Linear and Nonlinear W-Cells in C-Laminae of the Cat's Lateral Geniculate Nucleus

MRIGANKA SUR AND S. MURRAY SHERMAN

Department of Neurobiology and Behavior, State University of New York, Stony Brook, New York 11794

SUMMARY AND CONCLUSIONS

1. We used standard, single-cell recording techniques to study the response properties of 34 W-cells in the C-laminae of the cat's lateral geniculate nucleus. By W-cell, we mean a poorly responsive geniculate neuron that receives slowly conducting retinal afferents; these are quite distinct from geniculate X- and Y-cells. Our measurements included response latency to optic chiasm stimulation, plots of the receptive-field center, time course of response, and responses to counterphased, sine-wave gratings. This last measurement also involved the determination of contrast sensitivity, which is defined as the inverse of the contrast needed to evoke a threshold response at a particular spatial and temporal frequency of the grating. Many of these responses were compared to those of geniculate X- and Y-cells recorded in the A-laminae.

2. Each of the W-cells responded with a latency of at least 2.0 ms to optic chiasm stimulation, and most (76%) exhibited a latency of at least 2.5 ms. However, only 26 of these W-cells responded to visual stimuli, and these responses were weak or "sluggish," as has been reported previously. Receptive fields of these W-cells tended to be large, compared to those of X- and Y-cells, and included 11 on-center, 13 off-center, and 2 on-off center fields.

3. W-cells exhibited either linear (12 cells) or nonlinear (14 cells) spatial and temporal summation, as determined from their responses to counterphased, sine-wave gratings. Linearity of spatial summation was determined by measuring contrast sensitivity as a function of the grating's spatial phase. The linear W-cells' responses were sinusoi-

dally phase dependent, and the nonlinear Wcells' responses were independent of spatial phase. Linearity of temporal summation was determined by the presence or absence of harmonic distortion in the response relative to the grating's counterphase rate. Linear W-cells responded chiefly at the grating's fundamental temporal frequency, whereas much of the nonlinear W-cells' responses occurred at the second harmonic of the grating's temporal frequency. Thus, nonlinear W-cells exhibited many of the characteristics previously described for Y-cells.

4. Spatial and temporal contrast-sensitivity functions were determined for seven linear and eight nonlinear W-cells. Overall sensitivity values of the linear and nonlinear W-cells were comparable, but these groups differed in terms of the nature of the response component (linear or nonlinear) that was more sensitive.

5. The linear W-cells in our sample included both tonic (comparable to the "sluggish-sustained" type of retinal ganglion cells) and phasic (comparable to "sluggish-transient" ganglion cells) types, while all nonlinear W-cells were phasic. Otherwise, no difference between linear and nonlinear Wcells was seen for latency to optic chiasm stimulation, receptive-field size, overall contrast sensitivity, responsiveness to visual stimuli, overall spatial resolution, or temporal resolution.

6. The best contrast sensitivity exhibited by the W-cells was typically less than onefourth as great as the comparable sensitivity measured for 9 X- and 14 Y-cells with similar receptive-field locations. This relatively poor contrast sensitivity for W-cells may relate to their sluggish responses to visual stimuli.

7. There were systematic differences between the spatial contrast-sensitivity functions derived for linear W-cells and X-cells and for nonlinear W-cells and Y-cells. Linear W-cells did not exhibit the reduced sensitivity to low spatial frequencies seen in X-cells. At low temporal frequencies, the second-harmonic response component of nonlinear W-cells tended to be more sensitive than the fundamental component at all spatial frequencies, whereas for Y-cells the fundamental component was more sensitive than the second-harmonic component at lower spatial frequencies and less sensitive at higher ones. Furthermore, the spatial resolutions of W-cells were considerably less than those of either X- or Y-cells.

8. By a number of criteria, W-cells can be classified separately from X- and Y-cells. The division of W-cells into linear and nonlinear types reflects their responses to sinewave gratings but does not seem to relate to any other response property tested to date. The significance of this division for either cell classification or visual function is presently unclear. More generally, however, Wcells may be involved in vision related to low spatial frequencies at contrasts well above threshold.

INTRODUCTION

The cat's retinogeniculocortical system is comprised of at least two parallel, functionally independent pathways that contain either X- or Y-cells (see Refs. 24, 29, 33, 34, 39 for recent reviews). The X- and Y-cell classes include both retinal ganglion cells and neurons in the lateral geniculate nucleus, and the receptive-field properties of the geniculate X- or Y-cells are much like those of their retinal counterparts (3, 4, 20, 23, 32, 36). Other cell types with quite different response properties have been recently discovered among retinal ganglion cells and neurons of the geniculate C-laminae; these were originally termed W-cells (5, 6, 8, 37, 40, 41, 43, 44; see also Refs. 10, 25). X- and Y-cells have been intensively studied since their original description by Enroth-Cugell and Robson (11), and consequently we know a great deal about their response properties (e.g., Ref. 2-4, 9, 18-23, 32, 35, 36). Wcells, by contrast, were only recently described, and much less is known about their response properties (e.g., Refs. 5, 6, 8, 37, 40, 41, 43, 44). The purpose of this paper is to describe certain features of W-cells located in the C-laminae of the lateral geniculate nucleus.

Recent evidence has shown that retinal ganglion W-cells are a heterogeneous group of neurons and that use of the term W-cell may be a misleading oversimplification. Indeed, other terminology has been suggested (29). It is neither clear to what extent Wcells in the C-laminae of the lateral geniculate nucleus represent a genuine homogeneous functional class nor to what extent their response properties reflect those of their retinal afferents. These gaps in our knowledge form part of the reason that we initiated the present study. However, rather than adopt new terminology, we have elected to continue use of the term W-cell for these geniculate neurons (cf. Ref. 39). We both recognize and emphasize the shortcomings of such terminology, and we expect to adopt better terminology when these geniculate neurons and their retinal afferents are more completely characterized. This point will be considered more fully in DISCUSSION.

By certain criteria, X-cells show linear spatial and temporal summation within their receptive fields, and Y-cells do not (11, 18, 22, 23, 32). That is, for a visual stimulus consisting of a counterphased, sine-wave grating, a "null position" can be found at which X-cells cease to respond because inhibitory and excitatory influences of the stimulus linearly sum to zero. Also, X-cell responses to other grating positions occur predominantly at the fundamental temporal frequency of the counterphasing stimulus. No grating null position can be found for the Y-cells, and their responses are distorted temporally by harmonics of the stimulus frequency, particularly the second harmonic. Other differences between X- and Y-cells 35, 36). Compared to Y-cells, X-cells tend to exhibit 1) slower conducting axons, 2) poorer responses to low spatial frequencies or large shapes, 3) better responses to high spatial frequencies or small targets, 4) smaller receptive-field centers, 5) poorer responses to high temporal frequencies or fastmoving targets of contrast appropriate to excite their surrounds, and 6) more tonic responses to appropriate stimuli located in their receptive-field centers.

For the most part, W-cells in the retina and lateral geniculate nucleus can be distinguished from X- and Y-cells by two poorly defined criteria (e.g., Refs. 5, 6, 8, 40, 41, 43, 44). W-cells respond sluggishly to visual stimuli compared to the "brisk" responses of X- and Y-cells, and W-cells have more slowly conducting axons than do X- and Ycells, although the fastest conducting axons of W-cells overlap with the slowest of Xcells. We sought to clarify some of these differences between W-cells and X- or Ycells by noting the responses of W-cells to sine-wave gratings. We found that some Wcells exhibit linear spatial and temporal summation (like X-cells), and others appear nonlinear (like Y-cells). Also, W-cells have contrast sensitivity that is much worse than that of either X- or Y-cells. Preliminary results of this research have been recently reported (42).

MATERIALS AND METHODS

Experiments were performed on 10 adult cats. Methods of the physiological preparation, visual stimulation, electrophysiological recording, and data analysis were similar to those we have described in detail previously (20, 23). Briefly, cats were anesthetized with halothane for initial surgery and maintained thereafter on a 70/30 mixture of nitrous oxide/oxygen with continuous infusion of gallamine triethiodide (3.6 mg/h) and d-tubocurarine (0.7 mg/h) for paralysis. They were artifically ventilated, and end-tidal carbon dioxide was monitored and kept at 4.0%. Body temperature was maintained at 38°C. Pupils were dilated and nictitating membranes retracted with topical application of atropine sulfate and phenylephrine hydrochloride. The corneas were covered with contact lenses that included a 3-mm artificial pupil. Retinoscopy ensured that the retinas were conjugate with the visual stimuli delivered on a cathode-ray tube 57 cm away.

Bipolar stimulating electrodes (insulated tungsten wires with 0.5-mm exposed tips) were placed across the optic chiasm (one pair) and in the white matter underlying the visual cortex (two pairs). Electrical stimuli consisted of brief pulses (≤ 5 mA for $\leq 50 \ \mu$ s). Antidromic stimulation of geniculate neurons from visual cortex was determined by a spike-collision test by which an orthodromic spike collides with and blocks the antidromic one. Transynaptic activation of geniculate neurons from cortex was also seen in many cases. Latency of response was measured from the stimulus artifact to the foot of the action potential. Both the latency range and modal latency were measured for each cell, and measurements of the modal latency are indicated in RESULTS.

Visual stimuli consisted of bright or dark handheld targets moved on a tangent screen or counterphased, sine-wave gratings presented on a cathode-ray tube. The gratings were vertically oriented and had a mean luminance of 36 cd/m^2 . The spatial frequency, temporal frequency (counterphase rate), spatial phase angle,¹ and contrast (between 0 and 0.84) of the grating could be continuously varied. For many cells, responses were stored in a computer (PDP 11/34) for subsequent Fourier analysis.

Single-unit recording from geniculate cells was accomplished with glass micropipettes filled with 3 M KCl or NaCl (10-30 M Ω at 100 Hz). In every penetration in which W-cells were recorded in the C-laminae, receptive fields for X- and Ycells were noted in laminae A and A1 for comparison. Y-cells, and rarely some X-cells, were also recorded in the dorsal part of the C-laminae (43). X- and Y-cells responded briskly to visual stimuli and were distinguished from one another on the basis of a battery of tests (see INTRO-DUCTION and Refs. 20, 22, 23, 31). These tests, in decreasing order of priority, are 1) linearity of spatial summation to counterphased, sine-wave gratings; 2) latency of response to optic chiasm stimulation, 3) responsiveness of the surround to fast-moving $(>200^{\circ}/s)$, large targets; 4) size of the receptive-field center; and 5) time course (tonic or phasic) of responses to appropriate standing contrasts. W-cells were found in the Claminae, but not in the A-laminae, and were distinguished by long-latency responses to optic tract stimulation (≥ 2.0 ms) and weak or sluggish responses to visual stimuli (5, 6, 8, 40, 41, 43, 44). As we shall show (see RESULTS), W-cells exhibit markedly poor contrast sensitivity, and this probably relates to their sluggish responsiveness to visual stimuli.

We studied the responses of W-cells to handheld visual targets and to the counterphased, sinewave gratings. Hand-held targets (usually small, flashing spots of light) were used to determine the receptive-field borders; whether a cell was on-center, off-center, or generated responses to both onset and cessation of the stimulus (on-off center);

¹ Spatial phase angle refers to the relative position of the grating perpendicular to its orientation (i.e., horizontal position). One stimulus cycle of the sine-wave variation in luminosity is 360° of spatial phase angle. A phase shift of 90° is a movement of one-fourth of a stimulus cycle; 180°, half a cycle, etc.



FIG. 1. Histogram of response latencies to optic chiasm stimulation (OX latency) for 12 linear W-cells (mean, 2.7 ms), 14 nonlinear W-cells (mean, 3.0 ms), and 8 W-cells that were unresponsive to visual stimulation (mean, 3.8 ms).

responsiveness of the neurons to a fast-moving disk $(>200^{\circ}/s)$ of contrast appropriate to excite the surround; and whether the cell responded in a tonic or phasic manner to center stimulation (dark spots were used to test off-center cells). A cell was classified as tonic if it sustained a response above background to a centered spot for at least 10 s, and phasic if not. Phasic cells nearly always returned to background firing rates within 2 s of the stimulus presentation.

RESULTS

We obtained data from 34 W-cells in the C-laminae of the cat's lateral geniculate nucleus. W-cells were found in each of the C-laminae (i.e., laminae C and C_2 innervated by the contralateral eye and lamina C_1 innervated by the ipsilateral eye; see Ref. 43). No obvious differences in response properties were seen for W-cells among these laminae.

General characteristics of W-cells

Each of the 34 recorded W-cells responded to optic chiasm stimulation with a latency ≥ 2.0 ms, and 26 (76%) had response latencies ≥ 2.5 ms. Figure 1 shows the range

and distribution of these latencies. Eight cells could not be reliably activated by any visual stimuli used, even though neighboring neurons were visually responsive, and thus the presumed receptive-field locations were known. Nonetheless, these eight visually unresponsive cells responded reliably to optic chiasm stimulation, but they tended to have longer latencies than most visually responsive W-cells (see Fig. 1). We have tentatively grouped these eight unresponsive cells as W-cells for two reasons: their visual unresponsiveness may be an extreme characteristic in a continuum of poor W-cell responsiveness (e.g., Refs. 5, 6, 8, 40, 43, 44; see also below), and the morphological features of the visually responsive and unresponsive W-cells are essentially equivalent to one another (38). The remaining 26 Wcells responded weakly or sluggishly to visual stimuli. Their responses also seemed relatively variable compared to those of X- and Y-cells. More detailed receptive-field data are described below for 24 of these 26 Wcells, because the remaining 2 neurons, though visually responsive, were unreliable, weak, and inconsistent in their responses to visual stimulation.

From the cortical electrodes, we were able to activate only five W-cells antidromically (mean latency of 2.4 ms with a range of 2.1-3.5 ms). The failure to activate more W-cells might reflect the wide distribution of projections from the C-laminae (15, 28) and limited activation region of our cortical electrodes.

Of the visually responsive W-cells, 11 had on-center receptive fields, 13 were off-center, and 2 were on-off. Antagonistic surrounds were absent or too weak to detect in all but nine of these cells. One on-off cell had a suppressive surround. The receptive-field centers of W-cells were up to 5° across in the central 10° of visual field and were larger than those of all X-cells and nearly all Ycells at comparable eccentricities. However, due to both the weak responses of most Wcells to spots of light as well as the lack of sharply defined receptive-field borders, we are not very confident of these center size estimates. With this proviso, Fig. 2 shows the distribution of center sizes as a function of receptive-field eccentricity from the area centralis representation. The on-off center W-cells responded in a phasic manner to centered stimuli, but the on- or off-center cells were either tonic (5 cells) or phasic (19 cells). The tonic and phasic W-cells probably correspond, respectively, to the sluggish-sustained and sluggish-transient cells of Cleland et al. (5, 6, 8).

Contrast sensitivity and response versus contrast measurements

Contrast sensitivity is defined as the inverse of the minimum stimulus contrast needed to evoke a detectable or reliable neural response above the background level; it was measured by two methods that provided essentially the same result. First, a neuron's action potentials were displayed on a storage oscilloscope whose sweep was triggered by the temporal frequency wave form of the counterphased sine-wave grating. The temporal waveform of the stimulus was also displayed on the same time base. Repeated stimulus/response pairs were superimposed

and stored. From this we could determine for each spatial and temporal frequency the linear and nonlinear response components. The presence of action potentials on only one temporal half-cycle of the stimulus waveform denotes a fundamental or linear response component, while clustering of action potentials related to both half-cycles denotes a frequency-doubled response that is the second-harmonic or nonlinear component. The contrast threshold of each component was measured as the lowest contrast value needed to detect the component reliably. However, this method could overlook harmonic components in a response that occurred at either half-cycle of the stimulus.

The second method consisted of plotting response (size of either the fundamental or the second-harmonic component of response) versus contrast (Fig. 3). Such a graph has a portion where response increases in an approximately linear fashion with contrast and a portion where the response sat-



FIG. 2. Plot of receptive-field center size versus eccentricity of the field from the area centralis for 11 linear and 13 nonlinear W-cells. The center sizes for one linear and one nonlinear W-cell were not determined due to the unreliable responsiveness of these two cells, and they are excluded from this plot. For cells shown, there is a correlation between center size and receptive-field eccentricity (r = 0.6, P < 0.01).



Response versus contrast functions. The FIG. 3. dashed line in each graph represents the level of response component (fundamental or second harmonic) found in the spontaneous activity during an equivalent time period for each of the contrast-response measures (15 s). A: function for a linear W-cell's fundamental response component. The stimulus was a 0.2 cycle/deg sine-wave grating counterphased at 2 Hz. The grating's spatial phase angle was placed 90° from the fundamental response component's null position (see text and Fig. 4) and was thus at the most sensitive position for this reponse. B: function for a nonlinear W-cell's second-harmonic response component. The stimulus was a 0.1 cycle/deg sine-wave grating counterphased at 2 Hz. It was placed at the null position of the fundamental response component for this cell (see text and Fig. 4).

urates. From the linear portion of the graph, two measures of sensitivity could be obtained. First, one could choose a criterion response and obtain the contrast required to vield that response (11, 18). For example, the contrast needed to yield a response just higher than the background level could be obtained from the intersection of the linear portion of the contrast-response function with the fundamental component found in spontaneous activity (Fig. 3A, B). Alternatively, the slope of this linear portion provides a measure of sensitivity (18). For the two cells illustrated in Fig. 3A. B. the oscilloscope method yielded contrast thresholds of 0.09 (sensitivity, which is the inverse of the contrast at threshold, equals 11.1) for the stimulus used in Fig. 3A, and 0.17 (sensitivity equals 5.9) for the stimulus used in Fig. 3B. The contrast-response graph yielded contrast threshold values of 0.035 (sensitivity equals 28.6) and 0.13 (sensitivity equals 7.7) from the background response criteria. The slopes of the two graphs yielded sensitivity values of 22.1 and 16.0 (in spikes per second/contrast).

Contrast-sensitivity values obtained either by the oscilloscope method or from the contrast-response graphs were internally consistent and provided similarly shaped contrastsensitivity functions. For this reason, we employed the simpler and faster method of using the oscilloscope to determine contrast sensitivity, although these values were frequently verified with computer-generated contrast-response functions.

Linear and nonlinear W-cells

Two tests were used to assess the linearity of spatial and temporal summation of Wcells to visual stimuli. The test for linearity of spatial summation measured the evoked response as a function of spatial phase of a sine-wave grating. A linear response (e.g., from an X-cell) varies sinusoidally as a function of spatial phase between the phase angle of minimal response (i.e., null position) and one of maximal response, whereas a nonlinear response (e.g., from a Y-cell) is relatively independent of spatial phase (18, 19, 32). The test for linearity of temporal summation measured the neuronal response dynamics relative to the stimulus temporal frequency. A linear (X-cell) response occurs mostly at the fundamental temporal frequency of the stimulus, whereas a nonlinear (Y-cell) response contains considerable second- (and higher order) harmonic distortion of the stimulus (e.g., a "doubling" response that occurs at twice the stimulus frequency; see Refs. 18, 19). A combination of these nonlinearities for Y-cells leads to responses characterized by "phase-independent doubling" (18, 19), indicating that the response occurs at twice the temporal frequency of the stimulus and is unchanged with shifts in the spatial phase of the stimulus. Based on these criteria, our sample of W-cells included both linear (12 neurons) and nonlinear (14 neurons) types among those that responded to visual stimuli.

SPATIAL PHASE DEPENDENCE OF CONTRAST SENSITIVITY. Figure 4 shows the variation of contrast sensitivity (see above) with spatial phase for a linear W-cell and a nonlinear W-cell. The stimulus consisted of a sinewave grating counterphased at 2 Hz. The signs (positive or negative) of the contrastsensitivity values were derived from the relationship between the response and an arbitrarily chosen half-cycle of the temporal waveform of the stimulus (see Ref. 18). Figure 4A depicts the phase dependence of the linear W-cell's response, which occurs at the fundamental temporal frequency of the stimulus. This phase dependence is approximately sinusoidal. Figure 4B shows the phase dependence of the nonlinear W-cell's frequency-doubled response. The sensitivity is now plotted above and below zero, because responses occur at both temporal half-cycles. This response is basically independent of spatial phase. Although not illustrated, it was possible in some nonlinear W-cells to measure the spatial phase dependence of the fundamental response (which was typically much weaker or less sensitive than the second-harmonic response). Such a fundamental response had a sinusoidal phase dependence much like that illustrated in Fig. 4A.

Due to the phase dependence of the fundamental response component and phase-independent nature of the second-harmonic component, contrast thresholds could be determined by the oscilloscope method in the following manner (see above). To determine the sensitivity of the second-harmonic component, we canceled the fundamental component by adjusting the grating's spatial phase to the null position for the fundamental component. The remaining response was thus frequency doubled and independent of



FIG. 4. Plots of contrast sensitivity versus spatial phase angle. A: plot for a linear W-cell showing contrast sensitivity of the fundamental-response component to a 0.2 cycle/deg sine-wave grating counterphased at 2 Hz. Superimposed on the contrast-sensitivity values is a sinusoidal curve of the form Sm \cdot cos ϕ , where ϕ is the spatial phase angle of the grating and Sm is the mean of the sensitivities at 0 and 180°. B: plot for a nonlinear W-cell showing contrast sensitivity of the second-harmonic response component to a 0.1 cycle/deg sine-wave grating counterphased at 2 Hz. The sensitivity values are shown as positive and negative because the frequency-doubled responses occurred at both temporal half-cycles of the stimulus (see text).

spatial phase. Contrast was then reduced until the threshold response of this frequency-doubled component was obtained. To determine sensitivity of the fundamental response component, the grating was always placed 90° from this component's null po-



FIG. 5. Peristimulus time histograms of responses to counterphased sine-wave gratings with Fourier analysis of responses. A: response of a linear W-cell to a 0.1 cycle/deg grating counterphased at 2 Hz and placed 90° from the cell's null position. The grating's contrast was 0.3 and 50 cycles are averaged for this histogram. The inset shows the relative amplitude of the response components. B: response histogram for same cell to same stimulus as in A, but phase shifted by 90° to the null position. Twenty stimulus cycles are averaged. Note the decrease in the absolute size of the fundamental-response component from A to B (see insets). C: response histogram for a nonlinear W-cell to a 0.1 cycle/deg grating of 0.53 contrast and 2 Hz temporal frequency. The grating was positioned 90° from the null position to clicit the fundamental response component. Fifty stimulus cycles are averaged. Again, the inset shows the relative amplitudes for the Fourier components of the response. D: response histogram of same cell and to same stimulus as in C, but phase shifted by 90°. Fifty stimulus cycles are averaged. The major change from C to D is a reduction in the magnitude of the fundamental response component (see insets). The inset in B is scaled relative to that in A; and the inset in C is scaled relative to that in D.

sition, and two strategies were used. First, if the fundamental response component was more sensitive than was the second-harmonic component at the particular spatial and temporal frequency used, contrast was reduced until the fundamental component's threshold response was achieved. Second, if the second-harmonic response component was the more sensitive, contrast was reduced until the response at both stimulus half-cycles was equaled, since the presence of a fundamental component would create a larger response at one half-cycle than the other.

FUNDAMENTAL AND SECOND-HARMONIC RE-SPONSES. Figure 5 illustrates for two Wcells responses that characterize one as linear and the other as nonlinear. Figure 5A, *B* shows, for a typical linear W-cell, two peristimulus histograms of response rate versus time during one counterphase cycle of the stimulus. The stimulus had been shifted by 90° in spatial phase between histograms so that Fig. 5A depicts the maximum response and Fig. 5B the response at the null position. These responses were Fourier analyzed to determine the relative strength of responses at the fundamental temporal frequency and at higher harmonics of this frequency (insets). The ratio of the fundamental to second-harmonic component is 100 in Fig. 5A and 1.79 in Fig. 5B. This difference is due mainly to a dramatic loss of response at the fundamental frequency from Fig. 5Ato B. The response strength of the secondharmonic component lies in the background level in both cases.

Figure 5C, D depicts analogous responses for a tpical nonlinear W-cell. Again, the stimuli were separated by 90° in spatial phase between the two histograms. The responses of these cells are characterized by both a strong second-harmonic component that is independent of spatial phase as well as a phase-dependent response at the fundamental frequency. Figure 5C shows the maximum fundamental responses, and Fig. 5D shows the response at the null position for the fundamental component. The ratio of the fundamental response component to the second-harmonic response component is 1.34 for Fig. 5C and 0.22 for Fig. 5D. The magnitude of the fundamental response is reduced to the background level in Fig. 5Dwhile the second-harmonic component in both phase positions is quite large.

For X- and Y-cells in the retina and the lateral geniculate nucleus, the presence of a null position or of frequency-doubled responses at spatial frequencies that approach the spatial resolution of these cells (18) provides a reliable measure of the linearity of summation. The spatial resolution of W-cells in the lateral geniculate nucleus is generally much lower than that of X- or Y-cells (see below, and for example, Refs. 23, 35), and most nonlinear W-cells show clear phase-in-



FIG. 6. Spatial and temporal contrast-sensitivity functions of fundamental and second-harmonic response components. For each W-cell, the fundamental response component was assessed with a grating positioned at 90° from its null position, and the second-harmonic component was assessed at the fundamental component's null position. A: spatial function for a linear W-cell to a 2-Hz sine-wave grating. This cell had an off-center receptive field located 26.5° from the area centralis with a center 3° in diameter. B: temporal function for same cell as in A to a 0.1 cycle/deg sine-wave grating. C: spatial functions for a nonlinear W-cell to a 2-Hz sine-wave grating. This cell had an off-center receptive field located 17° from the area centralis with a center 5° in diameter. D: temporal functions for same cell as in C to a 0.05 cycle/deg sine-wave grating.

dependent, second-harmonic responses at spatial frequencies as low as 0.05 cycle/deg.

Contrast-sensitivity functions

We measured spatial and temporal contrast-sensitivity functions for seven linear and eight nonlinear W-cells. This was done by plotting contrast sensitivity as a function of spatial and temporal frequency. Furthermore, for 10 linear and 12 nonlinear W-cells, we measured spatial resolution at 0.84 contrast as the highest spatial frequency at 2 Hz to which the cell responded. For 10 linear and 9 nonlinear W-cells, we measured temporal resolution at 0.84 contrast as the highest temporal frequency at any spatial frequency (down to 0.05 cycle/deg) to which the cell responded.

Figure 6A, B shows spatial and temporal contrast-sensitivity functions for a typical linear W-cell, and Fig. 6C, D does likewise for a typical nonlinear W-cell. For the linear W-cell, no nonlinear or second-harmonic response component was detected, and the contrast-sensitivity functions decline monotonically with increasing spatial or temporal frequency (Fig. 6A, B; see also Fig. 10A).

Contrast-sensitivity measurements are more complicated in nonlinear W-cells because of the presence of appreciable linear or fundamental-response components in addition to the nonlinear or second-harmonic components (Fig. 6C, D). In seven of the eight nonlinear cells studied in this manner at a low temporal frequency (2 Hz), the nonlinear component was more sensitive than was the linear component at all spatial frequencies tested (Fig. 6C, see also Fig. 10B). The temporal contrast-sensitivity functions of these seven nonlinear W-cells are especially complicated. At low spatial frequencies, temporal-contrast sensitivity of the linear-response component, compared to that of the nonlinear component, was less at lower temporal frequencies but greater at higher temporal frequencies (Fig. 6D). Furthermore, the linear response component consistently exhibited a sensitivity loss at lower temporal frequencies, whereas no such loss was seen for the nonlinear component.

For the eighth nonlinear W-cell, contrastsensitivity functions were somewhat different at the same low temporal frequency (2 Hz) from the other seven and resembled

those of Y-cells (see, for example, Fig. 10B; see also Refs. 19, 36). That is, spatial-contrast sensitivity of the linear response component was greater than that of the nonlinear component at lower, but not higher spatial frequencies. Consequently, at lower spatial frequencies, temporal-contrast sensitivity for the linear response component was greater than that for the nonlinear component at all temporal frequencies and displayed no attenuation at low temporal frequencies. This eighth nonlinear W-cell may not be fundamentally different from the other seven, since it is possible that at other temporal frequencies all of the eight nonlinear Wcells, would exhibit similar spatial functions.

Spatial and temporal resolution

Figure 7A, B shows the distribution of spatial and temporal resolution values for W-cells. Linear and nonlinear W-cells are separately indicated. For nonlinear W-cells, spatial resolution was determined by the nonlinear response component (cf. Fig. 6Cand 10 B), but temporal resolution for these cells was determined by the linear component (cf. Fig. 6D). The temporal resolutions of the linear and nonlinear W-cells, therefore, represent the same aspect of response (i.e., the fundamental component), while the spatial resolutions of the two types represent different aspects of response (i.e., the fundamental versus second-harmonic component). In any case, no statistically reliable difference was seen between linear and nonlinear W-cells for either spatial or temporal resolution (P > 0.1 for each comparison on either a Mann-Whitney U test or an F test).

We measured both spatial- and temporalcontrast sensitivity for seven linear and eight nonlinear W-cells. Figure 8A plots for these cells the relationship between spatial and temporal resolution. As in Fig. 7, the values for the nonlinear W-cells represent secondharmonic responses for spatial resolution and fundamental responses for temporal resolution. W-cells that were generally more responsive or sensitive to visual stimuli exhibited higher spatial and temporal resolution.

The apparently equivalent spatial-resolution values of linear and nonlinear W-cells shown in Fig. 7A and 8A are somewhat misleading because the comparisons involve different response components (i.e., fundamental versus second harmonic). If, instead, the fundamental-response components are compared between linear and nonlinear Wcells, a different picture emerges. In only four of the nonlinear W-cells was the spatial resolution of the fundamental response component above 0.05 cycle/deg, and these are shown in Fig. 8B. Consequently, the spatial resolution of the fundamental response component of nonlinear W-cells is lower than either the second-harmonic component of these cells (P < 0.01 on a Mann-Whitney U test) or the spatial resolution of the linear W-cells (P < 0.01 on a Mann-Whitney U test).



FIG. 7. Frequency histograms of spatial and temporal resolutions. These are, respectively, the highest spatial frequency at 0.84 contrast and 2 Hz to which the cell responded and the highest temporal frequency at 0.84 contrast and any spatial frequency to which the cell responded. A: spatial resolution for 10 linear and 12 nonlinear W-cells. B: temporal resolution for 10 linear and 9 nonlinear W-cells.



FIG. 8. Spatial resolution of W-cells as a function of temporal resolution and receptive-field center diameter. *A*: spatial versus temporal resolution. The temporal resolution for each nonlinear W-cell was derived from the fundamental response component, but the spatial resolution was derived from the second-harmonic component (see text for details). *B*: spatial resolution versus the inverse of receptive-field center diameter. The resolution values represented for nonlinear W-cells include only their fundamental response components, and only the four nonlinear W-cells shown displayed resolution values above 0.05 cycles/deg (see text for details).

Figure 8B shows the surprising lack of correlation for W-cells between receptive-field center size and spatial resolution of the fundamental response component. No correlation was found for either linear or non-linear W-cells (P > 0.1 for either group). By contrast, for geniculate X- and Y-cells, there is a strong inverse relationship between these parameters of field size and spatial resolution (23, 36). Such a relationship for W-cells to have

much larger field centers than do linear Wcells, but Fig. 2 reveals no such difference. The conclusion derived from Fig. 8*B* must be qualified due to the uncertainty noted above in deriving precise receptive-field measurements for W-cells.

Contrast sensitivity: a comparison among W-, X-, and Y-cells

During the same experiments in which we studied W-cells in the C-laminae, we also recorded X- and Y-cells in the A-laminae and the top of lamina C. We measured spatial contrast-sensitivity functions, usually at a temporal frequency of 2 Hz, for 9 X- and 14 Y-cells. From these functions, we noted the peak contrast sensitivity. Figure 9 illustrates the comparisons of peak sensitivity both between linear W-cells and X-cells as well as between nonlinear W-cells and Ycells. For the latter comparison, peak sensitivity for both the fundamental and second-harmonic response components are illustrated, although only four of the nonlinear W-cells exhibited detectable fundamental responses at 2 Hz.

The W-cells were generally 2 octaves less sensitive than were the X- or Y-cells. These differences were clear between linear Wcells and X-cells (P < 0.001 on a Mann-Whitney U test), between the fundamental components of nonlinear W-cells and Y-cells (P < 0.001 on a Mann-Whitney U test), and between the second-harmonic components of nonlinear W-cells and Y-cells (P < 0.01on a Mann-Whitney U test). Furthermore, there was no overlap in sensitivity between either linear W-cells and X-cells or the fundamental response components of nonlinear W-cells and Y-cells. No difference, however, was seen in peak sensitivity between linear and nonlinear W-cells (P > 0.1 on a Mann-Whitney U test). Presumably the relatively poor contrast sensitivity of W-cells relates to their sluggish responsiveness to visual stimuli.

As a final comparison, Fig. 10 shows contrast-sensitivity functions derived at 2 Hz for typical W-cells and an X- and Y-cell. The W-cells are different from those illustrated in Fig. 6. Figure 10*A* compares a linear Wcell with an X-cell. Not only did the W-cell display generally poorer resolution and sensitivity than did the X-cell, but also the



FIG. 9. Frequency histograms of peak contrast-sensitivity values at a temporal frequency of 2 Hz for W-, X-, and Y-cells. A: values for linear cells (linear W-cells and X-cells). Arrows below each histogram indicate the mean values. B: values for nonlinear cells (nonlinear W-cells and Y-cells). Values for the fundamental and second-harmonic components are separately shown, and only four of the nonlinear cells had detectable fundamental components at 2 Hz. Arrows below each histogram denote the mean values, open for the fundamental components were assigned values of zero sensitivity for the determination of the mean value for this component.

shapes of the functions differed. The W-cell acted as a "low-pass" filter, since it displayed a gradually decreasing sensitivity for higher spatial frequencies, while the X-cell acted as a "band-pass" spatial filter, since it was most sensitive to middle spatial frequencies and was less so to lower and higher ones. Figure 10*B* compares the fundamental and second-harmonic response components for a nonlinear W-cell and a Y-cell. These functions generally matched those of low-pass spatial filters. However, not only were there



FIG. 10. Spatial-contrast sensitivity functions at 2 Hz. A: functions for a linear W-cell and an X-cell. B: functions for a nonlinear W-cell and a Y-cell. The fundamental and second-harmonic components are separately indicated.

differences in resolution and sensitivity between the W- and Y-cell, but there also existed differences in the relationship of the two response components for each cell. For the Y-cell, the fundamental component was more sensitive at lower spatial frequencies, and the second-harmonic component dominated at higher ones. For the W-cell, the second-harmonic component was more sensitive at all spatial frequencies tested. The greatest difference in contrast sensitivity between the nonlinear W-cell and Y-cell was evident in their fundamental response components; their second-harmonic response components displayed less of a difference from one another (Figs. 9B and 10B).

Other differences between linear and nonlinear W-cells

Although the W-cells in this study could be reliably and easily identified as linear or nonlinear, as described above, only one difference could be detected in response properties between linear and nonlinear W-cells. The one difference concerns response dynamics: each of the 5 tonic W-cells (i.e., possibly related to sluggish-sustained ganglion cells in the retina; see Refs. 5, 6, 8) was linear in spatial and temporal summation, whereas the phasic W-cells (i.e., possibly related to sluggish-transient ganglion cells in the retina; see Refs. 5, 6, 8) were either linear (7 cells) or nonlinear (14 cells) in spatial and temporal summation. The two on-off center W-cells were phasic and nonlinear. Otherwise, no difference between linear and nonlinear W-cells was seen for latency to optic chiasm stimulation (Fig. 1), receptive-field center size (Fig. 2), sensitivity to visual stimuli (e.g., Figs. 6 and 9), or overall spatial or temporal resolution (Fig. 7).

DISCUSSION

A major conclusion of this study is that W-cells found in the C-laminae of the cat's lateral geniculate nucleus can be reliably subdivided into types that exhibit linear or nonlinear spatial and temporal summation. These occur in roughly equal numbers. The linear or nonlinear classification does not appear to correlate with most other response properties (see RESULTS). The functional significance of this linear and nonlinear distinction and its implications for cell classification are considered below.

Cell classification

TERMINOLOGY. We have used the term Wcell to describe those cells in the geniculate C-laminae that do not exhibit X- or Y-cell properties. As noted in the INTRODUCTION, the main justification for this terminology is our desire to avoid new nomenclature without sufficient data for doing so. We thus used existing terminology (cf. Ref. 39).

However, we emphasize that W-cell as a descriptive phrase should not carry the same

implication as X-cell or Y-cell. X- and Ycells in retina and the lateral geniculate nucleus are each relatively homogeneous cell classes. Furthermore, geniculate X- or Ycells closely resemble their retinal counterparts in terms of reponse properties. Indeed, Cleland et al. (4) showed that each geniculate X- or Y-cell typically receives direct retinal input from one or very few ganglion cells of the same functional class.

The situation regarding W-cells is much less clear, partly because of the functional heterogeneity of the retinal W-cells and partly because so little is known of the properties of retinal or geniculate W-cells. It is not clear what subclasses of retinal W-cells innervate the C-laminae cells described in the present study. The observation that these C-lamina cells are fairly homogeneous compared to the multiplicity of response types described for retina suggests that a fairly small subset provides the retinogeniculate innervation to the C-laminae. However, without more detailed knowledge of the response properties of retinal and geniculate W-cells, the nature of the retinogeniculate circuitry is barely open to speculation. Indeed, it is possible that many W-cells of the C-laminae do not receive monosynaptic input from the optic tract, since their long response latencies to optic chiasm stimulation raise the possibility that many of these cells are polysynaptically activated. Finally, the C-laminae receive inputs from visual structures (e.g., the superior colliculus and nucleus of the optic tract) that do not innervate the A-laminae (16, 17, 27). For all of these reasons, circuitry in the W-cell pathway may be quite different from that in the X- and Y-cell pathways.

ARE W-CELLS A CLASS DISTINCT FROM X-AND Y-CELLS? X- and Y-cells in the retina were originally distinguished by Enroth-Cugell and Robson (11) on the basis of linearity of spatial summation to visual stimuli, and this has been extended to the lateral geniculate nucleus (32). X-cells are linear and Ycells are nonlinear in much the same way as W-cells in the lateral geniculate nucleus have been termed linear or nonlinear in the present paper. This raises the possibility that linear W-cells really represent a subclass of X-cells and nonlinear W-cells, a subclass of Y-cells. This, in turn, would require that linear W-cell properties be continuous with those of X-cells and, likewise, for nonlinear W-cells and Y-cells.

We feel that the present evidence, although incomplete, rather convincingly argues that W-cells, at least in the geniculate C-laminae, are quite distinct from geniculate X- and Y-cells. Indeed, linear and nonlinear W-cells resemble each other much more than either resembles X- or Y-cells, although more detailed quantitative comparisons among W-, X-, and Y-cells still need to be made. The features that seem to distinguish W-cells from X- and Y-cells in the lateral geniculate nucleus include axonal conduction velocity of retinal afferents (present study and Refs. 8, 43, 44), responsiveness and contrast sensitivity to visual stimuli (present study and Refs. 8, 43), receptivefield size (present study and Refs. 8, 43), spatial resolution (present study), contrastsensitivity functions (present study), neuronal distribution and morphology (8, 12, 13, 13)38, 43), and geniculocortical projection patterns (8, 10, 14, 15, 25, 26, 28, 30, 37, 43). W-cells in the retina differ from retinal Xand Y-cells along most of these same parameters as well (1, 5-7, 40, 41). Although certain of these properties exhibit limited overlap with some X- or Y-cell properties, a consideration of all of these features rather clearly distinguishes W-cells as a neuronal class quite different from X- or Y-cells.

DO LINEAR AND NONLINEAR W-CELLS REP-RESENT SEPARATE CLASSES? Although we feel that W-cells should not be placed in Xor Y-cell classes, we are not suggesting that W-cells are a single class, even in the C-laminae of the lateral geniculate nucleus. We cannot yet determine if more than one class is needed to encompass this rather heterogeneous group. In the present study, we have concentrated on the division of W-cells into linear and nonlinear types.

We emphasize that the terminology of linear and nonlinear, as applied to these neurons, refers only to limited linearity or nonlinearity of spatial and temporal summation measured with sine-wave gratings as the visual stimuli. This terminology does not encompass other aspects of the responses of these cells. For example, all neurons possess a number of nonlinearities of response that are independent of the spatial and temporal pooling that we have investigated in the present paper (e.g., the rectified action potential, refractory period, etc.). Similarly, many nonlinear W-cells and all Y-cells have response components that exhibit linearity of spatial and temporal summation.

By our criteria of identification, linear and nonlinear W-cells respond quite differently from one another to sine-wave gratings. That is, the fundamental-response component of linear W-cells exhibits greater contrast sensitivity and spatial resolution than does that of nonlinear W-cells. One obvious consequence of this is that the linear W-cells are more sensitive to the spatial phase, and thus position, of the stimulus than are the nonlinear W-cells. Similarly, the linear W-cells more faithfully represent temporal aspects of the stimulus in their response dynamics than do nonlinear W-cells.

Nevertheless, W-cells that exhibit linear and nonlinear spatial and temporal summation seem rather similar along other functional dimensions (response latency to optic chiasm stimulation, receptive-field size, etc.). There also appears to be no obvious morphological difference between linear and nonlinear W-cells, as judged by intracellular HRP injections placed into the physiologically identified types (L. R. Stanford, M. J. Friedlander, and S. M. Sherman, unpublished observation). Therefore, differences in spatial and temporal summation do not at present seem to justify the division of linear and nonlinear W-cells into two classes. Much more data are needed to resolve this issue.

Concluding remarks

The W-cells in the C-laminae of the cat's lateral geniculate nucleus seem quite different from X- and Y-cells, and at least two types have been noted. We have termed these linear and nonlinear W-cells on the basis of their spatial and temporal summation properties. The large areas of cortex that are directly innervated by the W-cell geniculocortical pathway (e.g., Refs. 10, 28) suggest an important functional role for these W-cells of the geniculate C-laminae. The poor contrast sensitivity, poor responsiveness, and poor spatial resolution that are characteristic features of these cells suggest that they function largely in the presence of large (i.e., low spatial frequency) stimuli of high contrast.

ACKNOWLEDGMENTS

We thank L. R. Stanford for assistance in some of the experiments, and Sally Gibson and Joan Sommermeyer for technical help.

This research was supported by Public Health Service Grant EY03038.

Received 29 June 1981; accepted in final form 14 December 1981.

REFERENCES

- 1. BOYCOTT, B. B. AND WÄSSLE, H. The morphological types of ganglion cells of the domestic cat's retina. J. Physiol. London 240: 397-419, 1974.
- 2. BULLIER, J. AND NORTON, T. T. X and Y relay cells in cat lateral geniculate nucleus: quantitative analysis of receptive-field properties and classification. J. Neurophysiol. 42: 244-273, 1979.
- 3. BULLIER, J. AND NORTON, T. T. Comparison of receptive-field properties of X and Y ganglion cells with X and Y lateral geniculate cells in the cat. J. Neurophysiol 42: 274-291, 1979.
- CLELAND, B. G., DUBIN, M. W., AND LEVICK, W. R. Sustained and transient neurones in the cat's retina and lateral geniculate nucleus. J. Physiol. London 217: 473-496, 1971.
- CLELAND, B. G. AND LEVICK, W. R. Brisk and sluggish concentrically organised ganglion cells in the cat's retina. J. Physiol. London 240: 421-456, 1974.
- 6. CLELAND, B. G. AND LEVICK, W. R. Properties of rarely encountered types of ganglion cells in the

cat's retina and an overall classification. J. Physiol. London 240: 457-492, 1974.

- CLELAND, B. G. LEVICK, W. R., AND WÄSSLE, H. Physiological identification of a morphological class of cat retinal ganglion cells. J. Physiol. London 248: 151–171, 1975.
- CLELAND, B. G., MORSTYN, R., WAGNER, H. G., AND LEVICK, W. R. Long-latency retinal input to lateral geniculate neurones of the cat. *Brain Res.* 91: 306-310, 1975.
- 9. DERRINGTON, A. M. AND FUCHS, A. F. Spatial and temporal properties of X- and Y-cells in the cat lateral geniculate nucleus. J. Physiol. London 293: 347-364, 1979.
- DREHER, B., LEVENTHAL, A. G., AND HALE, P. T. Geniculate inut to cat visual cortex: a comparison of area 19 with areas 17 and 18. J. Neurophysiol. 44: 804-826, 1980.
- 11. ENROTH-CUGELL, C. AND ROBSON, J. G. The contrast sensitivity of retinal ganglion cells of the cat. J. Physiol. London 187: 517-552, 1966.

- FRIEDLANDER, M. J., LIN, C.-S., AND SHERMAN, S. M. Structure of physiologically identified X- and Y-cells in the cat's lateral geniculate nucleus. *Science* 204: 1114-1117, 1979.
- FRIEDLANDER, M. J., LIN, C.-S., STANFORD, L. R., AND SHERMAN, S. M. Morphology of functionally identified neurons in the lateral geniculate nucleus of the cat. J. Neurophysiol. 46: 80-129, 1981.
- 14. FUKUDA, Y. AND STONE, J. Retinal distribution and central projections of Y-, X-, and W-cells of the cat's retina. J. Neurophysiol. 37: 749-772, 1974.
- GEISERT, E. E. Cortical projections of the lateral geniculate nucleus in the cat. J. Comp. Neurol. 190: 793-812, 1980.
- GRAHAM, J. An autoradiographic study of the efferent connections of the superior colliculus in the cat. J. Comp. Neurol. 173: 629-654, 1977.
- GRAYBIEL, A. M. AND BERSON, D. M. Autoradiographic evidence for a projection from the pretectal nucleus of the optic tract to the dorsal lateral geniculate complex in the cat. *Brain Res.* 195: 1–12, 1980.
- HOCHSTEIN, S. AND SHAPLEY, R. M. Quantitative analysis of retinal ganglion cell classifications. J. Physiol. London 262: 237-264, 1976.
- HOCHSTEIN, S. AND SHAPLEY, R. M. Linear and nonlinear subunits in Y cat retinal ganglion cells. J. Physiol. London 262: 265-284, 1976.
- HOFFMANN, K.-P., STONE, J., AND SHERMAN, S. M. Relay of receptive-field properties in dorsal lateral geniculate nucleus of the cat. J. Neurophysiol. 35: 518-531, 1972.
- IKEDA, H. AND WRIGHT, M. J. Receptive field organization of "sustained" and "transient" retinal ganglion cells which subserve different functional roles. J. Physiol. London 227: 769-800, 1972.
- KRATZ, K. E., WEBB, S. V., AND SHERMAN, S. M. Electrophysiological classification of X- and Y-cells in the cat's lateral geniculate nucleus. *Vision Res.* 18: 489-492, 1978.
- LEHMKUHLE, S., KRATZ, K. E., MANGEL, S. C., AND SHERMAN, S. M. Spatial and temporal sensitivity of X- and Y-cells in dorsal lateral geniculate nucleus of the cat. J. Neurophysiol. 43: 520-541, 1980.
- 24. LENNIE, P. Parallel visual pathways. Vision Res. 20: 561-594, 1980.
- LEVENTHAL, A. G. Evidence that the different classes of relay cells of the cat's lateral geniculate nucleus terminate in different layers of the striate cortex. *Exp. Brain Res.* 237: 349-372, 1979.
- LEVENTHAL, A. G., KEENS, J., AND TORK, I. The afferent ganglion cells and cortical projections of the retinal recipient zone (RRZ) of the cat's "pulvinar complex." J. Comp. Neurol. 194: 535-554, 1980.
- NIIMI, K., MIKI, M., AND KAWAMURA, S. Ascending projections of the superior colliculus in the cat. Okajimas Fol. Anat. Jpn. 47: 269-287, 1970.
- 28. RACZKOWSKI, D. AND ROSENQUIST, A. C. Con-

nections of the parvocellular C laminae of the dorsal lateral geniculate nucleus with the visual cortex in the cat. *Brain Res.* 199: 447-451, 1980.

- 29. RODIECK, R. W. Visual pathways. Ann. Rev. Neurosci. 2: 193-225, 1979.
- ROWE, M. H. AND STONE, J. Properties of ganglion cells in the visual streak of the cat's retina. J. Comp. Neurol. 169: 99-126, 1976.
- 31. ROWE, M. H. AND STONE, J. Naming of neurones. Brain Behav. Evol. 14: 185-216, 1977.
- SHAPLEY, R. AND HOCHSTEIN, S. Visual spatial summation in two classes of geniculate cells. *Nature London* 256: 411-413, 1975.
- SHERMAN, S. M. The functional significance of Xand Y-cells in normal and visually deprived cats. *Trends Neurosci.* 2: 192–195, 1979.
- 34. SHERMAN, S. M. Parallel pathways in the cat's geniculocortical system: W-, X-, and Y-cells. In: *Changing Concepts in the Nervous System*, edited by A. Morrison and P. Strick. New York: Academic, 1982, p. 337-359.
- 35. SO, Y.-T. AND SHAPLEY, R. M. Spatial properties of X and Y cells in the lateral geniculate nucleus of the cat and conduction velocities of their inputs. *Exp. Brain Res.* 36: 533-550, 1979.
- 36. SO, Y.-T. AND SHAPLEY, R. Spatial tuning of cells in and around lateral geniculate nucleus of the cat: X and Y relay cells and perigeniculate interneurons. J. Neurophysiol. 45: 107-120, 1981.
- SPEAR, P. D., SMITH, D. C., AND WILLIAMS, L. L. Visual receptive-field properties of single neurons in cat's ventral lateral geniculate nucleus. J. Neurophysiol. 40: 390-409, 1977.
- STANFORD, L. R., FRIEDLANDER, M. J., AND SHERMAN, S. M. Morphology of physiologically identified W-cells in the C laminae of the cat's lateral geniculate nucleus. J. Neurosci. 1: 578-584, 1981.
- 39. STONE, J., DREHER, B., AND LEVENTHAL, A. Hierarchical and parallel mechanisms in the organization of visual cortex. *Brain Res. Rev.* 1: 345–394, 1979.
- STONE, J. AND FUKUDA, Y. Properties of cat retinal ganglion cells: a comparison of W-cells with X- and Y-cells. J. Neurophysiol. 37: 722–748, 1974.
- STONE, J. AND HOFFMANN, K.-P. Very slow-conducting ganglion cells in the cat's retina: a major, new functional type? *Brain Res.* 43: 610–616, 1972.
- 42. SUR, M., STANFORD, L. R., AND SHERMAN, S. M. W-cells in the C laminae of the cat's lateral geniculate nucleus: contrast sensitivity and other response measures. Soc. Neurosci. Abstr. 7: 25, 1981.
- WILSON, P. D., ROWE, M. H., AND STONE, J. Properties of relay cells in the cat's lateral geniculate nucleus. A comparison of W-cells with X- and Y-cells. J. Neurophysiol. 39: 1193–1209, 1976.
- 44. WILSON, P. D. AND STONE, J. Evidence of W-cell input to the cat's visual cortex via the C-laminae of the lateral geniculate nucleus. *Brain Res.* 92: 472-478, 1975.