Re-distribution of Samurai Wasp (Trissolcus japonicus) on Fruit Farms in Ohio Celeste Welty¹, Kristina Fox Vik¹, & James Jasinski² ¹Ohio State University, Department of Entomology, Columbus OH THE OHIO STATE UNIVERSITY Π ²Ohio State University, Department of Extension, Urbana OH Methods, re-distribution tria Abstract Based on method used by Kaser et al. (2019) The samural wasp. Trissolcus igponicus (Ashmead) (Hymenoptera, Scelionidae), is a parasitoid that was first 15 parasitized egg masses per site =~420 parasitoids released one day before expected adult emergence of samurai wasp detected in Ohio from a sentinel egg sample of brown marmorated stink bug in Franklin County, August 2017. Additional detections in May 2018 were used to start a lab colony. Sentinel egg masses that were deployed at 10 fruit farms in central Ohio in June 2018 did not detect any samurai wasp, thus releases of the wasp were made at • each egg mass in mesh bag (Figure 9) in a 5-meter area of treeline adjacent to fruit crop (Figure 10) five farms, with the other five farms used as a no-release comparison. In August 2018, all 10 farms were sampled BMSB pheromone placed on stake at center of area at with sentinel eggs and with yellow sticky traps, and the samurai wasp was found at one release site and one norelease and no release sites (Figure 11) release site. In 2019, an additional 10 fruit farms were sampled with sentinel eggs, no samurai wasps were Figure 9. Each parasitized egg mass placed YEAR 1: 2018 detected, and releases were made at five of these farms with the other five farms as no-release comparisons. In pre-release sampling, 10 farms, June 2018 in mesh bag to shield it from predators August 2019 and again in July 2020, all 20 farms were sampled with sentinel eggs. Yellow sticky traps were also • parasitoid release at 5 farms from 7/19 to 8/9 used for sampling in 2020. No samurai wasps were detected by either sampling method, thus we have no evidence no release at 5 farms of the parasitoid being established at the release sites, however the sentinel egg sample size was likely too small or re-sampling, 10 farms, August-September 2018 too infrequent for adequate detection. More intensive sentinel egg mass sampling at the original site shows YEAR 2: 2019 seasonal trends in samurai wasp detection, and the benefit of more frequent sampling. pre-release sampling, 10 new farms, June 2019 parasitoid release at 5 new farms, 6/18 to 7/22 no release at 5 new farms Background The brown marmorated stink bug, Halyomorpha halys (Hemiptera: Pentatomidae; Figure 1), re-sampling, all 20 farms, August-September 2019 has been in Ohio since 2007. It is slowly spreading across the State, and there are increasing YEAR 3: 2020 reports of it causing injury to fruit crops (Figure 2) as well as to vegetable and grain crops. • The most common management tactic for control of the brown marmorated stink bug Figure 11. Pheromone lure on stake • no releases Figure 10. Clipping mesh at center of release area. • re-sampling, all 20 farms, June 2020 bag to tree in release area (BMSB) has been application of insecticides, which is not sustainable Biological control is of great interest to fruit growers. We have been aware of how well the samurai wasp is controlling BMSB eggs in Asia, but the Figure 1. Results Adult BMSB After release in 2018, the samural wasp was recovered from one sentinel egg mass USDA has not allowed this parasitoid wasp to be released in the USA. • Since an adventive population of the samurai wasp was found in Maryland in 2014 (Talamas at one release site, and on one sticky trap at one no-release site (Figure 12; Table 1) 🛨 = initial find After release in 2019, the samurai wasp was not recovered in sentinel egg samples et al. 2015), entomologists in many States have done sampling to find out whether this from any of the 10 release sites or the 10 no-release sites (Table 1) 800 parasitoid has spread to other States, and attempts are being made at its re-distribution (Kaser e 25 - 2 🛑 = release 2018 🔵 = no release 2018 In 2020, neither sentinel egg samples nor sticky trap samples found the samurai wasp at any of the 10 release or the 10 no-release sites (Table 1). O = release 2019 Retrieved egg masses showed the minimum number of samurai wasp emerged Figure 2. Typical ☆ = T.j. find, pos ranged from 101 to 379 per site, with an average of 270 for releases in 2018, and injury on apple by BMSB. 353 for releases in 2019 (Table 2). This is considered to be a high release rate. Parasitoid species found, other than T. japonicus, were: Anastatus mirabilis, was not detected Anastatus pearsalli, Anastatus reduvii, Telenomus persimilis, Telenomus podisi, Trissolcus brochymenae, Trissolcus euschisti. At the research farm where the samurai wasp was initially detected, sentinel egg Figure 12. Map of Ohio locations used for re distribution trial, marked with positive finds of samples resulted in positive finds of the samurai wasp each year (Table 1). Seasonal trends in detection of the samurai wasp at the research farm in 2020 samurai wasp EM TJ EM no Figure 3. Samurai wasps in (Figure 13A) show that this species was found from late May through late August. A even in small sample sizes (3-8 egg masses per day). Sampling at the rearing tube farm during the same time and in larger sample sizes did not result in any detections of the samurai wasp (Figure 13B). (76°F), 16:8 L:D igure 4. The colon Table 2. Emergence of the samurai wasp in Table 1. Parasitism of sentinel egg masses of samurai wasp. on Ohio fruit farms, 2017-2020. retrieved egg masses (EM) in Ohio re- distribution trials. year sile deploy # EM % min # 2018 win 719 15 14 92 343 2018 WH 7/19 15 14 92 345 2018 R7/17 15 14 97 365 2018 Ba 7/27 15 15 69 260 0101 E 7/27 15 15 69 260 distribution trials # egg masses with # #egg parasitoids emerged Year sites masses any species T. japonicus egg masses excised from bean plants in lab colony Commercial farms: Commercial farms: 2017 3 113 12 (10.6%) 2018 10 333 5 (1.5%) 2019 20 692 32 (4.6%) 2020 20 469 18 (3.8%) 1 (0.3%) 2018 St 7/27 15 12 2018 Hi 8/9 15 14 34 101 279 treeline adjacent to fruit plantings May (1) May 79 0 mean 74 2019 Hu 6/18 15 15 88 R 371 Research farm Research farm: 2017 1 505 24 (4.8%) 2 (0.4%) 2018 1 489 42 (8.6%) 31 (6.3%) 2019 1 512 21 (4.1%) 7 (1.4%) 2019 Hi 6/20 15 15 2019 GH 6/20 15 15 2019 AH 6/20 15 15 2019 JH 6/20 15 15 2019 Ro 7/22 15 15 2019 Ro 7/22 15 15 88 358 287 379 369 2019 1 2020 1 field lab 680 70 (10.3%) 41 (6.0%) (3 days) -> Figure 5. Sentinel egg preparation, distribution, Discussion evaluation. We could not confirm establishment of a samural wasp population at the 10 farms where a release was made. Figure 13. Seasonal distribution of sentinel egg Inadequate sample size or incorrect time of the season could explain why the Methods, yellow sticky trap sampling masses (EM) at Ohio sites, 2020, gray bars samurai wasp was not detected at sites where a release was made, but trends at double-sided traps from Alpha Scents (West Linn, OR) showing negative for samurai wasp, red bars the research farm in 2020 suggest that our sample size and timing were adequate deployed in treeline adjacent to fruit crop (Figure 6) showing positive for samurai wasp, A) at the for a site where this species is present. 5 traps per farm Franklin County site where the samurai wasp was An inadequate population of the host, brown marmorated stink bug, at the examined under microscope soon after retrieval from first detected in 2017, and B) at 20 farms used for Figure 6. Sticky trap clipped to commercial farms could have contributed to lack of success in re-distribution. field (Figures 7, 8) re-distribution tria ranch in treeline adjacent to We are hopeful that the samurai wasp will continue to spread on its own! fruit planting. YEAR 1: 2018 **References cited** traps deployed at 10 farms deployed for 4 weeks, in July-August Net CHICCS OF URCU AND CHICK AND deployed between time of parasitoid release and time of re-sampling by sentinel eggs owenstein, David M., Heather Andrews, Richard J. Hilton, Clive Kaiser and Nik G. Wiman. 2019. Establishment in an Introduced Range: Dispersal Capacity and Winter Survival of Trissolcus aponicus, an Adventive Egg Parasitoid. Insects, 10, 443; doi:10.3390/insects10120443 YEAR 2: 2019 2019: none used lamas, Elijah J., Megan V. Herlihy, Christine Dieckhoff, Kim A. Hoelmer, Matthew Buffington, Marie-Claude Bon, Donald C. Weber. 2015. Trissolcus japonicus (Ashmead) (Hymenoptera, elionidae) emerges in North America. Journal of Hymenoptera Research 43: 119-128. https://doi.org/10.3897/IHR.43.4661 YEAR 3: 2020 traps deployed at 20 farms Figure 7. Typical Figure 8. Sticky traps held in Greatly appreciated is permission for farm use by 20 fruit growers, technical assistance from Becca Welsh, Ariel Fisher, Cecilia Stone, Brittany Heigley, Vashti Tatman, Brooke Friend, and Madeline Fox, and funding from USDA-NIFA-SCRI Project: Management of Brown Marmorated Stink Bug in US Specialty Crops, grant # 2016-S1181-25409, and USDA NIFA EIP, grant # 2017-2006-2717A. Acknowledgements sticky trap after 2lab after deployment, deployed for 2 weeks awaiting evaluation.

- deployed after sentinel egg mass sampling completed
- week deployment.

et al. 2019, Lowenstein et al. 2019, Hoelmer et al. 2017). • Our initial objective in 2017 was to determine whether or not the samurai wasp could be

found in Ohio.

 Once we did detect the samural wasp in Ohio in 2017, our objectives were to survey additional sites to find out if it is present, and to attempt to re-distribute it to farms where it

Methods, lab colony of samurai wasp

in 50-ml plastic tubes with fine netting over tops (Figures 3. 4), held at 12°C (57°F), 9:15 L:D

held fed dilute honey once per week, brushed on netting for progeny production: in one tube, two females provided with one fresh BMSB egg mass for 1 week, held at 24°C

average of 12 days from egg provision until emergence of new adults

Methods, sentinel egg mass sampling

- BMSB, and taped to cards (Figure 5) cards clipped to underside of leaf on trees at edge of
- 15-25 fresh egg masses deployed per farm
 retrieved after 72 hours · held for 6+ weeks in lab to observe parasitoid emergence