

Non-invasive optical neuromonitoring of the temperature-dependence of cerebral oxygen metabolism during deep hypothermic cardiopulmonary bypass in neonatal swine

Tiffany S Ko^{1,2,3}, Constantine D Mavroudis⁴, Wesley B Baker³, Vincent C Morano², Kobina Mensah-Brown³, Timothy W Boorady³, Alexander L Schmidt⁵, Jennifer M Lynch⁶, David R Busch^{7,8}, Javier Gentile⁹, George Bratinov⁶, Yuxi Lin⁶, Sejin Jeong⁶, Richard W Melchior¹⁰, Tami M Rosenthal¹⁰, Brandon C Shade¹⁰, Kellie L Schiavo¹⁰, Rui Xiao¹¹, J William Gaynor⁹, Arjun G Yodh², Todd J Kilbaugh⁶ and Daniel J Licht³

Abstract

Management of deep hypothermic (DH) cardiopulmonary bypass (CPB), a critical neuroprotective strategy, currently relies on non-invasive temperature to guide cerebral metabolic suppression during complex cardiac surgery in neonates. Considerable inter-subject variability in temperature response and residual metabolism may contribute to the persisting risk for postoperative neurological injury. To characterize and mitigate this variability, we assess the sufficiency of conventional nasopharyngeal temperature (NPT) guidance, and in the process, validate combined non-invasive frequency-domain diffuse optical spectroscopy (FD-DOS) and diffuse correlation spectroscopy (DCS) for direct measurement of cerebral metabolic rate of oxygen ($CMRO_2$). During CPB, $n=8$ neonatal swine underwent cooling from normothermia to 18°C, sustained DH perfusion for 40 min, and then rewarming to simulate cardiac surgery. Continuous non-invasive and invasive measurements of intracranial temperature (ICT) and $CMRO_2$ were acquired. Significant hysteresis ($p < 0.001$) between cooling and rewarming periods in the NPT versus ICT and NPT versus $CMRO_2$ relationships were found. Resolution of this hysteresis in the ICT versus $CMRO_2$ relationship identified a crucial insufficiency of conventional NPT guidance. Non-invasive $CMRO_2$ temperature coefficients with respect to NPT ($Q_{10} = 2.0$) and ICT ($Q_{10} = 2.5$) are consistent with previous reports and provide further validation of FD-DOS/DCS $CMRO_2$ monitoring during DH CPB to optimize management.

¹Department of Bioengineering, University of Pennsylvania, Philadelphia, PA, USA

²Department of Physics and Astronomy, University of Pennsylvania, Philadelphia, PA, USA

³Division of Neurology, Children's Hospital of Philadelphia, Philadelphia, PA, USA

⁴Division of Cardiovascular Surgery, Department of Surgery, Hospital of the University of Pennsylvania, Philadelphia, PA, USA

⁵Department of Biology, James Madison University, Harrisonburg, VA, USA

⁶Department of Anesthesiology and Critical Care Medicine, Children's Hospital of Philadelphia, Philadelphia, PA, USA

⁷Department of Anesthesiology & Pain Management, University of Texas Southwestern, Dallas, TX, USA

⁸Department of Neurology & Neurotherapeutics, University of Texas Southwestern, Dallas, TX, USA

⁹Division of Cardiothoracic Surgery, Children's Hospital of Philadelphia, Philadelphia, PA, USA

¹⁰Department of Perfusion Services, Cardiac Center, Children's Hospital of Philadelphia, Philadelphia, PA, USA

¹¹Department of Pediatrics, Division of Biostatistics, Children's Hospital of Philadelphia, Philadelphia, PA, USA

Corresponding author:

Tiffany S Ko, Laboratory for the Research on the Structure of Matter, University of Pennsylvania, 3231 Walnut St., Philadelphia, PA 19104, USA.
 Email: tiko@seas.upenn.edu

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Introduction

Deep hypothermia (DH) is an important neuroprotective therapy used during cardiopulmonary bypass (CPB) in an attempt to mitigate hypoxic-ischemic brain injury by suppressing cellular metabolic demand in neonates with congenital heart disease during complex cardiac repairs.¹ Over the last two decades, survival rates for these children have substantially improved²; however, the incidence of neurological injury has remained constant and, in some cases, has resulted in developmental delays and lifelong neurological deficits.³ Despite widespread use of DH CPB, uncertainty predominates regarding optimal temperature management due to hitherto poorly defined individual cerebral metabolic responses to hypothermia.

Real-time neuromonitoring is needed to address a key challenge for DH protocols by confirming, on a patient-by-patient basis, that the suppression of metabolism is sufficient to prevent adverse neurological sequelae during procedural cerebral ischemia.^{1,4-6} Decreased cerebral blood flow (CBF), oxygen extraction, and metabolism in response to DH have been widely demonstrated, but significant inter-subject variability in temperature-response has also been observed.⁷ Specifically, the use and value of conventional core temperature guidance for assessment of the adequacy of metabolic suppression during DH have been questioned.⁸⁻¹² Systematic study of core temperature, brain temperature, and residual cerebral metabolism as neurological risk factors is needed. Such studies are hindered by a lack of noninvasive, cerebral metabolic monitoring tools suitable for the operative environment. If this limitation can be ameliorated, then assessment of current neuroprotection strategies, and development of new personalized strategies to optimize neurological outcomes, should be possible for these at-risk children.

Multimodal neuromonitoring, including clinical continuous-wave near-infrared spectroscopy (CW NIRS) for cerebral oxygen saturation and transcranial Doppler ultrasound (TCD) for CBF-velocity, has shown evidence of improving post-operative neurological complications.¹³ The quantitative uncertainty of CW NIRS and the logistical difficulty of employing and interpreting TCD in the operating room have prevented their combined use for routine metabolic monitoring and likely impacted efficacy of goal-directed

therapy.¹⁴ By contrast, the combination of frequency- or time-domain diffuse optical spectroscopy (FD-DOS, TD-DOS), to measure *absolute* cerebral tissue oxygen saturation (StO₂, %), with diffuse correlation spectroscopy (DCS) to measure CBF, enables a compact, all-optical method for continuous non-invasive monitoring of cerebral metabolism. This approach has been demonstrated in vulnerable pediatric populations outside the operating room.¹⁵⁻²⁸ Although recent work has established intraoperative feasibility for some of this technology,^{20,29} non-invasive diffuse optical measurements of CBF and metabolism have never been validated against invasive monitoring during the profound physiologic and temperature changes induced by DH CPB and subsequent rewarming.

Here we carry out an observational study of concurrent conventional monitoring of nasopharyngeal temperature (NPT) alongside invasive intracranial temperature (ICT) and invasive and non-invasive (i.e. diffuse optical) measurements of cerebral oxygen metabolism during a simulated cardiac surgical procedure using DH CPB with subsequent rewarming in neonatal swine. The sufficiency of NPT guidance is examined with respect to ICT, and the temperature-dependence of cerebral oxygen metabolism is assessed with respect to both modalities using high temporal resolution *in vivo* sampling methods. Non-invasive diffuse optical measurements are compared directly with invasive measurements, and similarities and differences of measured parameters are identified and understood.

Methods

Neonatal, female Yorkshire swine ($n=8$, 6–10 days old, 3–5 kg) were continuously monitored during CPB from induction of DH through recovery to normothermia. All procedures were approved by the CHOP Institutional Animal Care and Use Committee, performed in strict accordance with the NIH Guide for the Care and Use of Laboratory Animals, and reported according to the ARRIVE guidelines (<https://www.nc3rs.org.uk/arrive-guidelines>).

Selection of animal model

Pediatric large-animal model studies describing *CMRO*₂ temperature-dependence during DH CPB have been reported in dogs³⁰ and pigs using sparse

sampling methods.^{12,31–36} The neonatal swine model offers comparable anatomical size and cortical maturation with the human neonate, as well as excellent intersection of DH CPB and diffuse optical neuro-monitoring literature.^{23,37–39}

Neuromonitoring

Non-invasive and invasive neuromonitoring were placed and secured following anesthetic induction and intubation as detailed in Supplementary Materials, Section S.1.⁴⁰ Continuous non-invasive measurements comprised NPT (°C), for guidance of hypothermic therapy, and frequency-domain diffuse optical spectroscopy (FD-DOS) and DCS. NPT, which has been found to be a close surrogate of parenchymal brain temperature,⁴¹ was measured using a thermistor (Level 1 Thermistor GP Probe, Smith Medical) inserted 5 cm to the mid-nasopharynx and sutured into place. FD-DOS/DCS measurements were acquired in the left frontal cortex via an optical probe sutured to the left forehead (Figure 1). Continuous invasive neuromonitoring was performed symmetrically on the contralateral hemisphere through small burr holes made over the right frontal cortex (10 mm paramedian, 10 mm anterior to the coronal suture; Figure 1). Subcortical intracranial oxygen tension (P_{btO_2} , mmHg) and cortical ICT (°C) were measured 15 mm and 5 mm, respectively, below the cortical surface (Licox CC1-P1, Integra LifeSciences) near the junction of grey and subcortical white matter. Relative cortical cerebral blood flow (rCBF LD, %) was measured with a laser Doppler probe (PeriFlux 5000, Perimed Inc.) secured to the dura matter. A double lumen, 4F central venous catheter was placed in the superior vena cava and advanced into the internal jugular bulb for invasive discontinuous sampling of cerebral venous drainage.

DH protocol

Protocols for institution of DH CPB are detailed in Supplementary Materials, Section S.1 and closely mirrored clinical practice at our institution (Figure 2). Subjects were stabilized on CPB at normothermia (NPT=37°C) and baseline measurements acquired for 5 min. Guided by NPT, subjects were then cooled at a target rate of 1°C per minute to 18°C in order to characterize cerebral metabolism across the full range of temperatures currently used in neonates.^{42,43} Subjects were then maintained on continuous DH perfusion for 40 min. Rewarming to normothermia occurred at a target rate of 1°C per minute. Invasive arterial and venous blood gas sampling (0.3 cc of blood per draw from the bypass arterial outflow and jugular bulb, respectively) occurred at the start of cooling (i.e.

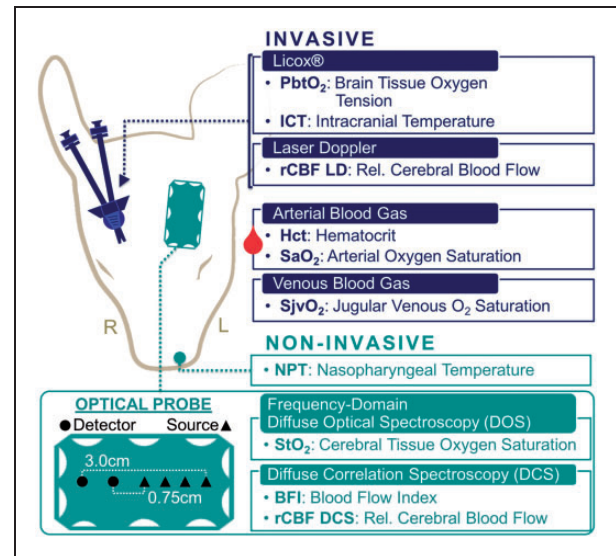


Figure 1. Neuromonitoring – Head placement of *non-invasive* (green; left hemisphere) and *invasive* (blue; right hemisphere) neuromonitoring technologies. A diagram showing details about the FD-DOS/DCS optical probe is given on the lower left (source positions, triangles; detector positions, circles).

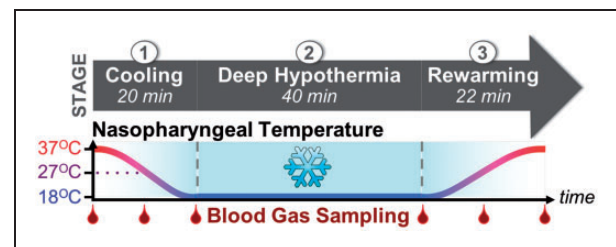


Figure 2. Temperature and blood gas sampling protocol – Each subject underwent three stages of treatment: (1) Induction of deep hypothermia (“Cooling”); (2) 40 min of continuous cold perfusion (“Deep Hypothermia”); (3) Recovery from deep hypothermia (“Rewarming”). Arterial and jugular venous blood gas sampling was performed at multiple time-points indicated by red droplets along the time-axis.

baseline, 37°C), midpoint of cooling (27°C), end of cooling (18°C), start of rewarming (18°C), midpoint of rewarming (27°C) and end of rewarming (37°C; Figure 2). Immediate analysis was facilitated through a point-of-care blood gas analyzer (GEM 3000, Instrumentation Laboratory). Blood pH was optimized by pH-stat management during cooling. During rewarming and while normothermic, blood pH was optimized by alpha-stat.

Diffuse optical techniques

Frequency-domain diffuse optical spectroscopy. Multi-distance FD-DOS was used to continuously measure

cerebral tissue oxygen saturation (StO_2 , %) and oxygen extraction fraction (OEF). FD-DOS employs radio-frequency intensity-modulated near-infrared light to quantify wavelength-dependent absorption and scattering properties of tissue.⁴⁴⁻⁴⁶

A customized, commercial instrument (Imagent, ISS Inc.), equipped with four 690, 725, 785, and 830 nm intensity-modulated (110 MHz) diode laser sources and two photomultiplier tube detectors, was coupled to the fiberoptic probe; source-detector separations ranged from 0.75 cm to 3 cm (Figure 1, lower left). For each subject, the source- and detector-fiber coupling coefficients to the tissue were estimated using a phantom-calibration approach⁴⁷ and used to correct continuous (10 Hz) AC intensity and phase data. Using a multi-distance linear fitting method, absolute absorption and scattering coefficients were calculated for each wavelength from these data. Coefficients were excluded if the linear fit Pearson correlation coefficient was <0.95 , and the data-point excluded if more than one of the four scattering or absorption coefficients were excluded. Assuming a cerebral water volume fraction of 75%,⁴⁸ the absolute cerebral tissue concentration of oxy-hemoglobin ($[HbO_2]$, $\mu\text{mol/L}$) and deoxy-hemoglobin ($[Hb]$, $\mu\text{mol/L}$) was quantified from the absorption coefficient⁴⁵ and StO_2 calculated as

$$StO_2(\%) = \frac{[HbO_2]}{[Hb] + [HbO_2]} \quad (1)$$

Importantly, the FD-DOS technique directly measures the tissue scattering coefficient and eliminates optical absorption measurement errors introduced by physiologic shifts in optical scattering,^{49,50} a parameter which cannot be determined by commercial CW NIRS.

DCS. DCS is a photon correlation technique that derives a CBF index (BFI, cm^2/s) from quantification of the rapid speckle intensity fluctuations of multiply scattered coherent near-infrared light induced by red blood cell motion.^{38,51,52} DCS measurements were made using a source-detector separation of 2.5 cm, wherein the detected light fields were sensitive to blood flow at an average depth of 1 cm below the scalp surface.⁵³

The DCS light source was a continuous-wave, long-coherence length ($>10\text{ m}$), 785 nm laser (RCL-080-785S, CrystaLaser, Inc.). A bundle of eight single-mode detection fibers coupled diffuse light emerging from the head onto two detection arrays of four single-photon-counting avalanche photodiodes (SPCM-AQ4C, Excelitas Technologies, Corp.). Calculation of the intensity autocorrelation curve for each detector was accomplished using a commercial eight-channel correlator board (FLEX03OEM-8CH,

Correlator.com) with a fixed integration time of 3 s per measurement. The tissue absorption and scattering coefficients, measured concurrently over the same tissue volume by FD-DOS, were used as inputs for the calculation of BFI from the intensity autocorrelation curve, averaged across all co-located detection fibers; the calculation was based on the semi-infinite homogeneous medium approximation of the diffusion correlation equation.⁵⁴ Individual measurements were rejected when the average detected photon count rate (light intensity) was $<5\text{ kHz}$, when the intensity autocorrelation curve failed to decay below 1.01, or when the fit of the intensity autocorrelation curve had greater than 10% error from the sampled curve.

For validation of DCS against invasive laser Doppler, relative CBF from DCS ($rCBF_{DCS}$, %) was computed from BFI normalized to the mean baseline BFI value.

Calculation of cerebral metabolic rate of oxygen

Cerebral metabolic rate of oxygen ($CMRO_2$) was calculated using the Fick principle⁵⁵

$$CMRO_2 = CaO_2 \cdot OEF \cdot CBF \quad (2) \leftarrow$$

where CaO_2 is the arterial blood concentration of oxygen and OEF is the cerebral OEF .

Invasive calculation of $CMRO_2$. Systemic arterial hematocrit (Hct, %), arterial oxygen saturation (SaO_2 , %) and jugular venous oxygen saturation ($SjvO_2$, %) were determined at each blood gas sampling time-point and assumed to estimate cerebral arteriole and venule oxygen content, respectively.⁵⁶ Using the piglet-specific mean corpuscular hemoglobin concentration (MCHC) of 32.2 g/mL⁵⁷ and a mammalian hemoglobin oxygen binding capacity of 1.36 mL O_2 /g Hgb,⁵⁸ OEF and CaO_2 were computed as

$$OEF = \frac{SaO_2 - SjvO_2}{SaO_2} \quad (3) \leftarrow$$

$$CaO_2 = \frac{1.36\text{mLO}_2}{1\text{gHgb}} \cdot \text{Hct}(\%) \cdot \text{MCHC} \left(\frac{\text{gHgb}}{\text{mLblood}} \right) \cdot SaO_2(\%) \quad (4) \leftarrow$$

Continuous laser Doppler measurements of relative CBF ($rCBF_{LD}$, %) were calculated with respect to the mean baseline value such that the baseline blood gas draws at the start of cooling corresponded to an $rCBF_{LD}$ of 100%. For each subsequent blood gas sample, the mean $rCBF_{LD}$ value in the 30 s preceding the time of blood gas draw was used. CaO_2 and OEF were also

normalized to baseline and combined into an invasive measure of relative $CMRO_2$ (invasive $rCMRO_2$, %)

$$Invasive\ rCMRO_2 = \frac{CaO_2}{CaO_{2,baseline}} \frac{OEF}{OEF_{baseline}} \frac{rCBF\ LD}{rCBF\ LD_{baseline}} \quad (5) \leftarrow$$

Non-invasive FD-DOS/DCS calculation of $CMRO_2$. Non-invasive $CMRO_2$ calculation utilized the baseline arterial blood gas oxygen concentration ($CaO_{2,baseline}$, %); this was assumed to remain constant. OEF was derived from FD-DOS-measured cerebral tissue oxygen saturation (StO_2 , %), baseline arterial oxygen saturation ($SaO_{2,baseline}$, %), and an assumed cerebral arteriovenous mixing fraction (γ) of 0.75⁵⁹⁻⁶¹

$$OEF = \frac{SaO_{2,baseline} - StO_2}{SaO_{2,baseline}} \quad (6) \leftarrow$$

The DCS-measured BFI was used as a surrogate for CBF. Baseline CaO_2 , OEF , and BFI, were combined into an absolute index of non-invasive $CMRO_2$ ($CMRO_{2,i}$), calculated continuously as^{18,26,62}

$$CMRO_{2,i} = CaO_{2,baseline} \ OEF \ BFI \quad (7) \leftarrow$$

For comparison with invasive quantification, a relative non-invasive $CMRO_2$ (non-invasive $rCMRO_2$, %) was also calculated for each blood gas sample. Corresponding non-invasive OEF and BFI values were calculated as the mean value measured in the 30 s prior to blood gas draw. These values were then normalized to their respective baseline blood gas values and, assuming constant CaO_2 and γ , were combined to calculate non-invasive $rCMRO_2$

$$Noninvasive\ rCMRO_2 = \frac{OEF}{OEF_{baseline}} \frac{BFI}{BFI_{baseline}} \quad (8) \leftarrow$$

Modeling $CMRO_2$ temperature-dependence

The van't Hoff equation has been widely applied in human and animal studies to describe the relationship between temperature and cerebral metabolism.^{7,63} Here, we also employ the van't Hoff equation, either reformulated as the empirical Arrhenius equation^{64,65} (equation (9)) or as the Q_{10} temperature coefficient (equation (11)), to quantify the temperature-dependence of cerebral oxygen metabolism.

Arrhenius equation approach. In the Arrhenius relationship (equation (9)), a rate of reaction (k) depends on

temperature (T), the universal gas constant (R), an activation free energy barrier (E_a), and a pre-exponential factor (A) which is related to the reaction attempt frequency

$$k = Ae^{-\frac{E_a}{RT}} \quad (9) \leftarrow$$

Here we assume this rate of reaction (k) to be the metabolic rate, $CMRO_2$. Rearrangement and substitution yield a linear expression ($y = ax + b$) that models the relationship between $CMRO_2$ and temperature

$$\ln(CMRO_2) = \frac{E_a}{R} \left(\frac{1}{T} \right) + \ln(A) \quad (10) \leftarrow$$

Model parameters, $a = \frac{E_a}{R}$, $b = \ln(A)$, are obtained from data using linear regression.

Assessment of model accuracy. Arrhenius-type approaches, such as the version we utilize, represent an oversimplification of cerebral metabolism, which depends on many chemical reactions and other factors.⁷ The model selection we have made might be valid, for example, if a single rate-limiting reaction exists for the metabolic process, or if multiple important reactions had roughly the same free energy barrier height. Dense temperature sampling in vitro has shown good agreement⁶⁶; however, multiple reports of in vivo characterization using sparse sampling methods have found non-Arrhenius or multiphasic behavior, depending on temperature range.^{10,67,68} Thus, using continuous quantification of non-invasive $CMRO_{2,i}$ and temperature, we evaluated model robustness in vivo by goodness-of-fit of the linear regression (equation (10)).

Temperature coefficient, Q_{10} . Most commonly, the temperature-dependence of metabolism has been assessed using the *temperature coefficient*, Q_{10} . This metric is a reduction of the van't Hoff equation for use under physiologic conditions (see Supplementary Materials, Section S.2). Q_{10} is defined as the relative change in cerebral metabolic rate per 10°C change in temperature. It can be calculated using an initial and subsequent measurement of temperature and $CMRO_2$

$$Q_{10} = \left(\frac{CMRO_{2,1}}{CMRO_{2,2}} \right)^{\frac{10}{T_1 - T_2}} \quad (11) \leftarrow$$

The Q_{10} for both invasive and non-invasive $rCMRO_2$ measurements was quantified with respect to nasopharyngeal and ICTs.

Statistical analysis

All statistical analyses were carried out using MATLAB 2014a. Summary statistics were reported as mean and standard deviation, unless otherwise noted.

Continuous time-series data were synchronized using 15 s epoch averages.

The sufficiency of NPT guidance was assessed via the relationship between *non-invasive* NPT and *invasive* ICT, and especially via the functional relationship between non-invasive $CMRO_{2,i}$ and each temperature source during cooling and rewarming (i.e. using the linear form of the Arrhenius equation; equation (10)). These relationships were individually examined using linear mixed-effects models that incorporated subject-specific random intercept and slope effects to allow for variation in the intercept and slope among individuals. To quantify the potential hysteresis between cooling and rewarming periods, each model included period-specific (e.g. cooling, rewarming) fixed slope (**a**, **a**) and intercept (**b**, **b**) effects reported as mean and standard error. The modeled relationship during *cooling* is expressed as

$$y = ax + b \quad (12)$$

and the modeled relationship during *rewarming* expressed as

$$y = (a + \leftarrow a)x + (b + \leftarrow b) \quad (13)$$

Parameters Δa and Δb should be interpreted as the incremental effects of rewarming on the slope (**a**) and intercept (**b**), respectively, of cooling. The goodness-of-fit of these models was evaluated using the coefficient of determination (R^2) of a separate generalized linear regression model with slope and intercept interaction terms for each subject and period.

Validation of non-invasive against invasive $CMRO_2$ measures at discontinuous blood gas sampling time-points was conducted by paired *t*-test, assuming equal variances and evaluated at a pooled significance level of $\alpha = 0.05$ with Bonferroni correction for multiple comparisons. Consequently, the five individual time-point *t*-tests were evaluated at an adjusted significance level of $\alpha = 0.01$. Given a type II error rate of $\beta = 0.2$ and an assumed within-subject correlation of 0.875, the analysis was powered to detect a 10% difference in paired observations. Secondly, to assess the *continuous* relationship between *invasive* $rCBF$ LD versus *non-invasive* $rCBF$ DCS and *invasive* $rCMRO_2$ versus *non-invasive* $rCMRO_2$, linear mixed-effects models with random slope effects were used to quantify the slope relating the change from baseline ($rCBF$, %; $rCMRO_2$, %; respectively) between modalities; slope is reported as mean and standard error. To balance continuous $rCBF$ data across the full range of temperatures from normothermia to DH, the synchronized time-series data were bin-averaged by corresponding temperature in 1°C intervals from 18°C to 37°C. Linearity was

evaluated by the goodness-of-fit, as described above. Finally, agreement between non-invasive measures versus invasive measures of $rCBF$ and $rCMRO_2$ was evaluated by the bias and precision from repeated-measures Bland–Altman analysis.^{69,70}

Calculated $CMRO_2 Q_{10}$ coefficients for both nasopharyngeal and ICT-dependence were compared between modalities and to values reported in the literature. Due to their non-normal sample distributions, these results were reported as median and interquartile range (IQR), and intra-subject Q_{10} comparisons were made using the Wilcoxon signed-rank test.

Results

Neonatal swine ($n = 8$), with a mean (SD) weight of 4.1 (0.5) kg, were cooled to DH in 25.7 (5.3) min, maintained at DH for 42.6 (1.1) min, and subsequently rewarmed to normothermia in 27.2 (7.0) min.

Summary statistics of experimental parameters are listed in Table 1.

Temperature-dependence of cerebral oxygen metabolism: Comparison of nasopharyngeal and ICT

Here we examine the impact of NPT guidance on cerebral metabolic parameters. An example of non-invasive optical time-series data is available in Supplementary Materials, Section S.3. The mean and standard deviation of the non-invasive optical measurements of OEf , $rCBF$ DCS, and $rCMRO_2$ with respect to NPT during cooling and rewarming periods is plotted in Figure 3. Cooling to DH caused a decrease in all parameters. During rewarming, OEf values are in agreement with cooling, but a hysteresis in $rCBF$ and $rCMRO_2$ is apparent.

Characterization of the relationship of NPT and ICT provided additional insights regarding the hysteresis (Figure 4, left). A significant slope effect ($a = 0.63$ [0.06], $p < 0.001$) confirmed an association between the two temperature monitors with the effect size < 1 indicating a lag of ICT behind NPT (Figure 4, left). Significant rewarming slope ($\Delta a = -0.08$ [0.01], $p < 0.001$) and intercept ($\Delta b = -5.0$ [0.3], $p < 0.001$) effects demonstrated a hysteresis between the two temperature monitors with an increased lag and offset in ICT. The mismatch in ICT between the end of cooling and the beginning of rewarming indicated that, despite NPT indicating attainment of DH, the brain had not reached thermal equilibrium.

The NPT-dependence of non-invasive $CMRO_{2,i}$ exhibited a significant slope effect ($a = -3.7$ [0.4] 10^3 , $p < 0.001$) which verified an association between metabolism and temperature (Figure 4, center). Confirming the observed hysteresis, rewarming

Table 1. Summary statistics.

	Start of cooling	End of cooling	Start of rewarming	End of rewarming
Arterial blood gas^a				
pH	7.4 (0.1)	7.1 (0.1)	7.1 (0.1)	7.4 (0.1)
pCO ₂ (mmHg)	45.8 (17.2)	86.5 (16.9)	83.3 (20.5)	39.8 (8.4)
pO ₂ (mmHg)	278.6 (92.7)	327.3 (58.5)	286.6 (34.8)	181.7 (80.5)
Glu (umol/L)	163.9 (56.1)	156.1 (45.7)	162.9 (37.7)	142.1 (36.7)
Lac (mmol/L)	3.5 (1.2)	3.8 (1.2)	4.1 (1.4)	6.0 (4.9)
Hct (%)	29.9 (2.8)	29.3 (6.4)	33.2 (7.5)	35.6 (7.0)
HCO ₃ (mmol/L)	29.5 (3.1)	29.6 (3.2)	27.2 (5.0)	24.8 (2.9)
SaO ₂ (%)	99.2 (2.5)	100.0 (0.0)	99.9 (0.2)	98.8 (2.3)
Invasive neuromonitoring				
ICT (°C)	34.5 (2.4)	22.6 (2.1)	18.8 (1.1)	30.2 (3.1)
PbtO ₂ (mmHg)	8.0 (2.7)	8.6 (4.9)	11.1 (8.7)	6.2 (4.9)
SjvO ₂ (%) ^a	76.6 (13.2)	95.3 (5.3)	97.3 (2.2)	84.2 (7.3)
OEF	0.23 (0.13)	0.047 (0.053)	0.026 (0.022)	0.15 (0.06)
rCBF LD (%)	100.0 (-)	45.9 (18.1)	41.9 (21.2)	49.4 (9.9)
rCMRO ₂ (%)	100.0 (-)	9.3 (9.9)	3.4 (3.2)	32.3 (7.9)
Non-invasive neuromonitoring				
NPT (°C)	37.3 (0.5)	17.8 (0.3)	18.2 (0.5)	37.3 (0.5)
μ_a (1/cm)				
$\lambda = 690$ nm	0.17 (0.02)	0.14 (0.02)	0.15 (0.02)	0.19 (0.03)
$\lambda = 725$ nm	0.14 (0.02)	0.13 (0.02)	0.14 (0.01)	0.16 (0.02)
$\lambda = 785$ nm	0.14 (0.02)	0.15 (0.03)	0.15 (0.03)	0.16 (0.03)
$\lambda = 830$ nm	0.15 (0.02)	0.16 (0.03)	0.17 (0.03)	0.17 (0.02)
μ_s' (1/cm)				
$\lambda = 690$ nm	12.2 (1.7)	11.6 (1.6)	11.7 (1.3)	12.4 (1.6)
$\lambda = 725$ nm	10.2 (1.5)	10.0 (1.4)	10.1 (1.2)	10.6 (1.6)
$\lambda = 785$ nm	9.9 (1.5)	10.0 (1.5)	10.0 (1.4)	10.3 (1.7)
$\lambda = 830$ nm	9.4 (2.0)	10.1 (2.1)	9.9 (1.8)	9.9 (1.7)
THC ($\mu\text{mol/L}$)	75.3 (9.3)	81.0 (14.0)	85.1 (14.9)	86.1 (12.6)
StO ₂ (%)	57.2 (5.3)	68.6 (8.7)	69.9 (11.4)	58.6 (6.7)
OEF	0.56 (0.07)	0.41 (0.10)	0.39 (0.14)	0.54 (0.08)
BFI (10^{-8} cm ² /s)	1.2 (1.2)	0.57 (0.55)	0.49 (0.51)	0.69 (0.73)
rCBF DCS (%)	100.0 (-)	50.1 (17.5)	45.9 (28.9)	64.5 (31.6)
rCMRO ₂ (%)	100.0 (-)	35.8 (8.7)	27.8 (11.4)	63.2 (35.5)

^aValues are corrected to 37°C. Glu: Glucose; Lac: Lactate; Hct: Hematocrit; SjvO₂: internal jugular venous oxygen saturation; rCBF LD: relative cerebral blood flow measured using laser Doppler; ICT: intracranial temperature; PbtO₂: partial pressure of oxygen in brain tissue; StO₂: cerebral tissue oxygen saturation from FD-DOS; BFI: blood flow index measured using DCS; rCBF DCS: relative cerebral blood flow measured using DCS; NPT: nasopharyngeal temperature; THC: total hemoglobin concentration measured using FD-DOS.

had a significant and dampening effect on slope ($\mathbf{a} = +1.4 [0.4] 10^3$, $p = 0.001$), which suggests metabolism had diminished sensitivity to NPT. Linear regression resulted in a strong coefficient of determination ($R^2 = 0.87$) and affirms use of Arrhenius-type models for examining cerebral metabolic temperature-dependence at physiologic temperatures. The ICT-dependence of non-invasive $CMRO_{2,i}$ also demonstrated a strong coefficient of determination with linear regression ($R^2 = 0.90$) and had a significant slope effect

($\mathbf{a} = -5.2 [1.0] 10^3$, $p < 0.001$; Figure 4, right). Surprisingly, rewarming did not have a significant effect on slope ($\mathbf{a} = +0.3 [0.6] 10^3$, $p = 0.647$) or intercept ($\mathbf{b} = -1.1 [2.1]$, $p = 0.585$).

These results highlight critical insufficiencies of NPT guidance to accurately reflect ICT or metabolic status during DH CPB. Furthermore, they suggest that the presence of cerebral metabolic hysteresis with respect to temperature may be an artifact resulting from use of NPT to approximate ICT.

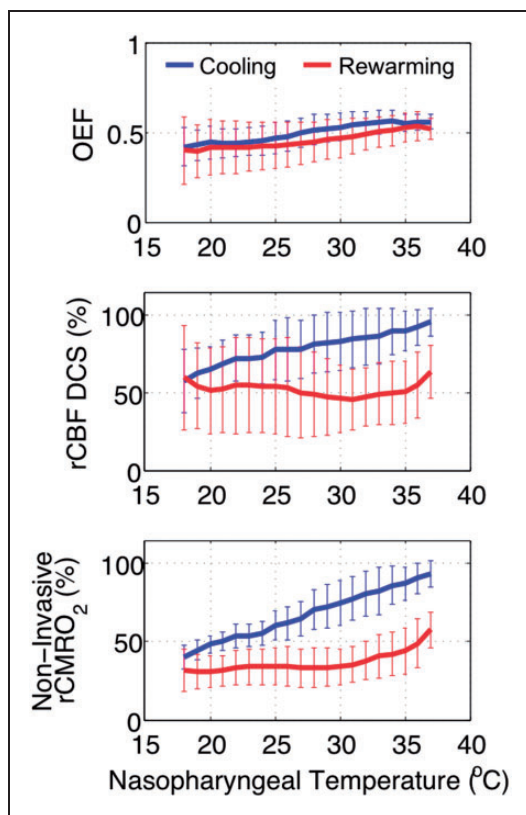


Figure 3. Temperature-dependence of non-invasive optical measures of cerebral oxygen extraction fraction, flow, and metabolism during cooling (blue) from normothermia to deep hypothermia, and during rewarming (red) from deep hypothermia back to normothermia. Mean values across subjects ($n=8$) are plotted along with standard deviation. Abbreviations: *OEF*: oxygen extraction fraction; *rCBF DCS*: DCS measure of relative cerebral blood flow; *rCMRO₂*: relative cerebral metabolic rate of oxygen.

Validation of non-invasive quantification of cerebral oxygen metabolism

The results of models examining discontinuous and continuous repeated measures of *CMRO₂* parameters to assess differences between invasive and non-invasive modalities are reported. Blood gas analysis was hampered by a machine malfunction in a single animal, resulting in the inclusion of seven of eight animals.

Arterial concentration of oxygen (*CaO₂*). *CaO₂* quantified discontinuously from invasive blood gas samples, were used to examine the non-invasive assumption that *CaO₂* remained constant through DH and recovery to normothermia. Importantly, no significant differences from baseline were observed in relative *CaO₂* (*rCaO₂*; Figure 5, top left). From these results, we conclude that the assumption of constant *CaO₂* for non-invasive quantification during DH was reasonable.

***OEF*.** Significant differences between non-invasive versus discontinuous invasive measures of relative *OEF* (*rOEF*; Figure 5, top right) were found at the end of cooling (difference = +53.9% (24.3), $p=0.001$), start of rewarming (difference = +62.4% (22.6), $p<0.001$), midpoint of rewarming (difference = +50.8% (20.4), $p=0.002$), and end of rewarming (difference = +34.2% (17.2), $p=0.005$). Non-invasive sampling demonstrated consistently greater *rOEF* across all time-points.

The non-invasive calculation of *OEF* is derived from baseline blood gas *SaO₂* and from continuous non-invasive measurement of cerebral tissue oxygen saturation (*StO₂*; equation (6)). Given the limited range in *SaO₂* (98.0–100.0%) during cooling and rewarming, significant differences in non-invasive and invasive *OEF* must be attributed to a disagreement between non-invasive *StO₂* and invasive sampling of jugular venous oxygen saturation (*SjvO₂*). Additional analysis of the relationship of *StO₂* and *SjvO₂* is included in Supplementary Materials, Section S.4.

***CBF*.** Non-invasive, continuous *rCBF DCS* measurement demonstrated good agreement with invasive, continuous *rCBF LD*. Significant differences were not found at all discontinuous blood gas sampling time-points (Figure 5, bottom left). Examination of the *continuous* relationship between modalities demonstrated that the change from baseline (*rCBF*, %) of non-invasive DCS significantly predicted (Figure 6, left) invasive laser Doppler with a slope effect of 1.26 [0.15] ($p<0.001$). Linear regression with subject-specific slope interactions resulted in a good coefficient of determination ($R^2=0.73$), suggesting a strong linear relationship between modalities. Using Bland–Altman analysis, the comparison of mean *rCBF* (%) between modalities was found to have a bias of –10.0% and precision of 13.1% (Supplementary Materials, Section S.5). These findings support the use of DCS for non-invasive measurement of *rCBF* during DH CPB.

***CMRO₂*.** Non-invasive versus invasive relative *CMRO₂* measured discontinuously exhibited significant differences at the end of cooling (difference = +26.9% (9.7), $p<0.001$), start of rewarming (difference = +25.1% (11.7), $p=0.001$), and midpoint of rewarming (difference = +18.3% (8.1), $p=0.003$; Figure 5, bottom right). Due to the dependence of *CMRO₂* on *OEF*, significant differences in *rOEF* directly contributed to differences in *rCMRO₂*, whereby non-invasive sampling reflected higher levels of metabolism versus invasive sampling.

Examination of the continuous relationship between modalities by linear mixed-effects model analysis

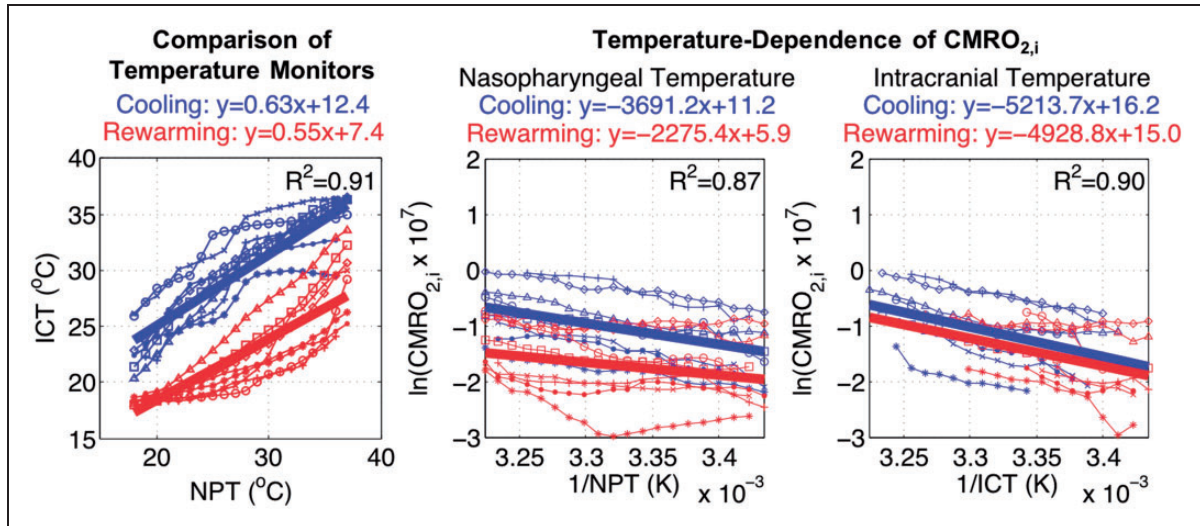


Figure 4. (Left) Intracranial temperature hysteresis: significant hysteresis of intracranial temperature (ICT) with respect to nasopharyngeal temperature (NPT) during rewarming (red) versus cooling (blue); rewarming slope $p < 0.001$, intercept $p < 0.001$. Non-invasive $CMRO_{2,i}$ Temperature-dependence: (Center) $CMRO_{2,i}$, with respect to $(NPT)^{-1}$, also demonstrates significant hysteresis in rewarming slope ($p = 0.001$) and intercept ($p < 0.001$). (Right) $CMRO_{2,i}$, with respect to $(ICT)^{-1}$, has improved concordance between rewarming versus cooling; rewarming slope $p = 0.647$, intercept $p = 0.585$. Thick lines represent fitted linear mixed-effects models; unique symbols connected by thin lines represent individual subject data ($n = 8$).

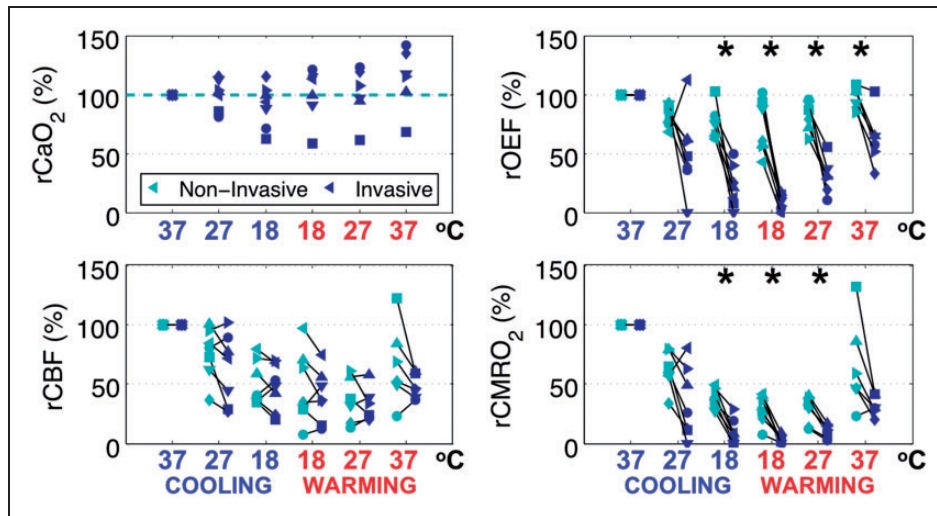


Figure 5. Comparison of invasive versus non-invasive tissue sampling at each blood gas time-point demonstrate validity of optical assumptions of arterial concentration of oxygen (CaO_2), agreement of relative cerebral blood flow ($rCBF$) measurements, but a discrepancy in relative oxygen extraction fraction ($rOEF$) and relative metabolic rate of oxygen ($rCMRO_2$). Paired data at each time-point are displayed with a unique symbol for each subject ($n = 7$) colored by modality (i.e. non-invasive, green; invasive, blue), and the x-axis labeled by respective nasopharyngeal temperature with stage designated by color (i.e. cooling, blue; rewarming, red). Asterisks (*) denote significant differences ($p < 0.01$) in paired t -tests between invasive and non-invasive sampling.

demonstrated that non-invasive $rCMRO_2$ (%) significantly predicted (Figure 6, right) invasive $rCMRO_2$ (%) with a slope effect of 1.31 [0.07] ($p < 0.001$); however, linear regression resulted in only a fair coefficient of determination ($R^2 = 0.53$) suggesting underlying non-linearity. Using Bland-Altman analysis, agreement

of $rCMRO_2$ between modalities was found to have a bias of -25.8% and precision of 12.5% (Supplementary Materials, Section S.4). Despite modest quantitative agreement, these findings show a highly significant association between non-invasive and invasive $rCMRO_2$ measurements; this association, in

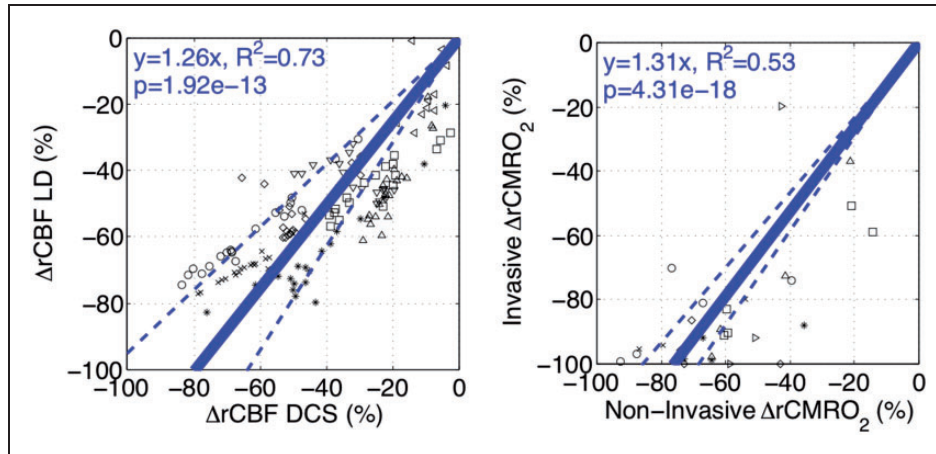


Figure 6. Validation of non-invasive diffuse correlation spectroscopy (DCS, left) and $CMRO_2$ (right) – data from individual subjects are indicated by a unique symbol. Measurements of change in relative cerebral blood flow using laser Doppler ($rCBF_{LD}$, %) and DCS ($rCBF_{DCS}$, %) are compared using a linear mixed-effects model ($n = 8$; left). DCS measurements demonstrate a good linear correlation (fixed slope effect $p < 0.001$; $R^2 = 0.73$) against laser Doppler measurements. Similarly, invasive and non-invasive measurements of $rCMRO_2$ (%) are compared using a linear mixed-effects model ($n = 7$; right). Non-invasive $rCMRO_2$ quantification demonstrates a significant association but limited linearity (fixed slope effect $p < 0.001$; $R^2 = 0.53$) against invasive sampling. Fitted linear relationships (solid line) with 95% confidence intervals (dotted line) are plotted in blue.

turn, supports use of non-invasive measurements as an indicator of cerebral status.

$CMRO_2$ temperature coefficient (Q_{10}). Computed $CMRO_2$ temperature coefficient Q_{10} derived from invasive sampling differed significantly from that of non-invasive sampling, whether assessed with respect to NPT (non-invasive versus invasive median [IQR]: 2.0 [1.6, 2.3] versus 4.9 [3.6, 6.3], $p = 0.016$) or ICT (non-invasive versus invasive median [IQR]: 2.5 [2.0, 3.5] versus 9.0 [6.9, 11.0], $p = 0.016$). This finding further affirms our discrepant findings at individual blood gas sampling time-points. As may be expected from the presence of significant temperature hysteresis, Q_{10} based on ICT was significantly different from Q_{10} based on NPT, i.e. whether measured non-invasively (NPT versus ICT median [IQR]: 2.0 [1.6, 2.3] versus 2.5 [2.0, 3.5], $p = 0.016$) or invasively (NPT versus ICT median [IQR]: 4.9 [3.6, 6.3] versus 9.0 [6.9, 11.0], $p = 0.016$). Regardless of metabolic sampling method, sensitivity to NPT was lower than to ICT.

Discussion

Continuous, non-invasive optical metabolic neuromonitoring using FD-DOS combined with DCS permits understanding of the physiologic alterations of cerebral metabolism that occur during therapeutic hypothermia. This approach has the potential to address critical shortfalls in conventional temperature guidance of hypothermia, as well as to enable individualized

neuroprotective strategies. The present study takes important steps towards this goal.

Temperature-dependence of cerebral oxygen metabolism: Comparison of nasopharyngeal and ICT

NPT is an established source of guidance for DH CPB management¹ and has been found, among other non-invasive sites of core temperature measurement, to best approximate cerebral temperature.⁴¹ However, our data add to mounting evidence that NPT does not adequately reflect ICT, nor the metabolic state of the brain.^{71,72} This finding is evident from the metabolic hysteresis with respect to NPT between cooling and rewarming periods, and in the significantly different temperature coefficients exhibited by nasopharyngeal versus ICT. The finding that ICT has more concordant metabolic temperature-dependence between cooling and rewarming periods is supported by prior in vitro observations of the reversibility of hypothermic metabolic inhibition.⁶⁶ The continued decline of ICT following cooling indicates the clinical cooling interval used was insufficient for thermal equilibrium and underscores the importance of directly measuring the metabolic state of the brain in lieu of using NPT as a surrogate.

By coupling continuous FD-DOS/DCS measurements of cerebral oxygen metabolism with continuous temperature measurements during DH, we were additionally able to examine the validity of temperature-dependent models for the metabolic rate based on the

Arrhenius equation. These models were consistent with our data, whether using NPT ($R^2=0.87$) or ICT ($R^2=0.90$). Our results support the continued use of Q_{10} to characterize the temperature-dependence of cerebral oxygen metabolism.

Validation of non-invasive quantification of cerebral oxygen metabolism

Non-invasive $rCMRO_2$ significantly predicted invasive $rCMRO_2$ ($p < 0.001$). However, we found significant discrepancies in measured values with a bias of -25.8% and precision of 12.5% . This mismatch is discussed, and rationalized, with respect to each component of the $CMRO_2$ calculation below.

Arterial concentration of oxygen. A critical assumption of non-invasive $CMRO_2$ quantification is that CaO_2 remains constant from baseline. While increases in CaO_2 are expected during hypothermia,⁷³ avoidance of hyperoxia during CPB has been established.⁷⁴ Fortuitously, oxygen administration is intentionally adjusted during hypothermia to maintain constant arterial blood oxygen tension. In-line with these recommendations, we found that only the last sampling time-point after recovery to normothermia demonstrated a significant difference from baseline. Thus, the assumption of constant CaO_2 for non-invasive quantification during deep hypothermic CPB seems reasonable and is concordant with invasive sampling.

OEF. Absolute OEF was found to differ significantly between invasive and non-invasive sampling methods at all time-points. We determined that this effect resulted from a disagreement between optically measured cerebral tissue oxygen saturation (StO_2), and the compartment model computation of StO_2 via $StO_2 = (1-\gamma)SaO_2 + \gamma SjvO_2$, wherein an arteriovenous mixing fraction of $\gamma = 0.75$ was assumed.^{59,61} In fact, StO_2 was consistently lower than $SjvO_2$, thus violating a necessarily positive γ . This phenomenon has been reported in the context of CW NIRS instruments and FD-DOS measurements in animals models^{47,75} and human subjects,^{16,60} and remains to be further explored in future studies. Our findings support the reproducibility of this phenomenon and sheds further light on the issue.

The use of jugular venous sampling in this study was based on its wide utilization in pediatric cardiac surgery for this purpose^{8,76} and for comparison with non-invasive cerebral oximetry.^{60,77-79} Prior reports of jugular venous sampling in swine models of CPB agree with our observations. In Sasaki et al.⁷⁵ $SjvO_2$ were calculated from reported OEF as $74.5 \pm 7.1\%$ at normothermia and $89.5 \pm 4.2\%$ at 18°C in comparably aged

neonatal piglets.⁷⁵ In slightly older (three to four weeks) and larger piglets ($9.4 \pm 0.8\text{kg}$), Walther et al.³⁶ reported $SjvO_2$ of $86\text{--}88\%$ at normothermia, 91% at a body temperature of 25°C , and 93% at a body temperature of 18°C .³⁶ While these values are in the range of our $SjvO_2$ measurements at normothermia ($76.6 \pm 13.2\%$) and at DH ($95.3 \pm 5.3\%$), further comparison with sagittal sinus sampling suggests these measurements may have been affected by reported limitations of jugular venous sampling to access cerebral venous saturation.⁸⁰

Animal studies wherein sagittal sinus oxygen saturation ($SssO_2$) was directly sampled during hypothermic CPB show consistently lower values than reported $SjvO_2$. Two observations in moderately larger piglets ($5\text{--}13\text{kg}$) reported an $SssO_2$ of $75 \pm 10\%$ at normothermia and $85 \pm 5\%$ at DH ($18\text{--}20^\circ\text{C}$).^{12,35} While normothermic values are comparable to our observations, hypothermic saturations are markedly lower. These comparisons indicate that internal jugular venous sampling overestimates sagittal sinus saturations, resulting in an underestimation of true cerebral OEF in pediatric swine models. We believe this to be the primary source of error in our invasively quantified OEF and, subsequently, $CMRO_2$.

Probable physiologic mechanisms that could elevate $SjvO_2$ with respect to $SssO_2$ include systemic venous contamination of jugular venous sampling due, in part, to the logistical difficulty of advancing a catheter into small neonatal vessels. Contributions from the external jugular vein or superior vena cava would reflect higher saturations due to lower somatic oxygen utilization rates compared to the brain. This hypothesis is corroborated by central venous saturations of 75% at normothermia and $98 \pm 2\%$ at 18°C in Cavus et al.,³⁵ our $SjvO_2$ agrees at baseline and is only 3% lower at DH. Future studies conducted in neonatal swine should be wary of systemic contributions with this sampling method that may inaccurately diminish cerebral OEF.

CBF. Significant agreement was observed between invasive laser Doppler (LD) and non-invasive DCS measurement of CBF throughout DH. Our invasive data, in particular, advance recent cross-validation of DCS with TCD.²⁹ In principle, quantitative variation between LD and DCS can be attributed to regional variability in metabolic and cerebrovascular response to hypothermia, which has been previously reported,⁸¹⁻⁸³ and can also result from extracerebral contributions to the optical signal.⁸⁴⁻⁸⁶ Measuring tissue thickness post-mortem, we found 0.5cm of superficial tissue (e.g. skull, scalp) above the brain. The potential contribution from this tissue should be explored in future studies using advanced multi-layered optical extraction.

$CMRO_2$. Taken in total, non-invasive optical measurement of $CMRO_2$ demonstrated lower temperature-sensitivity and higher residual metabolic rates at DH than invasively sampled $CMRO_2$. As discussed above, we believe this to be a direct result of systemic contributions to invasive jugular venous sampling. For further validation, calculated Q_{10} temperature coefficients were compared to those reported in the literature.

In studies utilizing non-invasive temperature methods and jugular venous sampling, Greeley et al.⁸ reported an average Q_{10} of 3.65 in neonates and children, and McCullough et al.⁹ reported a Q_{10} of 2.3 in adults. While these measurements are potentially confounded by pathologic conditions necessitating the use of hypothermic CPB, plausibility is provided for our non-invasive ($Q_{10}=2.0$) versus invasive ($Q_{10}=4.9$) measurements with respect to NPT.

In healthy animal models with invasive ICT and sagittal sinus sampling, Michenfelder and Milde¹⁰ saw a Q_{10} of 2.23 for mild hypothermia (ICT between 37°C and 27°C) and 4.53 for DH (27 to 14°C) in canines; CBF was measured using a flow-through electromagnetic flow probe placed in the sagittal sinus.¹⁰ Using both radioactive and fluorescent microspheres for CBF determination, Ehrlich et al.¹² observed a Q_{10} of 2.46 with a 95% CI of 2.1 to 2.9 (38°C to 8°C) in piglets ranging in weight from 7 to 13 kg.¹² These values support the validity of our non-invasive metabolic measurements, which exhibited a Q_{10} of 2.5 with respect to ICT, versus our invasive measurements ($Q_{10}=9.0$) which incorporated jugular venous sampling. In sum, despite significant differences between our non-invasive and invasive sampling methods, we are confident that our non-invasive metabolic measurements are in agreement with prior studies of cerebral metabolic temperature-response and hold tremendous clinical promise to measure an individual patient's $CMRO_2$.

Limitations of animal model

Several considerations affect the interpretation of our animal model results for application to neonatal cardiac patients. First, we utilized only female animals; in light of reported sexual dimorphisms with respect to brain development⁸⁷ and tolerance to neurological injury,^{88,89} further study is required to understand potential sex differences in metabolic temperature-dependence. Additionally, piglet resting core temperatures are slightly higher than human neonates (38.5°C vs. 36.5°C).^{82,90} The impact of relative hypothermia in the animal model may have resulted in lower metabolic values than an animal at natural baseline. Variation in anesthetic management, rate and duration of temperature derangement, and the use of circulatory arrest among pediatric cardiac surgical practices may also

impact generalizability.^{32,91,92} Notably, the oxygen binding affinity of swine hemoglobin has been shown to be significantly lower than that of human hemoglobin.⁹³ This effect could account for the lower baseline cerebral oxygen tissue saturations measured versus baseline values in human subjects. Furthermore, there is less impact of cooling on oxygen binding affinity.⁹⁴ Therefore, greater changes in cerebral metabolism (i.e. larger temperature coefficients) may be observed in humans than those measured here.

Clinical applications of cerebral metabolic monitoring during therapeutic hypothermia

Clinical imaging modalities that permit access to cerebral metabolism include stand-alone PET, which typically requires the injection or inhalation of radioactive tracers such as ^{15}O -H₂O and ^{18}F -FDG for glucose metabolism⁹⁵ or $^{15}O_2$ gas for oxygen metabolism,⁹⁶ and stand-alone MRI,⁹⁷ which uses a combination of arterial spin labeling or phase-contrast mapping for CBF and calibrated blood-oxygen-level-dependent T2* signal mapping for oxygenation. More recently, integrated PET/MRI paradigms that decrease the invasive vascular access requirements of stand-alone PET have been demonstrated.⁹⁸ These modalities have profoundly impacted clinical care, as well as our understanding of developmental and pathologic alterations in cerebral metabolism; however, extensive patient transport and operating room requirements, the incremental ionizing radiation exposure of PET, and the prolonged durations and limited throughput associated with MRI data acquisition prohibit real-time, intraoperative monitoring, particularly in neonates.

FD-DOS/DCS sacrifices spatial resolution and sensitivity for temporal resolution and portability that, as specifically demonstrated in the present work, permit real-time guidance during procedural hypothermia. Frequently, neonatal surgical protocols utilize DH for neuroprotection during subsequent circulatory arrest. Temperature of initiation and what duration of circulatory arrest is safe remain controversial.^{1,6,99} Precise determination of residual cerebral metabolism using diffuse optics could be used to individually guide cooling to adequate levels of cerebral metabolic suppression as well as provide a subject-specific estimate of safe arrest duration based on rate of [HbO₂] depletion in cerebral tissue. Alternatively, mild hypothermia has also demonstrated therapeutic potential to improve mortality^{100,101} and neurological¹⁰² and neurodevelopmental outcomes¹⁰³ in infants with hypoxic-ischemic encephalopathy. Non-invasive optical CW NIRS and FD-DOS/DCS have already been used at the bedside in these infants to examine alterations in cerebral autoregulation¹⁰⁴ and metabolism.¹⁰⁵ We anticipate that

our findings will enable and motivate closer examination of the magnitude and rate of hypothermia-induced cerebral metabolic suppression and neurological outcomes in these patients.

Conclusions

This study identifies critical limitations in conventional NPT guidance during deep hypothermic CPB in neonates, and it provides strong evidence for the validity and utility of non-invasive diffuse optical measurements of cerebral oxygen metabolism to address these limitations. Continuous measurements throughout cooling and rewarming enabled novel high-fidelity determination of metabolic temperature-dependence in vivo and validation of Arrhenius-type models (i.e. the van't Hoff Law and Q_{10}). The relationship between non-invasive $CMRO_2$ and NPT demonstrated a problematic hysteresis between cooling and rewarming periods. The finding that ICT-dependence improved concordance suggests that NPT inadequately reflects cerebral metabolic state and may be a significant source of uncertainty in the clinical guidance of hypothermia for brain protection. The application of non-invasive FD-DOS/DCS for direct quantification of cerebral oxygen metabolism thus offers promise for improved guidance of therapeutic hypothermia and for mitigation of neurological injury in vulnerable pediatric populations.

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Declaration of conflicting interests

The author(s) declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: Authors (in parenthesis) disclose partial ownership of active relevant patents applications. Pending: WO2013/090658A1 (AGY), PCT/US2012/069626

(AGY), PCT/US2015/017286 (AGY, DRB), PCT/US2015/017277 (AGY, DRB). Granted: US8082015B2 (AGY). No author currently receives royalties or payments from these patents.

Authors' contributions

TSK, CDM, WBB, JML, DRB, JG, RWM, JWJ, TJK, AGY, and DJL contributed to the conception and design of the study. TSK, CDM, VM, KM, TWB, ALS, JG, GDB, YL, SJ, RWM, TMR, BCS, KLS, and TJK contributed to acquisition of data. TK, CDM, WBB, VM, KMB, TWB, ALS, JML, DRB, RX, AGY, TJK, and DJL contributed to analysis and/or interpretation of data. TSK, CDM, and RWM drafted the manuscript. WBB, DRB, RX, AGY, TJK, and DJL contributed to revising the manuscript critically for important intellectual content. All authors have approved the final version of the manuscript for publication.

Supplementary material

Supplementary material for this paper can be found at the journal website: <http://journals.sagepub.com/home/jcb>

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