 monkeys, control condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S6b	Fisher's exact test, two-sided	Fig. legend	504	neurons from 2 monkeys, experimental condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S6c	Fisher's exact test, two-sided	Fig. legend	255	neurons from 2 monkeys, control condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S6d	Fisher's exact test, two-sided	Fig. legend	504	neurons from 2 monkeys, experimental condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S6e	Fisher's exact test, two-sided	Fig. legend	553	neuronal responses from 2 monkeys, control condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S6f	Fisher's exact test, two-sided	Fig. legend	1171	neuronal responses from 2 monkeys, experimental condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S6g	Fisher's exact test, two-sided	Fig. legend	353	neurons from monkey V, experimental condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S6h	Fisher's exact test, two-sided	Fig. legend	151	neurons from monkey C, experimental condition	Fig. legend	two-sided p value	Fig. legend	As shown in Fig.	Fig. legend		
+ -	S7	Breslow-Day test	Fig. legend	759	neurons from 2 monkeys	Fig. legend	Breslow-Day statistic	Fig. legend	As shown in Fig. 5e	Fig. legend	d.f. = 1	Fig. legend

► Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

NA

If so, what figure(s)?

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

NA

If so, where is this reported (section, paragraph #)?

► Statistics and general methods

1. Is there a justification of the sample size?

If so, how was it justified?

Where (section, paragraph #)?

Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

We used 2 animals, which is standard practice for non-human primate studies. The number of cells collected for each condition and the number of trials ran for each cell (sample size) were not pre-determined using any statistical method, but were comparable to those of previous studies (Padoa-Schioppa and Assad, 2006). Methods, paragraph 4.

2. Are statistical tests justified as appropriate for every figure?

Where (section, paragraph #)?

Yes.

Results, paragraph 4-5 (fig. 3), 8 (fig. 4), 10 (fig. 5), 13-14 (fig. 6).

- a. If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?

Yes.

Methods, paragraph 5-16.

- b. Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?

Where is this described (section, paragraph #)?

The primary results of this study were based on analyses of categorical data. Fisher's exact test and the bootstrap procedure had minimal assumptions, and the Breslow-Day test had a cell count requirement (Agresti, 2007) that was met by our reduced contingency tables. Conversely, these analyses followed neuronal classification procedures based on least-squares linear regressions, which assume normality and equal variance. As previously discussed (Padoa-Schioppa and Assad, 2006), data in our experiments approximately satisfied normality because individual data points in the regressions were averages of 10-20 trials. In contrast, variances were generally unequal, with high-variance data points closer to the indifference point (where choices were split). Because correcting for unequal variance would effectively reduce the weight of these data points (Neter et al., 1990), which were in many respects the most informative, we deemed it preferable to use uncorrected data. However, a control analysis in which linear regressions were corrected for unequal variance provided similar results ((Padoa-Schioppa and Assad, 2006)), not shown). Methods, paragraph 16.

- c. Is there any estimate of variance within each group of data?

Is the variance similar between groups that are being statistically compared?

Where is this described (section, paragraph #)?

NA

- d. Are tests specified as one- or two-sided?

Yes.

Figure legend 3, 5, methods paragraph 11.

- e. Are there adjustments for multiple comparisons?

Yes.

3. Are criteria for excluding data points reported?

Was this criterion established prior to data collection?

Where is this described (section, paragraph #)?

NA

<p>4. Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data.</p> <p>If no randomization was used, state so.</p> <p>Where does this appear (section, paragraph #)?</p>	<p>Experimental and control sessions were interleaved in the experiments, but the order was not determined using a formal randomization.</p> <p>Methods, paragraph 4.</p>
<p>5. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included?</p> <p>If no blinding was done, state so.</p> <p>Where (section, paragraph #)?</p>	<p>Blinding was not used.</p> <p>Methods, paragraph 4.</p>
<p>6. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included?</p> <p>Where (section, paragraph #)?</p>	<p>Yes.</p> <p>Methods, paragraph 17.</p>
<p>7. Is the species of the animals used reported?</p> <p>Where (section, paragraph #)?</p>	<p>Yes.</p> <p>Methods, paragraph 1.</p>
<p>8. Is the strain of the animals (including background strains of KO/transgenic animals used) reported?</p> <p>Where (section, paragraph #)?</p>	<p>NA</p>
<p>9. Is the sex of the animals/subjects used reported?</p> <p>Where (section, paragraph #)?</p>	<p>Yes.</p> <p>Methods, paragraph 1.</p>
<p>10. Is the age of the animals/subjects reported?</p> <p>Where (section, paragraph #)?</p>	<p>Yes.</p> <p>Methods, paragraph 1.</p>
<p>11. For animals housed in a vivarium, is the light/dark cycle reported?</p> <p>Where (section, paragraph #)?</p>	<p>NA</p>
<p>12. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported?</p> <p>Where (section, paragraph #)?</p>	<p>NA</p>
<p>13. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)?</p> <p>Where (section, paragraph #)?</p>	<p>NA</p>
<p>14. Is the previous history of the animals/subjects (e.g. prior drug administration, surgery, behavioral testing) reported?</p> <p>Where (section, paragraph #)?</p>	<p>Yes.</p> <p>Methods, paragraph 1-2.</p>

- a. If multiple behavioral tests were conducted in the same group of animals, is this reported?

Where (section, paragraph #)?

15. If any animals/subjects were excluded from analysis, is this reported?

Where (section, paragraph #)?

- a. How were the criteria for exclusion defined?

Where is this described (section, paragraph #)?

- b. Specify reasons for any discrepancy between the number of animals at the beginning and end of the study.

Where is this described (section, paragraph #)?

► Reagents

1. Have antibodies been validated for use in the system under study (assay and species)?

- a. Is antibody catalog number given?

Where does this appear (section, paragraph #)?

- b. Where were the validation data reported (citation, supplementary information, Antibodypedia)?

Where does this appear (section, paragraph #)?

2. Cell line identity

- a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by [ICLAC](#) and [NCBI Biosample](#)?

Where (section, paragraph #)?

- b. If yes, include in the Methods section a scientific justification of their use--indicate here in which section and paragraph the justification can be found.

- c. For each cell line, include in the Methods section a statement that specifies:

- the source of the cell lines
- have the cell lines been authenticated? If so, by which method?
- have the cell lines been tested for mycoplasma contamination?

Where (section, paragraph #)?

► Data deposition

Data deposition in a public repository is mandatory for:

- a. Protein, DNA and RNA sequences
- b. Macromolecular structures
- c. Crystallographic data for small molecules
- d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available [here](#). We encourage the provision of other source data in supplementary information or in unstructured repositories such as [Figshare](#) and [Dryad](#).

We encourage publication of Data Descriptors (see [Scientific Data](#)) to maximize data reuse.

1. Are accession codes for deposit dates provided?

Where (section, paragraph #)?

► Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

1. Identify all custom software or scripts that were required to conduct the study and where in the procedures each was used.

The behavioral task was controlled through custom software (<http://www.monkeylogic.net>) based on Matlab (MathWorks). Data analyses were performed using Matlab.

2. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

Codes are available upon request.
Methods, paragraph 17.

► Human subjects

1. Which IRB approved the protocol?

Where is this stated (section, paragraph #)?

2. Is demographic information on all subjects provided?

Where (section, paragraph #)?

3. Is the number of human subjects, their age and sex clearly defined?

Where (section, paragraph #)?

4. Are the inclusion and exclusion criteria (if any) clearly specified?

Where (section, paragraph #)?

5. How well were the groups matched?

Where is this information described (section, paragraph #)?

6. Is a statement included confirming that informed consent was obtained from all subjects?

Where (section, paragraph #)?

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?

Where (section, paragraph #)?

► fMRI studies

For papers reporting functional imaging (fMRI) results please ensure that these minimal reporting guidelines are met and that all this information is clearly provided in the methods:

1. Were any subjects scanned but then rejected for the analysis after the data was collected?

a. If yes, is the number rejected and reasons for rejection described?

Where (section, paragraph #)?

2. Is the number of blocks, trials or experimental units per session and/or subjects specified?

Where (section, paragraph #)?

3. Is the length of each trial and interval between trials specified?

4. Is a blocked, event-related, or mixed design being used? If applicable, please specify the block length or how the event-related or mixed design was optimized.

5. Is the task design clearly described?

Where (section, paragraph #)?

6. How was behavioral performance measured?

7. Is an ANOVA or factorial design being used?

8. For data acquisition, is a whole brain scan used?

If not, state area of acquisition.

a. How was this region determined?

9. Is the field strength (in Tesla) of the MRI system stated?
- a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated?
- b. Are the field-of-view, matrix size, slice thickness, and TE/TR/flip angle clearly stated?
10. Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated?
11. Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)?
12. If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)?
13. How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.?
14. Were any additional regressors (behavioral covariates, motion etc) used?
15. Is the contrast construction clearly defined?
16. Is a mixed/random effects or fixed inference used?
- a. If fixed effects inference used, is this justified?
17. Were repeated measures used (multiple measurements per subject)?
- a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated?
18. If the threshold used for inference and visualization in figures varies, is this clearly stated?
19. Are statistical inferences corrected for multiple comparisons?
- a. If not, is this labeled as uncorrected?

20. Are the results based on an ROI (region of interest) analysis?

a. If so, is the rationale clearly described?

b. How were the ROI's defined (functional vs anatomical localization)?

21. Is there correction for multiple comparisons within each voxel?

22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined?

► Additional comments

Additional Comments